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Angiocrine control of tissue metabolism

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Abstract

Endothelial cells are important gatekeepers of organ homeostasis and metabolism. It has long been determined that impaired vascular function is associated with tissue dysregulation and metabolic disorders, but whether endothelial cells contribute to metabolic homeostasis beyond ensuring tissue perfusion is poorly described. Using an unbiased proteomics approach to analyze endothelial secretome, we identified GDF15 to be abundantly secreted. As GDF15 has been identified as an important modulator of metabolism through the control body weight, in this dissertation, the regulation of angiocrine GDF15 was explored and whether it influences other aspects of metabolism. It was determined that GDF15 expression is negatively controlled by FOXO1, master regulator of endothelial quiescence, but not NOTCH1, master regulator of endothelial specification. Inhibiting FOXO1 using the inhibitor AS1842856, increased GDF15 expression. *In vivo*, endothelial specific deletion of FOXO1 increased GDF15 expression (>4 fold) in endothelial cells isolated from different tissues and was associated with significant upregulation of GDF15 in urine and serum. Furthermore, endothelial specific deletion of FOXO1 provoked a significant decrease in food intake, suggesting that small changes in circulating GDF15 suffice to control food intake. Finally, it was determined that GDF15 has an acute insulin sensitizing effect, which is primary to the acute administration of GDF15 and independent of weight loss. This work identifies FOXO1 as a direct regulator of GDF15 in ECs and highlights the critical role of endothelial cells in whole body metabolic control.

Zusammenfassung

Endothelzellen sind wichtige Regulatoren der Organhomöostase und des Stoffwechsels. Es ist allgemein bekannt, dass eine gestörte Gefäßfunktion mit Gewebedysregulationen und Stoffwechselstörungen einhergeht. Jedoch ist bis heute nur unzureichend untersucht, ob und inwiefern Endothelzellen über die Sicherstellung der Gewebepерfusion hinaus zur metabolischen Homöostase beitragen. Unter Verwendung eines unvoreingenommenen Proteomik-Ansatzes zur Analyse des endothelialen Sekrets haben wir festgestellt, dass GDF15 reichlich sezerniert wird. Basierend auf der Feststellung, dass GDF15 mit Hilfe des Körpergewichts als wichtiger Regulator des Stoffwechsels fungiert, wurde in dieser Dissertation die Regulation von angiokrinem GDF15 untersucht und dessen Effekte auf den Stoffwechsel. Es wurde festgestellt, dass die GDF15-Expression durch FOXO1 (Hauptregulator der endothelialen Stilllegung) negativ reguliert wird, nicht aber durch NOTCH1 (Hauptregulator der endothelialen Spezifikation). Die Hemmung von FOXO1 mit dem Inhibitor AS1842856 erhöhte die GDF15-Expression. In vivo steigerte die endothelspezifische Deletion von FOXO1 die GDF15-Expression (>4fach) in den Endothelzellen, welche aus verschiedenen Geweben isoliert wurden. Zudem konnte eine signifikante Hochregulierung von GDF15 in Urin und Serum festgestellt werden. Darüber hinaus bewirkte die endothelspezifische Deletion von FOXO1 eine signifikante Abnahme der Nahrungsaufnahme, was darauf hindeutet, dass kleine Veränderungen im zirkulierenden GDF15 ausreichen, um die Nahrungsaufnahme zu kontrollieren. Schließlich konnte beobachtet werden, dass GDF15 einen akuten Insulin-sensibilisierenden Effekt hat, der primär auf die akute Verabreichung von GDF15 zurückzuführen ist und unabhängig vom Gewichtsverlust ist. Diese Arbeit konnte FOXO1 als einen direkten Regulator von GDF15 in Endothelzellen identifizieren und unterstreicht die entscheidende Rolle von Endothelzellen bei der Ganzkörper-Stoffwechselkontrolle.

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Figure 13 GDF15 stimulation increases insulin sensitivity and glucose handling in human myotubes. **a.** *GDF15* knockdown efficiency in E4ORF1 HUVECs using sh- α GDF15 lentiviral viral particles. (n= 3)(Student-*t* test). **b.** Insulin signaling cascade

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Figure 14 Administration of rhGDF15 promotes increased insulin sensitivity and greater glucose uptake in skeletal muscle in mice fed chow diet. Mice were administered 8 nmol of rhGDF15 at the beginning of their dark cycle. **a** There are no differences in changes in body weight after 5 days between controls (n= 6) and GDF15 treated animals (n= 6) (Two-way ANOVA with Sidak's multiple comparison). **b** GDF15 treated mice show an increased insulin signaling upon stimulated with 1 IU/kg body weight insulin for 15 min, the total load shows comparable amounts of protein loaded in the representative blot. **c. d.** Downstream insulin signaling was increased as measured by phosphorylation of AKT T308 and AS160 T642(Two-way ANOVA and Tukey's multiple comparison). **e.** rhGDF15 treatment improves insulin sensitivity as shown by ITT (vehicle, n= 4, rhGDF15, n= 3) (Two-way ANOVA and Sidak's multiple comparison) **f.** but treatment does not influence glucose handling as assessed in

ipGTT (n= 4) (Two-way ANOVA and Sidak's multiple comparison). **g.** *In vivo* insulin-stimulated radioactive 2-Deoxy glucose uptake of several tissue (Student-t test). *Ta*, tibialis anterior; *Edl*, extensor digitorum longus; *Sol*, soleus; *Rgas*, red gastrocnemius; *Wgas*, white gastrocnemius; *Rquad*, red quadriceps; *Ewat*, epididymal white adipose tissue; *Liv*, liver; *Bat*, brown adipose tissue; *Dia*, diaphragm. Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001

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Extended data figures

Extended data figure 1 Workflow of generation and quantification of secretome of HUVECs produced in serum-free conditions. **a.** Heatmap of the top 40 most upregulated proteins present in CM. Uniquely identified proteins were included in the analysis, relative abundance is shown as escalated expression compared to control. **b** GDF15 accumulates in HUVECs culture supernatants in standard culturing conditions over time (0 h, 24 and 48 h), GDF15 ELISA. (Student-*t* test) **P*<0.05, ***P*<0.01, ****P*<0.001, and *****P*<0.0001 116

Extended data figure 2 FOXO1, but not NOTCH1, regulates expression of GDF15 in E4ORF1 and WT HUVECs. **a.** Dose response secretion of GDF15, 48 hrs. after induction of FOXO1CA with 50, 150 and 300 ng/ml doxycycline in Tet-On FOXO1CA E4ORF1 HUVECs in serum free conditions (n=4) (Two-way ANOVA and Tukey's multiple comparison). **b** FOXO1 inhibition, AS1842856, increases in a dose response expression of *ATF4* but not *CHOP* (*CHOP* was upregulated with 50 nM but not 100 nM AS1842856 treatment) or *BIP* (Two-way ANOVA with Tukey's multiple comparison). **c.** Knockdown of *ATF4* in HUVECs (n=3) **d** did not blunt *GDF15* upregulation upon FOXO1 pharmacological blockade (n=3) (Student-*t* test). Significant values are represented in asterisks as follows **P*<0.05, ***P*<0.01, ****P*<0.001, and *****P*<0.0001 117

Extended data figure 3 A block of proliferation by Mitomycin C treatment does not blunt FOXO1 negative regulation on GDF15 in vitro. **a.** GDF15 secretion of HUVECs that were treated with FOXO1 inhibitor, AS1842856, 100 nM (n= 9), or an equivalent volume of vehicle, DMSO (n= 9), both in the presence of Mitomycin C 1 µg/ml. Cell culture supernatants were collected after 72 hours. (Student-*t* test) **b.** GDF15 secretion of FOXO1CA HUVECs (n= 9) or controls cells (n= 9) that were treated with Mitomycin C 1 µg/ml. Cell culture supernatants were collected after 72 hours. (Student-*t* test). Significant values are represented in asterisks as follows **P*<0.05, ***P*<0.01, ****P*<0.001, and *****P*<0.0001 118

Supplementary tables

Supplemental table 1. Full list of identified proteins in proteomic analysis. In yellow are highlighted uniquely identified proteins..... 120

Abbreviations

AEC	Alveolar epithelial cell
2DG	2-Deoxy Glucose
3PO	3-(3-Pyridinyl)-1-(4-pyridinyl)-2-propen-1-one
6PG	6-phosphogluconate
6PGD	6-phosphogluconate dehydrogenase
ACAD	FAD-dependent acyl-CoA dehydrogenase
ACTB	Actin beta
AGO	Argonaute proteins
AKT	Protein kinase B
AMP	Adenosine monophosphate
AMPK	Adenosine monophosphate kinase
ANCOVA	Analysis of covariance
ANTP	Homeobox A7
AS160	Akt substrate 160
AS1842856	FOXO1 inhibitor
ATF4	Activating transcription factor 4
ATP	Adenosine triphosphate
BAT	brown adipose tissue
BCA	Bicinchoninic acid
BIP	Binding-Immunoglobulin Protein
BPTES	Bis-2-(5-phenylacetamido-1,2,4-thiadiazol-2-yl)ethyl sulfide

BSA	Bovine serum albumin
CD31	Cluster of differentiation 31
CD36	Cluster of differentiation 36
CD45	Cluster of differentiation 45
CHOP	C/EBP homologous protein
CM	Conditioned medium
CNS	Central nervous system
CONACyT	Mexican research council
CPT1	Carnitine palmitoyltransferase 1
CT	Cycle threshold
DAMP	Damage-associated molecular patterns
DAPT	N-[N-(3,5-Difluorphenacetyl)-L-alanyl]-S-phenylglycin-tert-butylester
DDA	Data-dependent mode
DHAP	Dihydroxyacetone phosphate
DLL4	Delta ligand-like 4
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DTT	Dithiothreitol
E4ORF1	E4 open reading frame 1
EC	Endothelial cell
ECM	Extra cellular matrix
EDL	Extensor digitorum longus
EDTA	Ethylenediamine tetra acetic acid
EGCS	Endothelial cell growth factor supplements

EGF	Epidermal growth factor
EGM2	Endothelial cell growth medium 2
EGTA	Ethylene Glycol Tetra acetic Acid
ELISA	Enzyme-linked immunosorbent assay
ER	Endoplasmic reticulum
ERC	European research council
ERK	Extracellular Signal-Regulated Kinase
ETC	electron transport chain
ETH	Swiss Federal Institute of Technology
FA	Fatty acids
FABP	Fatty acid binding protein
FACS	Fluorescence-activated cell sorting
FAD	Flavin adenine dinucleotide
FADH2	Flavin adenine dinucleotide (reduced)
FAO	Fatty acid oxidation
FATP	Long-chain fatty acid transport protein
FATP3	Long-chain fatty acid transport protein 3
FATP4	Long-chain fatty acid transport protein 4
FBS	Fetal bovine serum
FDR	False discovery rate
FELASA	Federation of European laboratory animal science associations
FGF	Fibroblast growth factor
FGF21	Fibroblast growth factor 21
FGFR	Fibroblast Growth Factor Receptor

FOXO	Forkhead Box O
FOXO1A3	Forkhead Box O1A3
FOXO1 CA	FOXO1 constitutively active
FOXO3	Forkhead Box O 3
FOXO4	Forkhead Box O 4
G3P	Glyceraldehyde-3-phosphate
G6P	Glucose-6-phosphate
G6PD	Glucose-6-phosphate dehydrogenase
GDF15	Growth differentiation facto 15
GDH	Glutamate dehydrogenase
GFP	Green fluorescent protein
GFRAL	GDNF family receptor alpha like
GLS1	Glutaminase 1
GLUT1	Glucose transporter 1
GLUT4	Glucose transporter 4
GPNA	L-γ-glutamyl-p-nitroanilide
GSEA	Gene set enrichment analysis
GTT	Glucose tolerance test
HADH	L-3-hydroxyacyl-CoA dehydrogenase
HEK	Human embryonic kidney
HES1	Hairy and enhancer of split 1
HEY1	Hes related family BHLH transcription factor with YRPW motif 1
HEY2	Hes related family BHLH transcription factor with YRPW motif 2
HGF	Hepatocyte growth factor

HIF	Hypoxia Inducible Factor
HIFU	High Intensity Focused Ultrasound
HIV	Human immunodeficiency virus
HK2	Hexokinase 2
HRP	Horseradish peroxide
HUVEC	Human umbilical cord vein endothelial cells
ICAM	Intercellular Adhesion Molecule
IDH	Isocitrate dehydrogenase
IGF1	Insulin Like Growth Factor 1
IR	Insulin receptor
ISR	Integrated stress response
ITT	Insulin tolerance test
IU	International units
IV	Intravenous
αKG	alpha ketoglutarate
KLF2	Kruppel like factor 2
LC	Liquid chromatography
LPS	Lipopolysaccharides
M199	Medium 199
ME	Malic enzyme
MEM	Minimal essential medium
MMP	Matrix metalloprotein protease
MMP2	Matrix metalloprotein protease 2
MOI	Multiplicity of infection

MS	Mass spectrometry
MXI1	MAX interactor 1, dimerization protein
MZ	Mass charge
NAD	Nicotinamide adenine dinucleotide
NADPH	Nicotinamide adenine dinucleotide phosphate
NC	Non-conditioned medium
NF	Nuclear factor
NO	Nitric oxide
NOS	Nitric oxide synthase
NRARP	NOTCH regulated ankyrin repeat protein
NRP1	Neuropilin 1
NTP	Nucleoside triphosphate
OAA	Oxaloacetic acid
OGDH	α -ketoglutarate dehydrogenase
OPD	o-Phenylenediamine
PAMP	Pathogen-associated molecular patterns
PAX2	Pair boxed 2
PBS	Phosphate buffer saline
PC	Pyruvate carboxylase
PCA	Perchloric acid
PCR	Polymerase chain reaction
PDH	Pyruvate carboxylase
PEP	Phosphoenolpyruvate
PFA	Paraformaldehyde

PFKFB3	6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3
PPAR	Peroxisome Proliferator Activated Receptor
PPP	Pentose phosphate pathway
PVDF	Polyvinylidene difluoride
RCF	Relative centrifugal force
RER	Respiratory exchange ratio
RLT	Lysis Buffer
RNA	Ribonucleic acid
ROCK1	Rho Associated Coiled-Coil Containing Protein Kinase 1
ROS	Reactive oxygen species
RT	Room temperature
Ru5P	Ribulose-5-phosphate
SEM	Standard error of mean
SGK	Serum/glucocorticoid –regulated kinase
SLC1A5	Solute Carrier Family 1 Member 5
SYBR Green	N',N'-dimethyl-N-[4-[(E)-(3-methyl-1,3-benzothiazol-2-ylidene)methyl]-1-phenylquinolin-1-ium-2-yl]-N-propylpropane-1,3-diamine
TA	Tibialis anterior
TBS	Tris buffer saline
TBX3	T-box transcription factor 3
TCA	Tricarboxylic acid cycle
TGF	Transforming growth factor
TLR	Toll-like receptors

TNF	Tumor necrotic factor
TSP1	Thrombospondin 1
UTR	Untranslated region
UV	Ultraviolet light
VCAM	Vascular Cell Adhesion Molecule
VEGF	Vascular endothelial growth factor
VEGFR1	Vascular endothelial growth factor receptor
VSMC	Vascular smooth muscle cells
VWF	Von Willebrand factor
WAT	white adipose tissue
WNT	Wingless-Type MMTV Integration Site Family
WT	Wild type

1. Introduction

The vascular endothelium is the organ system in charge of transporting oxygen and nutrients from the sites they are taken up towards the peripheral tissues, while at the same time, carrying by-products of metabolism for their disposal. Over the last decades, it has however become clear that the role of this widespread organ system goes far beyond, and that the endothelium plays an active role in the regulation of tissue homeostasis.

In this chapter, a short overview of vascular physiology will be given as an example of how the vascular endothelium regulates several important aspects of organ homeostasis. Furthermore, emphasis will be made on the emerging properties of vascular biology as a secretory organ and as a regulator of metabolic homeostasis, and how these two properties could be controlled by master regulators of endothelial cell specification.

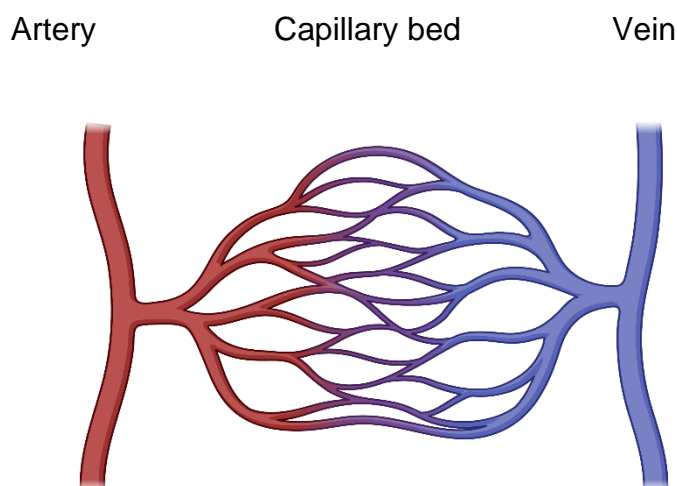


Figure 1 . Schematic representation of vascular tree. Three distinctive functional and morphological areas: arteries, veins, and capillaries. Arteries and veins are thick vessels composed of endothelial cells, perivascular cells, smooth muscle cells and elastic collagen fibers. On the other hand, capillaries are intricate vessel meshwork only formed by a single cell layer. This creates a large surface area through which it communicates with its surrounding tissue, and vice versa, for homeostatic communication and regulation.

1.1. Regulation of organ homeostasis through angiocrine signaling

1.1.1. Endothelial modulation of immune response.

The immunoregulatory role of endothelial cells is broad and fundamental to ensue a healthy immunogenic response (Jambusaria, Hong et al. 2020) and, ultimately, to resolve it (Kadl and Leitinger 2005). Upon inflammatory or necrotic stimulation, endothelial cells become active, they increase their leakiness, adhesiveness of leukocytes amplifies, platelet adhesion commences, and angiogenesis is triggered (Pober and Sessa 2007). Furthermore, endothelial cells participate in the innate immunity by reacting to pathogen-associated molecular patterns (PAMPs) (Pober and Sessa 2007) and damage-associated molecular patterns (DAMPs) (Vénéreau, Ceriotti et al. 2015). These molecular signatures interact with Toll-like receptors (TLRs) that trigger signals which lead to pro inflammatory cytokine expression, leukocyte recruitment, phagocytosis, and cytotoxicity (Maaser, Heidemann et al. 2006). However, during endothelial immunomodulatory events, endothelial cells are not only a recipient of inflammatory signals, but they also play a central role by directing the immune response by signaling immune cells contained in their milieu. This has been recently shown to occur through the secretion of metabolites. Zhang *et al*/demonstrated that endothelial-derived lactate drives macrophage polarization to a regenerative M2-like phenotype, which ultimately is responsible of driving skeletal muscle regeneration in a hind limb ischemia model. (Zhang, Muri et al. 2020). Overall, it suggests that secretory capacity of endothelial cells cement their role in modulation of immune response and tissue regeneration

1.1.2. Endothelial modulation of vascular tone via nitric oxide production.

Blood vessels have the property of regulating blood pressure through the production of nitric oxide. Particularly, endothelial cells have a strong influence on vascular tone by releasing a battery of vasodilators like NO, prostacyclin and bradykinin, and vasoconstrictors such as endothelin and endothelium/derived constrictor factor (Andresen 2006).). Hypertension is the process in which the heart pumps to a greater resistance in peripheral tissues. Structural and functional modifications in arteries are the underlying mechanism to these changes (Andresen 2006). To counteract these effects, endothelial-derived vasoactive molecules take part in coordinating the

vasodilation of arterioles and in focalized adjustments of flow and shear stress in response to mechanical queues (Iadecola 2004). For instance, EC release nitric oxide to reestablish adequate vascular tone via signaling to vascular smooth muscle cells (VSMCs). Increased laminar shear stress stimulates production of nitric oxide by the enhanced expression of endothelial nitric oxide synthase (eNOS) and through increased activity via Kruppel-like factor 2 (Klf-2) and NF-kB. (Davis, Grumbach et al. 2004, Grumbach, Chen et al. 2005).

1.1.3. Angiocrine factors important in tissue regeneration.

It has been described that endothelial cells possess the capacity to regenerate their surrounding tissue upon injury. Beyond their known function to restore oxygen and nutrient supply through the formation of new blood vessels (angiogenesis), they have the capacity to mediate tissue regeneration and homeostasis through the secretion of angiocrine factors. These factors amongst others contribute to sustaining the clonal expansion of progenitor cells and to maintain an adequate vascular niche permissive for resident stem cells to return to their quiescent state, once they have been activated, and thus preventing stem cell exhaustion that could impair ulterior bouts of injury. The following cases are not meant to be an exhaustive revision of all regenerative properties of ECs described in literature, but rather to show case how the secretion of angiocrine factors take a central role in these processes.(Rafii, Butler et al. 2016).

Regeneration and fibrosis in liver. In basal conditions, angiocrine signals control the growth and expansion of hepatocytes by allowing the proliferation of diploid *AXIN2*^{+/+} and T-box transcription factor (*TBX3*)^{+/+} cells that replenish the liver (Wang, Zhao et al. 2015). These progenitors are located adjacent to EC in the central vein. These specialized EC produce WNT2 and WNT9b, which maintain a healthy population of an *AXIN2*^{+/+} and *TBX3*^{+/+} double-positive cells. In other words, WNT2 and WNT9b are angiocrine factors. Tissue-specific deletion of endothelial *Wntless* in the adult mice exhausts these hepatic repopulating cells. *Wntless* is a specific transporter for WNT-ligands, which mechanistically explains the downregulation of these angiocrine factors in the extracellular space. Additionally, defined angiocrine expression of *Rspondin3*, a WNT agonist, by ECs in the central vein of the liver determines a β -catenin-dependent

metabolic zonation (Rocha, Vidal et al. 2015). Hence, both, repopulating potential and metabolic zonation of hepatocytes is determined by extrinsic, yet adjacent, angiocrine factors that arise from an instructive endothelial niche at the central vein of the liver.

Regeneration of lung epithelium. Lung capillary EC are instrumental in the regeneration of pulmonary epithelium. This has been demonstrated by removal of the left lung in mammals, procedure also known as pneumonectomy. This surgical ablation of the respiratory capacity leads to compensatory expansion of lung mass in the right lung, due to growth and further differentiation of alveolar epithelial stem and progenitor cells, in which alveolar type (AT) AT EC2 epithelial cells proliferate and regenerate lung epithelium. (Hogan, Barkauskas et al. 2014). This process is dependent of an intact angiogenic potential in ECs. Abrogation of the expression of FGFR-1 and VEGFR2 in adult ECs impairs this process (Ding, Nolan et al. 2011). Mechanistically, a growing vascular niche upregulates MMP-14, which later stimulates the epidermal growth factor (EGF) receptor on alveolar epithelial stem and progenitor cells via the cryptic EGF-like motif in HB-EGF and the gamma 2 chain of Laminin-5, and thus the proliferation of these cells leads to neo alveologenesi. Finally, endothelial cell specific knock out of MMP-14 in the adult blunts the regeneration capacity of alveolar epithelium with no repercussion on vascular perfusion. Taken altogether, endothelial-derived MMP-14 is an angiocrine signal that triggers the propagation of alveolar epithelial cells and basal cell progenitors.

1.1.4. Secretion of endothelial microRNA (miRNAs) in the modulation of cardiovascular biology.

Endothelial cells are capable of secreting microvessels containing micro RNAs. They have the main objective of silencing genes in distant tissues to modulate tissue homeostasis. Of note, MicroRNAs are short sequences of RNA, with an approximate length of 22 bp that bind to the untranslated region 3' (3' UTR) of their target messenger RNA to downregulate its expression. Mechanistically, Argonaute proteins (AGO) use these miRNAs as guide RNA to drive gene silencing, which may occur via 2 distinct processes, via a translation blockade and/or degradation of messenger RNA (mRNA)(Swarts, Makarova et al. 2014). miRNA can be assembled into a wide variety of membrane-derived vesicles for instance exosomes (30 to 150 nm), microvesicles (100 to 1000 nm) and apoptotic bodies (1 to 5 µm) to influence gene expression in

distant tissues and cells in a paracrine fashion (Rodrigues, Nimrichter et al. 2008). Due to the stability shown in the bloodstream, endothelial cells are thought to be an important contributor to the circulating pool of vesicle containing miRNA. Indeed, it has been shown that endothelial cells can secrete vesicle-containing miRNAs. To exemplify the extent of regulation endothelial cells, exert on tissues, the following EC-derived miRNAs are described. miR-222 is secreted by endothelial cells to reduce the tumor necrosis alpha (TNF- α) induced expression of intercellular adhesion molecule - 1 (ICAM-1), which recruits monocytes, in both, *in vivo* and *in vitro*. Importantly, in coronary artery disease (CAD), the relative abundance of miR-222 is reduced which may be partially responsible of an increased inflammatory state. (Jansen, Yang et al. 2015). Cardiac endothelial cells produce miR-126 and miR-210 to mediate survival of cardiac progenitor cells in transplanted cardiac cells in a myocardial infarction model. Mechanistically, miR-126 and miR-210 drive the upregulation of pro survival kinases and mediate a glycolytic switch in terminally differentiated cells cardiac which improves ejection fraction in mice. (Ong, Lee et al. 2014). Finally, endothelial-derived miRNAs can also be mediators of pathological states in target tissues. For instance, endothelial cells can produce miR-146a during peripartum cardiomyopathy. (Davis, Arany et al. 2020) a pregnancy-associated type of cardiomyopathy that could present itself as fatal condition, which contributes to an antimetabolic phenotype, impairing contractile and metabolic properties in cardiomyocytes (Halkein, Tabruyn et al. 2013). All in all, this exemplifies how endothelial cells are not only able to secrete metabolites, growth factors, hormones, and other archetypical molecules described during cell to cell communication, but also, as novel mechanisms of crosstalk are discovered, we see endothelial cells, and the whole vasculature in general, playing an important role and furthermore, assuming the central stage.

1.2. Regulation of organ homeostasis via controlling nutrient transport

1.2.1. Vascular endothelium as gatekeeper of substrate access.

Vascular endothelium, generally, can be rendered as a continuum lining of cells that barriers the contents present in the circulation (metabolites, nutrients, and growth factors) and the tissue they perfuse. However, ECs can form different types of capillary structures depending on the permeability needed in each capillary bed. They can be identified as continuous, fenestrated, and discontinuous. A continuous lining of

endothelial cells is characterized with tight junctions and a continuous basement membrane with the least permeability. Organs with continuous capillaries are skeletal muscle, lung, skin, central nervous system (CNS), heart, and adipose tissue (Bazzoni and Dejana 2004). Fenestrated capillaries are characterized by gaps, also referred as clefts, between adjacent ECs. Of note, despite these clefts, their basement membrane remains a continuum, therefore, they can be considered as the capillary structure with an intermediate permeability. Organs with these structures are typically endocrine organs which can secrete hormones more readily into circulation (Pavelka and Roth 2010). Finally, capillary structures with the highest permeability are discontinuous, also referred as sinusoids, where both endothelial cells and their basement membrane are discontinuous, which ensure maximal access of nutrient, metabolites, and factors to tissues (Buckley, Dickson et al. 1985, Braet and Wisse 2002, Shetty, Lalor et al. 2018). Organs with these vascular structures are liver, spleen, and bone marrow. Therefore, endothelial cells play different roles in modulating the access to different substrates in several organs, which ultimately furthers endothelial cell specificity as nutrient gatekeeper and modulator of metabolism.

Initially, it was believed that ECs were not able to actively modulate transport of glucose due to an allegedly inability to control expression of glucose transporter 1, GLUT1. A constitutive expression of GLUT1 would secure a continuous transendothelial transport and supply to tissues (Kaiser, Sasson et al. 1993). However, this view has been challenged by findings pointing that ECs downregulate expression of GLUT1 in CNS and heart when challenged for prolonged periods of time to high concentrations of glucose (Rajah, Olson et al. 2001, Alpert, Gruzman et al. 2005). Furthermore, it has been shown that when endothelial cells acquire a quiescent state, glycolysis is downregulated which leads to a GLUT1 upregulation in a NOTCH1-dependent manner. (Veys, Fan et al. 2020). Additionally, GLUT1 has been identified as a hypoxia inducible factor alpha (HIF1 α) (Huang, Lei et al. 2012).), which improves endothelial cell survival in poorly irrigated and thus hypoxic zones, as endothelial cells heavily rely on glycolysis for ATP generation (De Bock, Georgiadou et al. 2013). As central glucose can be to cellular energy homeostasis, it is actively regulated in endothelial cells depending their physiological state.

To continue exemplifying the role of vascular endothelium to control the access to nutrients and metabolites in organ tissues, endothelial cells express a variety of fatty acid transporters (FA) in continuous capillaries which form a tight barrier to FA of carbon chains equal or greater to 12 carbons. On the contrary, in the liver these tight junctions are not necessary as FA present in circulation are in direct contact with hepatocytes through liver sinusoids. Among the different FA transporters found in vascular tissue there are several worth to mention, for instance, fatty acid binding proteins (FABPs) (Antohe, Popov et al. 1998, Elmasri, Karaaslan et al. 2009), which mediate their intracellular binding and mobility, fatty acid transport proteins (FATPs) which are transporters located at EC cell membrane that mediate FA uptake and CD36 which is a translocase, key in the transport of fatty acid in skeletal muscle and heart tissue (Greenwalt, Watt et al. 1990). Over expression of FATP3 and FATP4 in ECs synergistically enhance lipid accumulation, which highlights their role in the uptake of long-chain fatty acid in ECs. Importantly, their co-expression is controlled by the stimulation of Vascular endothelial growth factor B (Hagberg, Falkevall et al. 2010)

1.3. Regulation of organ homeostasis through angiogenesis

Blood vessels are the functional unit of the vascular system. They are an assembly of endothelial cells, pericytes and smooth muscle cells that form the tubing fundamental in maintaining tissue homeostasis. In adults, around 95% of endothelial cells remain quiescent. However, ECs keep their capacity to respond to oxygen and nutrient shortage through the secretion of angiogenic growth factors from hypoxic and/or nutrient deprived cells which leads to the formation of new blood vessels in a process termed angiogenesis (Wang Y. 2010). Among those angiogenic growth factors, VEGFA is the canonical molecule to signal towards vascular growth. It acts through the stimulation of VEGF receptor 2, VEGFR2, at the cell membrane of EC. Through the establishment of a VEGFA gradient in the capillary milieu and an intricate cell-to-cell signaling, three functionally distinct transient cell fates can be identified. These are: *tip cells* which are present at the forefront of sprouting region, they are distinguished by high migration capability but suppressed proliferation; *stalk cells*, on the contrary are highly proliferative with limited migratory capacity; and finally *phalanx cells*

characterized by their quiescent state (Zecchin, Kalucka et al. 2017). (De Bock, Georgiadou et al. 2013).

1.3.1. Tip cells

The main function of tip cells is to lead the angiogenic sprout upon stimulation with angiogenic factors. Tip cells are characterized by their high migratory capacity and decreased proliferative potential. They exhibit a unique morphology: highly elongated shape with multiple cell membrane projections like filopodia and lamellipodia. In addition, their cell membrane is abundant in receptors and molecules in charge of extracellular matrix remodeling, helping them to reach tissue not yet perfused. (De Bock, Georgiadou et al. 2013, Schoors, Bruning et al. 2015).

1.3.2. Stalk cells

The angiogenic stimuli at the tip cell is pivotal for the generation of proliferative stalk cells. Stimulation of VEGFR2 elicits a transcriptional program that results in the upregulation of delta-like ligand 4 (Dll4) in the tip cells. Dll4 activates NOTCH signaling in adjacent EC. This signal begins a transcriptional program that leads, among other expression changes, to the downregulation of several cognate receptors of angiogenic factors like VEGFR2, VEGFR3, and NRP1, while at the same time, upregulating VEGFR1, a decoy receptor that dampens VEGFA stimuli to inhibit tip cell formation and promote a rather proliferative phenotype (Blanco and Gerhardt 2013).

1.3.3. Quiescent phalanx cells

After a new blood vessel has formed, ECs become quiescent again. Under steady state conditions most of ECs, up to 95%, are quiescent (Ricard, Bailly et al. 2021). Despite being the prevailing vascular growth state in mammals, little is known about the physiological role it has in the body or the metabolic determinants of a *functionally quiescent endothelium*.

Upon exiting the proliferative stage into quiescence, endothelial cells undergo a dramatic metabolic change, reducing their glycolysis ~40% while increasing their relative ATP production from mitochondria. A similar phenotype, can be observed

when overexpressing the Kruppel like factor 2 (KLF2), a transcription factor induced by high laminar shear stress in EC. In the nuclei, KLF2 suppresses the expression of PFKFB3 . Interestingly, overexpressing PFKFB3 in EC resumes their glycolytic activity and overrides KLF2 inhibition of growth (Lin, Natesan et al. 2010, Doddaballapur, Michalik et al. 2015).

The transcription factor fork head box O 1 (FOXO1) is yet another driver of EC quiescence. Interestingly, activation of FOXO1 is dependent of concentration of growth factors exposed to the cell. In the lack of growth factors, FOXO1 is present in the nucleus where it drives a gene transcription program. When angiogenic factors are present, such as VEGFA, FOXO1 suffers phosphorylation by AKT or serum/glucocorticoid –regulated kinase (SGK) which promote their displacement outside from the nucleus where is degraded by ubiquitin-proteasome pathway. In EC, FOXO1, along with FOXO3 and FOXO4 control growth state, survival, cell progression, energy metabolism, differentiation, and oxidative stress resistance. Specifically, FOXO1 acts as a pivotal modulator of proliferation and metabolic output. Endothelial specific murine FOXO1 overexpression constrains vascular development, provokes reduction of branching points and reduction of capillary diameter. Furthermore, FOXO1 controls endothelial quiescence through the inhibition of metabolic activity, reducing glycolysis and oxidative phosphorylation via blocking MYC signaling(Paik, Kollipara et al. 2007, Roudier, Milkiewicz et al. 2013, Wilhelm, Happel et al. 2016).

Despite that transcriptional programs that are determinant in endothelial cell identity are well-characterized and their resulting metabolic output largely described , little is known regarding how the secretion of angiocrine factors is modulated by these states to ultimately control whole body metabolic control and energy homeostasis.

1.4. Aims of the thesis

The sections above give a brief overview of aspects in which vasculature plays a central role in maintaining whole body homeostasis. However, the mechanistic understanding of whether and how vascular growth controls skeletal muscle homeostasis is largely lacking. In this project, I focused to answer four major questions in EC-Skeletal Muscle crosstalk

1. Whether and how a metabolic cross talk between EC and skeletal muscle tissue occurs.
2. Characterization of endothelial cell specific secretome *in vitro* using an unbiased proteomic approach.
3. Validation of potential targets using differential vascular states such as FOXO1 or NOTCH1.
4. Metabolic characterization of a novel angiocrine able to control skeletal muscle metabolism.

Importantly, the initial aims put forth in the thesis proposal at the commencement of this doctoral project shifted to the ones presented on this dissertation, which should be considered a final version. This was caused by the fact that the onset of metabolic adaptations to exercise, specially to the ones corresponding to *ad libitum* running wheel, require several weeks of training. Therefore, it compromised the time allocated to finish this project.

2. Measuring glycolytic and mitochondrial fluxes in endothelial cells using radioactive tracers

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2.1. Abstract

Endothelial cells (ECs) form the inner lining of the vascular network. Although they can remain quiescent for years, ECs exhibit high plasticity in both physiological and pathological conditions, when they need to rapidly form new blood vessels in a process called angiogenesis. EC metabolism recently emerged as an important driver of this angiogenic switch. The use of radioactive tracer substrates to assess metabolic flux rates in ECs has been essential for the discovery that fatty acid, glucose, and glutamine metabolism critically contribute to vessel sprouting. In the future, these assays will be useful as a tool for the characterization of pathological conditions in which deregulation of EC metabolism underlies and/or precedes the disease, but also for the identification of anti-angiogenic metabolic targets. This chapter describes in detail the radioactive tracer substrate assays that have been used for the determination of EC metabolic flux *in vitro*.

Key words. Endothelium, Metabolic flux rate, Radioactive tracer substrates, Glycolysis, Fatty acid oxidation, Glucose oxidation, Glutamine oxidation

2.2. Authors contributions

The general content and perspective of this chapter was proposed by Prof. Dr. Katrien De Bock. All the authors contributed to the technical expertise necessary to perform the experiments described in this protocol paper for the measurement of metabolic fluxes using radioactive tracers in endothelial cells. The design of the content, this is, the order and writing of each section of the manuscript, as well as the figure design was done by Abdiel Alvarado-Diaz and Dr. Koen Veys. The final figure formatting was carried out by Koen Veys. All authors reviewed and corrected the full contents of this manuscript chapter.

2.3. Introduction

Endothelial cells (ECs) line the inner surface of the vascular network and can stay quiescent for years. But in response to injury or during pathological conditions such as cancer, ECs are able to rapidly initiate the formation of new blood vessels in a tightly controlled but highly dynamic process termed sprouting angiogenesis. Sprouting angiogenesis is initiated by the secretion of pro-angiogenic factors from nearby cells into the microenvironment. Subsequently, one single EC is selected to become the leading “tip” cell and migrates toward the hypoxic and/or nutrient deprived area, while at the same time instructing its follower cells not to become tip cells. Instead, those “stalk” cells proliferate, extend the growing vascular sprout, and form a lumen. When two sprouts fuse, blood flow is reinitiated and ECs secrete a basement membrane while they return to their quiescent “phalanx” cell phenotype. Complex growth factor signaling networks and transcriptional signals control sprouting angiogenesis, but over the last few years it has become clear that these signals converge into metabolic changes that subsequently drive EC subtype (either tip, stalk, or phalanx cell) specification(Adams and Alitalo 2007, Carmeliet and Jain 2011).

ECs are glycolysis-addicted since the great majority of their ATP is produced by the glycolytic conversion of glucose to lactate, and only a limited number of glucose-derived pyruvate molecules are shunted into the TCA cycle for further oxidation(De Bock, Georgiadou et al. 2013). . Nonetheless, levels of glycolysis are strictly controlled in the quiescent endothelium(Doddaballapur, Michalik et al. 2015, Wilhelm, Happel et al. 2016) and reducing glycolysis suffices to promote a quiescent phenotype, even within the abnormal tumor environment (Schoors, De Bock et al. 2014, Cantelmo, Conradi et al. 2016). . During sprouting, glycolysis is upregulated in the migratory tip cells. In these cells, glycolytic enzymes, such as 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase 3 (PFKFB3) and hexokinase 2 (HK2), compartmentalize with F-actin at the so-called ATP hotspots of their membrane ruffles and thereby facilitate fast and local supply of ATP for migration (De Bock, Georgiadou et al. 2013, Yu, Wilhelm et al. 2017). Interestingly, proliferating stalk cells also require high glycolysis(De Bock, Georgiadou et al. 2013), but at the same time use fatty acid β -oxidation (FAO) for the synthesis of nucleotides for DNA replication. Fatty acid-derived carbons substantially

replenish the TCA cycle, and knock-down of carnitine palmitoyltransferase 1a (CPT1a), an enzyme involved in fatty acyl-CoA transport into the mitochondria and the rate-limiting enzyme for fatty acid β -oxidation, impairs EC proliferation in vitro (Schoors, Bruning et al. 2015, Wong, Wang et al. 2017), and reduces radial expansion and number of branch points of the retinal vascular network in vivo (Schoors, Bruning et al. 2015). Glutamine, however, is the main source of TCA cycle carbons (Kim, Li et al. 2017) and preventing glutamine anaplerotic via glutamine depletion or knock-down of glutaminase 1 (GLS1) reduced both nucleotide synthesis, protein synthesis, as well as lipid biosynthesis via reductive carboxylation (Huang, Vandekeere et al. 2017, Kim, Li et al. 2017). Interestingly, glutamine metabolism is also linked to asparagine metabolism, since angiogenic defects, observed upon inhibition of glutamine metabolism, could only be rescued by combined α -ketoglutarate and asparagine availability (Huang, Vandekeere et al. 2017). Other metabolic pathways have been studied in ECs but their role in EC specification in vivo is less clear. For instance, glucose flux into the oxidative pentose phosphate pathway (oxPPP) is required for EC proliferation and migration in vitro (Leopold, Walker et al. 2003). At the same time, oxPPP flux, via its rate-limiting enzyme glucose-6-phosphate dehydrogenase (G6PD), supports vascular redox homeostasis by limiting oxidative stress through the production of NADPH and the maintenance of sufficient nitric oxide levels (Leopold, Dam et al. 2007). For a detailed overview on EC metabolism, the reader is referred to elsewhere (Uebelhoer and Iruela-Arispe 2016, Sawada and Arany 2017, Eelen, de Zeeuw et al. 2018, Rohlenova, Veys et al. 2018).

The assessment of metabolic fluxes using radioactively labeled substrates has been fundamental to increase our understanding about EC metabolism. Here, we provide a detailed description on how to assess flux rate through several key EC metabolic pathways. The measurement of fatty acid β -oxidation (using palmitate as the lipid substrate) and glycolysis relies on the incorporation of ^3H into water, whereas the determination of glucose oxidation and oxPPP, as well as glutamine oxidation is dependent on the incorporation of ^{14}C into carbon dioxide.

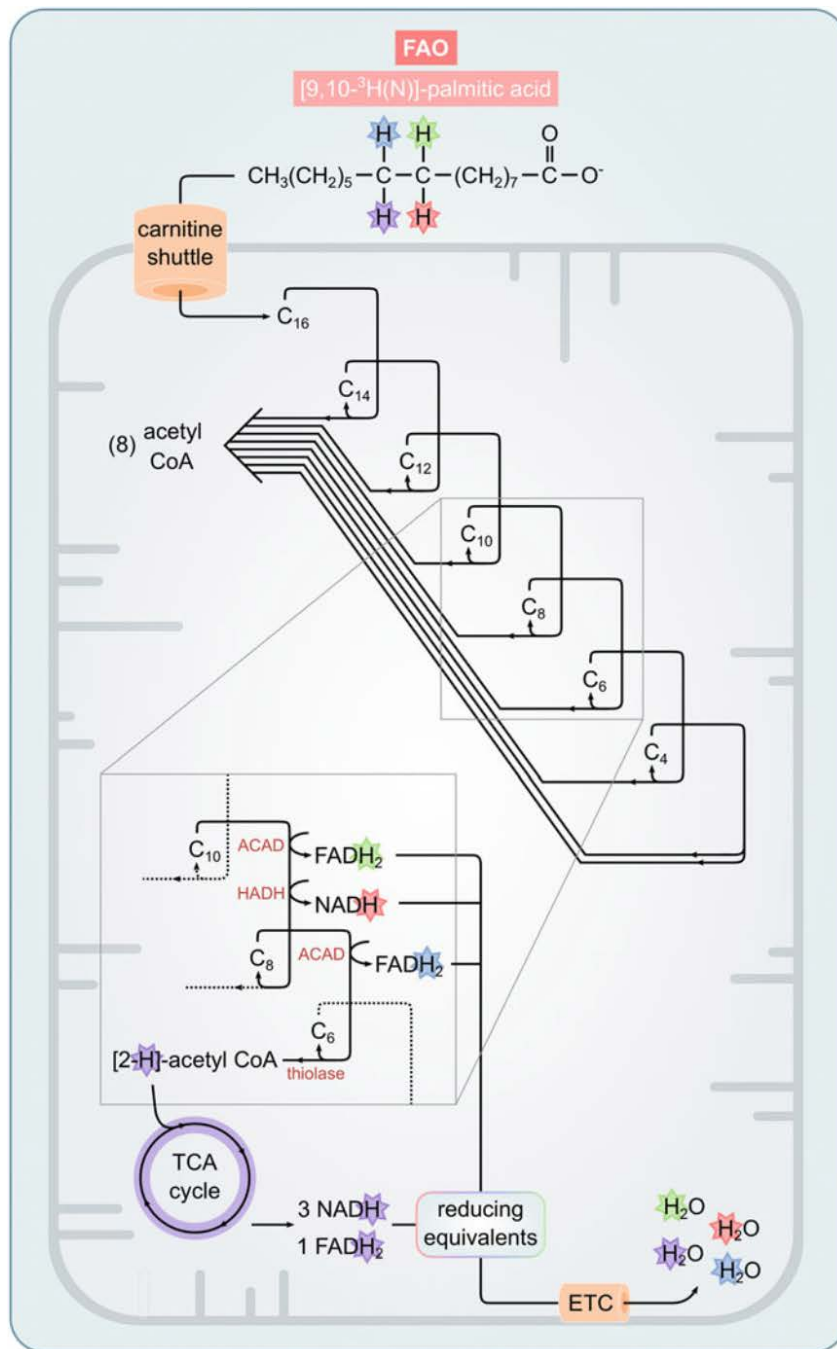
2.3.1. Fatty Acid β -Oxidation Flux

Palmitate, the 16-carbon, saturated long chain fatty acid is the product of fatty acid synthesis and is a precursor for longer fatty acids as well as a substrate for oxidation through fatty acid β -oxidation (FAO). FAO progresses in four-step cycles, each time

cleaving two carbons that form acetyl-CoA at the end of every cycle. Subsequently, these acetyl-CoA units can be used for further oxidation in the TCA cycle.

The determination of FAO flux in ECs can be done using *[9,10-³H(N)]-palmitic acid*. It is worth noting that the tritium atoms are chiral and that all four stereoisomers ((1) 9R-10R; (2) 9R-10S; (3) 9S-10R; (4) 9S-10S; see **Figure 2**). are present in equal amounts. The release of the hydrogen atoms is stereospecific and either in the form of [2-³H]-acetyl-CoA units or as tritiated FADH₂ or NADH molecules, which eventually produce ³H₂O in the electron transport chain (ETC) (see **Figure 2**) (Kler, Sherratt et al. 1992). This implies that for the β-oxidation of *[9,10-³H(N)]-palmitic acid*, 75% of the ³H atoms are indirectly released as ³H₂O and 25% are released as [2-³H]-acetyl-CoA. The fate of the ³H-label derived from [2-³H]-acetyl-CoA depends on the efficiency of its oxidation in the TCA cycle, and because TCA cycle flux is not always proportional to fatty acid β-oxidation (FAO) flux, the full recovery of the [2-³H]-acetylCoA into ³H₂O can thus be affected by changes in TCA cycle flux and lead to deviations in the experimental readout (Kler, Sherratt et al. 1992). Nonetheless, the utilization of *[9,10-³H(N)]-palmitic acid* is considered more accurate than the utilization of ¹⁴C-labeled palmitate tracers (Kler, Sherratt et al. 1992).

Fatty acids such as palmitate use the carnitine shuttle as a transport system for mitochondrial import. The first enzyme in the carnitine shuttle, CPT1, is rate-limiting for both fatty acylCoA transport and fatty acid β-oxidation (Drynan, Quant et al. 1996). Thus, pharmacological blockade of CPT1 using etomoxir can be used as a negative control for this assay



Palmitate starting positions

- ★ ³H at C-10 in pro-R
- ★ ³H at C-9 in pro-R
- ★ ³H at C-10 in pro-S
- ★ ³H at C-9 in pro-S

Figure 2. Schematic representation of FAO measurement using [9, 10-³H(N)]-palmitic acid. During the fourth cycle of FAO, palmitate is reduced to a 10-carbon molecule and the ³H atom that started at position 9 is released as ³H-labeled FADH₂ by the FAD-dependent acyl-CoA dehydrogenase (ACAD) from the pro-R form or released as ³H-labeled NADH by the NAD⁺-dependent L-3-hydroxyacyl-CoA dehydrogenase (HADH) from the pro-S form. In the next cycle, the 8-carbon molecule releases the ³H atom that started at position 10 by the ACAD enzyme as ³H-labeled FADH₂ from the pro-R form, whereas the pro-S form releases it as [2-³H]-acetyl-CoA, when its two carbons are removed by the thiolase enzyme. Further oxidation of [2-³H]-acetyl-CoA in the TCA cycle yields labeled FADH₂ and NADH. In the ETC, ³H₂O is produced from labeled FADH₂ and NADH. ACAD FAD-dependent acyl-CoA dehydrogenase, ETC electron transport chain, HADH NAD⁺-dependent L-3-hydroxyacyl-CoA dehydrogenase.

2.3.2. Glycolytic Flux

To measure glycolytic flux, tritium labeled glucose is used as the substrate tracer. The tritium is positioned at the fifth glucose carbon which means that when glycolysis is active, the ^3H label from the *D*-[5- ^3H (N)]-glucose tracer appears at the middle carbon of glyceraldehyde-3-phosphate (G3P). Subsequently, 2-phosphoglycerate (2PG) is converted to phosphoenolpyruvate (PEP) by the enolase enzyme, thereby producing tritium labeled H_2O (see **Figure 3**). Thus, flux rate through glycolysis is assessed by measuring the amount of tritium labeled water produced over time. Several inhibitors of glycolysis have been described in ECs. For instance, adding 2-deoxyglucose (2DG) to the assay medium in equal concentrations when compared to glucose rapidly reduces glycolytic flux (Schoors, De Bock et al. 2014). 2DG competes with glucose for uptake in the cell but cannot be further metabolized by glycolysis after its phosphorylation by hexokinase. Subsequently, the accumulation of the intracellularly trapped 2-deoxyglucose-6-phosphate leads to the inhibition of hexokinase activity and thus glycolytic flux. Also, pharmacological inhibition of the glycolytic regulator PFKFB3 using the small molecule 3-(3-pyridinyl)-1-(4-pyridinyl)-2-propen-1-one (3PO) reduces endothelial glycolysis with approximately 30%. (Clem, Telang et al. 2008, Schoors, De Bock et al. 2014)

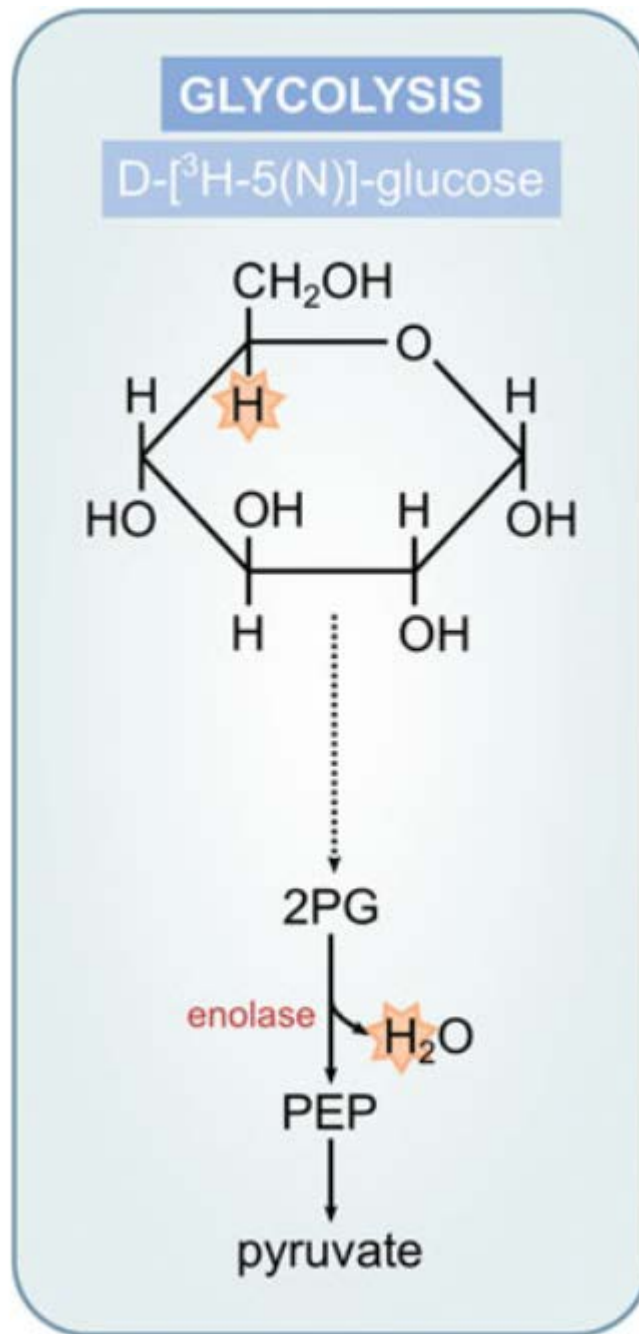


Figure 3. Schematic representation of glycolysis measurement using *D*-[5-³H(N)]-glucose. *D*-[5-³H(N)]-glucose produces ³H₂O when 2PG is converted to PEP at the enolase-step of glycolysis. 2PG 2-phosphoglycerate, PEP phosphoenolpyruvate

2.3.3. Glucose Oxidation and oxPPP Flux

To measure the oxidation of glucose, D-[6-¹⁴C]-glucose is used as a tracer. The sixth carbon does not necessarily end up in dihydroxyacetone phosphate (DHAP) and rather travels through glycolysis as the third triose carbon from G3P to pyruvate. Next, pyruvate enters the TCA cycle after decarboxylation to acetyl-CoA and in the theoretical model, half of the D-[6-¹⁴C]-glucose molecules lose their ¹⁴C label in the form of ¹⁴CO₂ during the third TCA cycle. By the seventh cycle, more than 95% of the ¹⁴C labels will have been released. Labeled carbons will arise as ¹⁴CO₂ during the conversion of isocitrate to α-ketoglutarate by isocitrate dehydrogenase (IDH) and during the conversion of α-ketoglutarate (α-KG) to succinyl-CoA by α-ketoglutarate dehydrogenase (OGDH), thereby being a measure of glucose oxidation **Figure 4a**. Theoretically, pyruvate could also enter the TCA cycle as oxaloacetate (OAA) through pyruvate carboxylation (Lee, Kang et al. 2003). In that case, the ¹⁴C label could already exit as ¹⁴CO₂ after the first TCA cycle. However, a significant contribution of pyruvate carboxylation flux in ECs remains largely unexplored.

Glucose oxidation flux in endothelial cells is detectable (De Bock, Georgiadou et al. 2013), but under normal culture conditions it is only a small fraction of the glucose that went through glycolysis. To determine oxPPP flux, a combination of two tracers is used in parallel experiments: *D*-[1-¹⁴C]-glucose and *D*-[6-¹⁴C]-glucose. As mentioned above, the latter is an indicator of glucose oxidation in the TCA cycle, whereas the first one produces ¹⁴CO₂ as a result of both glucose oxidation and oxPPP activity. Indeed, the first carbon of G6P can be released in the oxPPP as ¹⁴CO₂ when 6-phosphogluconate (6PG) is converted to ribulose-5-phosphate (Ru5P). Ru5P formation is particularly relevant as a precursor for nucleotide biosynthesis (Pandolfi, Sonati et al. 1995). If *D*-[1-¹⁴C]-glucose does not enter the oxPPP, it can run down the glycolytic pathway but unlike *D*-[6-¹⁴C]-glucose, where the ¹⁴C-labeled carbon bypasses DHAP, the labeled carbon needs to take a detour through DHAP before it ends up in G3P. At that point, both the [1-¹⁴C] and the [6-¹⁴C] end up at the same carbon position within G3P, making them indistinguishable. From there, both labeled carbons will follow the same route and therefore, their metabolic fate will be identical (see **Figure 4b**). To determine net oxPPP flux, two experiments – one using *D*-[1-¹⁴C]-glucose and one for *D*-[6-¹⁴C]-glucose – are run in parallel and ¹⁴CO₂ production from *D*-[6-¹⁴C]-glucose is subtracted from the ¹⁴CO₂ production from *D*-[1-¹⁴C]-glucose (this is C1-¹⁴CO₂ minus C6-¹⁴CO₂).

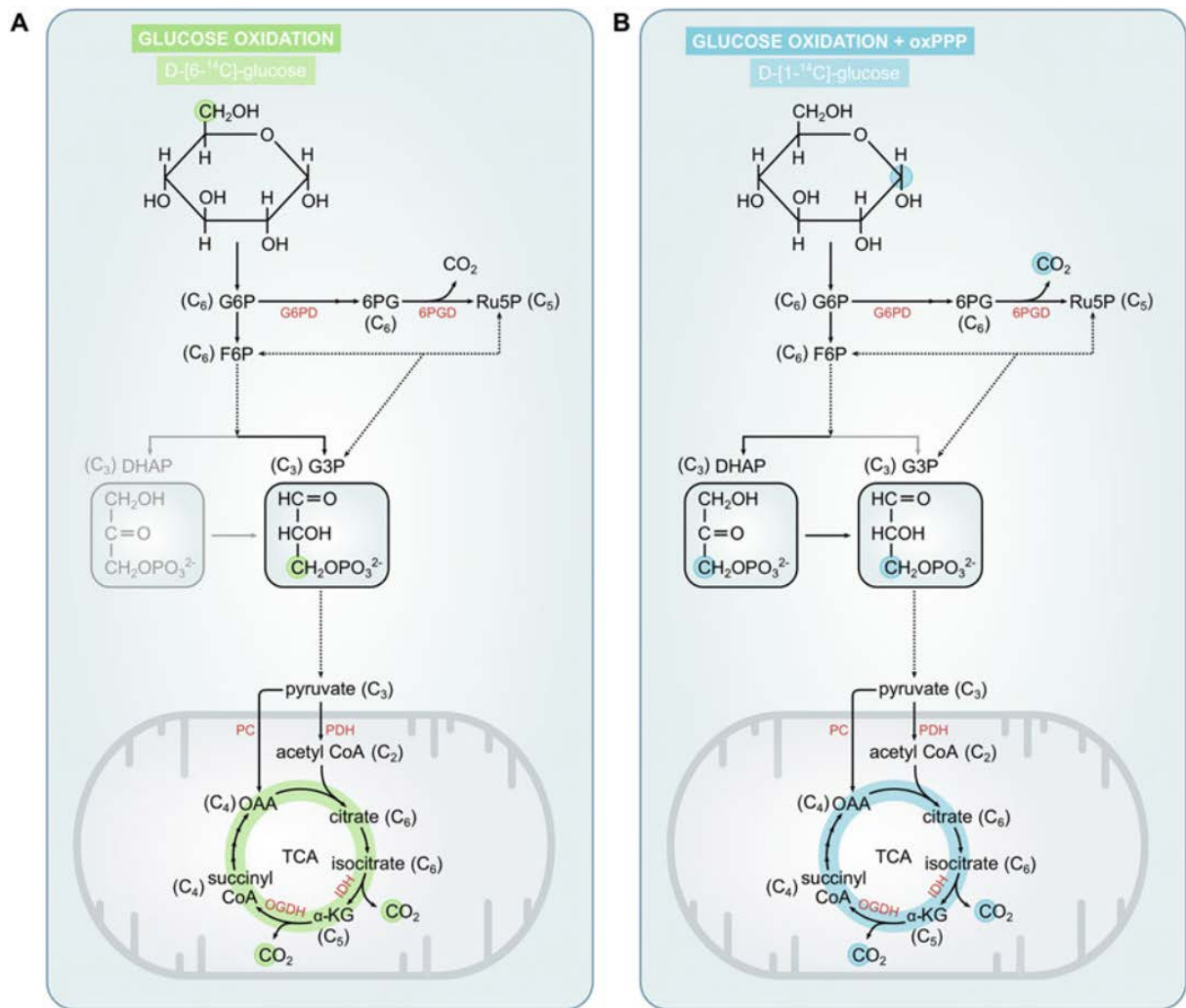


Figure 4. Schematic representation of glucose oxidation/oxPPP measurement using *D*-[6-¹⁴C]-glucose and *D*-[1-¹⁴C]-glucose. (a) Glucose oxidation: ¹⁴CO₂-production from the oxidation of *D*-[6-¹⁴C]-glucose only starts during the third TCA cycle at the IDH- and OGDH-step. (b) Glucose oxidation + oxPPP: *D*-[1-¹⁴C]-glucose releases CO₂ at the same steps of the TCA cycle, but can also release CO₂ at the 6PGD-step of the oxPPP. (a, b) Therefore, the subtraction of *D*-[6-¹⁴C]-glucose-derived ¹⁴CO₂-production from *D*-[1-¹⁴C]-glucose-derived ¹⁴CO₂-production yields a measure for the net oxPPP flux. 6PGD 6-phosphogluconate dehydrogenase, G6PD glucose-6-phosphate dehydrogenase, IDH isocitrate dehydrogenase, OGDH α-ketoglutarate dehydrogenase, PC, pyruvate carboxylase, PDH pyruvate dehydrogenase.

2.3.4. Glutamine Oxidation Flux

To assess glutamine oxidation flux, *L*-[¹⁴C(*U*)]-glutamine has been used as a tracer. Glutamine is first converted to glutamate by GLS1 (Leighton, Curi et al. 1987), which in turn is metabolized to α -ketoglutarate before entering the TCA cycle. There, fully labeled α -ketoglutarate immediately releases a first ¹⁴CO₂ when converted into succinyl-CoA by OGDH and a second ¹⁴CO₂ when isocitrate is converted to α -ketoglutarate by IDH. In the second cycle of the theoretical model, OGDH releases a third ¹⁴CO₂ from α -ketoglutarate, but due to the symmetry of succinate, the remaining ¹⁴C label is randomized and will gradually be released in the following cycles. It has recently been reported that about 10% of glutamine in ECs takes an alternative route via malic enzyme and pyruvate dehydrogenase in two subsequent decarboxylation steps before reentering in the TCA cycle (Huang, Vandekeere et al. 2017, Kim, Li et al. 2017). In this scenario, three ¹⁴CO₂ molecules are released in the first cycle but the start of the second cycle will not result in ¹⁴CO₂ release at OGDH. Thereafter, the metabolic fate of the label remains the same (see **Figure 5**). Although this alternative pathway only marginally affects the kinetics of ¹⁴CO₂ production, it coincides with the production of NADPH for biomass generation or redox maintenance rather than the generation of NADH. In vitro pharmacological interventions have proven useful for the study of endothelial glutamine metabolism. To determine the role of exogenously administered glutamine in ECs, the use of the SLC1A5 transporter inhibitor L- γ -glutamyl-p-nitroanilide (GPNA) has been reported (Sanchez, Carroll et al. 2015). Furthermore, inhibitors of GLS1, such as CB-839 or bis-2-(5-phenylacetamido-1,3,4-thiadiazol-2-yl)ethyl sulfide (BPTES), have also been used to show that glutamine is required for tip cell competitiveness in vessel sprouting (Durante, Liu et al. 2017, Huang, Vandekeere et al. 2017).

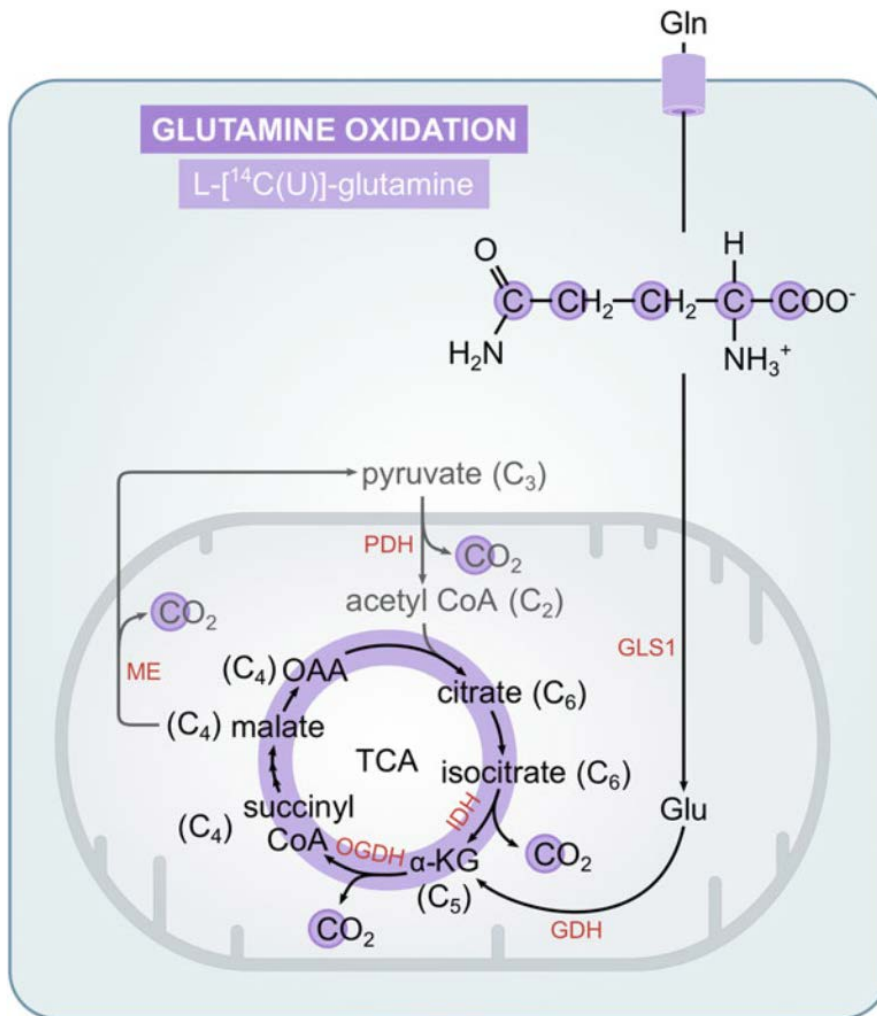


Figure 5. Schematic representation of glutamine oxidation measurement using L -[$^{14}\text{C}(\text{U})$]-glutamine. Glutamine oxidation: L -[$^{14}\text{C}(\text{U})$]-glutamine releases a first $^{14}\text{CO}_2$ when converted into succinyl-CoA by OGDH, and a second $^{14}\text{CO}_2$ when isocitrate is converted to α -ketoglutarate. In the second cycle of the theoretical model, α -ketoglutarate releases a third $^{14}\text{CO}_2$ by OGDH activity, but due to the symmetry of succinate, the remaining ^{14}C label is randomized and will gradually be released in the following cycles. About 10% of glutamine in ECs takes an alternative route via malic enzyme and pyruvate dehydrogenase in two subsequent decarboxylation steps before reentering in the TCA cycle. In this scenario, three $^{14}\text{CO}_2$ molecules are released in the first cycle but the start of the second cycle will not result in $^{14}\text{CO}_2$ release at OGDH. Thereafter, the metabolic fate of the label remains the same. *GDH* glutamate dehydrogenase, *GLS1* glutaminase 1, *IDH* isocitrate dehydrogenase, *ME* malic enzyme, *OGDH* α -ketoglutarate dehydrogenase.

2.4. Materials

2.4.1. General

1. β -counter.
2. Polyethylene hinge cap scintillation vials with 8 mL capacity.
3. Scintillation cocktail.
4. Regular endothelial cell culture medium such as EGM2 or M199 complete medium: 5.5 mM glucose, 2 mM glutamine, 20% FBS, supplemented with endothelial cell growth factors and heparin. Of note, the medium composition should be carefully considered depending on the specific research question. For instance, serum-free or fatty acid-free bovine serum albumin medium is required for the quantification of FAO flux because the precise fatty acid concentration should be known to calculate oxidation fluxes.

2.4.2. $^3\text{H}_2\text{O}$ Recovery for Fatty Acid β -Oxidation and Glycolysis Measurement

1. FAO tracer: *[9,10- ^3H (N)]-palmitic acid* at a stock concentration of 1 mCi/0.2 mL EtOH; this is 5 $\mu\text{Ci}/\mu\text{L}$. For the labeling solution, use 0.4 μL from the stock solution per mL culture medium. This results in a final concentration of 2 $\mu\text{Ci}/\text{mL}$ medium. Use medium that is serum-deprived or supplemented with fatty acid-free bovine serum albumin. Add 100 μM palmitate (nonradioactive), 50 μM fatty acid-free bovine serum albumin and 50 μM carnitine.
2. 10 mM palmitate stock solution (nonradioactive) in absolute EtOH. Store at -20 C in a glass vial.
3. 50 mM carnitine stock solution in PBS.
4. Glycolysis tracer: *D-[5- ^3H (N)]-glucose* at a stock concentration of 1 mCi/mL EtOH:Water (9:1); this is 1 $\mu\text{Ci}/\mu\text{L}$. For the labeling solution, use 0.4 μL from the stock solution per mL culture medium. The final concentration will be 0.4 $\mu\text{Ci}/\text{mL}$ medium.
5. Glass vials.
6. Rubber stoppers.
7. Hanging wells.
8. Filter papers: 1 cm x 6 cm.

9. Ultrapure water.

2.4.3. $^{14}\text{CO}_2$ Recovery for Glucose, Glutamine Oxidation and oxPPP Measurement.

1. Glucose oxidation tracer: *D*-[6- ^{14}C]-glucose at a stock concentration of 50 $\mu\text{Ci}/0.5$ mL EtOH:Water (9:1); this is 0.1 $\mu\text{Ci}/\mu\text{L}$. For the labeling solution, use 5.5 μL from the stock solution per mL culture medium. The final concentration therefore is 0.55 $\mu\text{Ci}/\text{mL}$ medium.

2. Glucose oxidation + oxPPP tracer: *D*-[1- ^{14}C]-glucose at a stock concentration of 50 $\mu\text{Ci}/0.5$ mL EtOH:Water (9:1); this is 0.1 $\mu\text{Ci}/\mu\text{L}$. For the labeling solution, use 5.5 μL from the stock solution per mL culture medium. The final concentration is 0.55 $\mu\text{Ci}/\text{mL}$ medium.

3. Glutamine oxidation tracer: *L*-[$^{14}\text{C}(\text{U})$]-glutamine at a stock concentration of 50 $\mu\text{Ci}/\text{mL}$ EtOH:Water (2:98); this is 0.05 $\mu\text{Ci}/\mu\text{L}$. For the labeling solution, use 5.5 μL from the stock solution per mL culture medium. The final concentration is 0.275 $\mu\text{Ci}/\text{mL}$ medium.

4. 10X hyamine-hydroxide. Dilute to 1X hyamine solution in ultrapure water. Only use the hyamine solution under the hood. 5. 12% perchloric acid. 6. Filter papers: 2.6 cm x 2.6 cm. 7. Ultrapure water.

2.5. Methods

2.5.1. $^3\text{H}_2\text{O}$ Recovery for Fatty Acid β -Oxidation and Glycolysis Measurement

A schematic representation of this assay is depicted in **Figure 6**

1. Day 1: the day before the experiment, seed 150,000 endothelial cells per well in a gelatin-coated 12-well plate (see Note 1).

2. Day 2: replace the culture medium (see **Note 2**) with 500 μL labeling solution. For FAO, use 2 $\mu\text{Ci}/\text{mL}$ [9,10- $^3\text{H}(\text{N})$]-palmitic acid supplemented with 100 μM palmitate (nonradioactive) (see **Note 3**) bound to 50 μM fatty acid-free bovine serum albumin and 50 μM carnitine. For glycolysis, use 0.4 $\mu\text{Ci}/\text{mL}$ D-[5- $^3\text{H}(\text{N})$]-glucose). Incubate the cells with labeling solution at 37 C for 6 h (FAO) or 2 h (glycolysis). Likewise, add the same volume of labeling solution to empty wells of a 12-well plate for the determination of background signal. Make sure to keep a small fraction of labeling solution (>12 μL) to calculate the specific activity (see **Note 6**).

3. Prepare rubber stoppers with hanging wells. Roll the filter papers (1 cm x 6 cm) and insert one into each hanging well. Hydrate each filter paper with 200 μ L ultrapure water.
4. After tracer incubation, transfer 400 μ L out of 500 μ L labeling solution from the 12-well plate to the glass vials. Aspirate the leftover tracer solution and prepare cell lysates for the determination of protein content. Close the glass vials with the rubber stoppers with hanging well and paper. If ECs attach poorly, add perchloric acid to stop any metabolic activity of unintentionally transferred cells.
5. Incubate the glass vials for at least 48 h at 37 C (see **Note 4**).
6. Prepare scintillation liquid vials containing 5 mL scintillation liquid and transfer each filter paper to its corresponding scintillation vial. Optional: wash the hanging well with 100 μ L H₂O and transfer the water into the same scintillation vials.
7. Shake the vials thoroughly and allow the filter papers to disperse their radioactive material into the scintillation cocktail for at least 2 h and up to 16 h before counting.
8. Count the scintillation vials using the correct protocol, measuring the disintegrations per minute (dpm) of the ³H label for at least 1 min per sample.
9. Rinse the equipment (glassware, rubber stoppers, and hanging wells) (**see Note 5**) to prevent the leftover radioactivity from adding noise to future experiments.
10. Typically, FAO or glycolytic flux is reported as nmol/hr/ μ g protein. See **Note 6** for calculations.

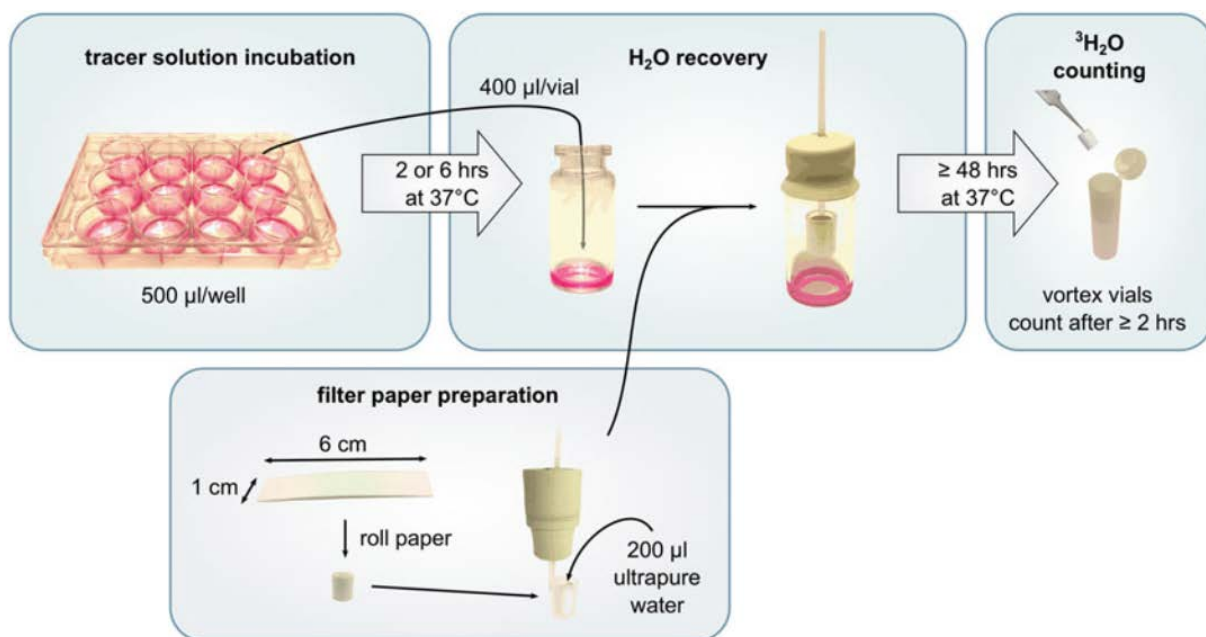


Figure 6. $^3\text{H}_2\text{O}$ recovery set-up (FAO/glycolysis). At the end of the tracer incubation period, $400\ \mu\text{L}$ of $500\ \mu\text{L}$ labeling solution is transferred from the 12-well plate to a glass vial. Once inserted inside the plastic hanging well, the filter paper is soaked with $200\ \mu\text{L}$ ultrapure water. Next, the glass vial is closed with its rubber stopper and incubated at $37\ \text{C}$ until equilibrium is reached (at least $48\ \text{h}$). Finally, the filter papers are transferred into scintillation vials containing scintillation cocktail and incubated at room temperature before counting

2.5.2. $^{14}\text{CO}_2$ Recovery for Glucose, Glutamine Oxidation, and oxPPP Measurement.

A schematic representation of this assay is depicted in **Figure 7**

1. Day 1: the day before the experiment, seed 150,000 endothelial cells per well in a staggered configuration (see **Note 7**) in a gelatin-coated 12-well plate.
2. Day 2: replace the culture medium with 500 μL labeling solution and incubate the cells for 6 h at 37 C. Likewise, add the same volume of labeling solution to empty wells of a 12-well plate for the determination of background signal.
3. Soak the filter papers (2.6 cm x 2.6 cm) in 1X hyamine solution.
4. At the end of the incubation period, add 100 μL of 12% perchloric acid to the labeling solution to lyse the cells, thereby releasing both labeled and unlabeled CO_2 (see **Note 8**). Close the well plate with the lid that contains hyamine-soaked papers on the inside to capture the CO_2 . Alternatively, place the hyamine-soaked filter paper on the top of the well immediately after adding perchloric acid (see **Note 9**). Repeat this for each individual well. When all wells are covered with a filter paper, carefully put the lid on the top of the plate. Wrap the plate in parafilm to prevent the CO_2 from escaping from the wells and incubate overnight at room temperature (see **Note 10**).
5. Day 3: after overnight capturing of CO_2 , transfer each filter paper to its corresponding scintillation vial containing 5 mL scintillation liquid.
6. Shake the vials thoroughly and allow the filter papers to disperse their radioactive material into the scintillation cocktail for at least 2 h and up to 16 h before counting.
7. Count the scintillation vials using the correct protocol, measuring the disintegrations per minute (dpm) of the ^{14}C label for at least 1 min per sample and calculate the flux rate after the necessary corrections (see **Note 11**).

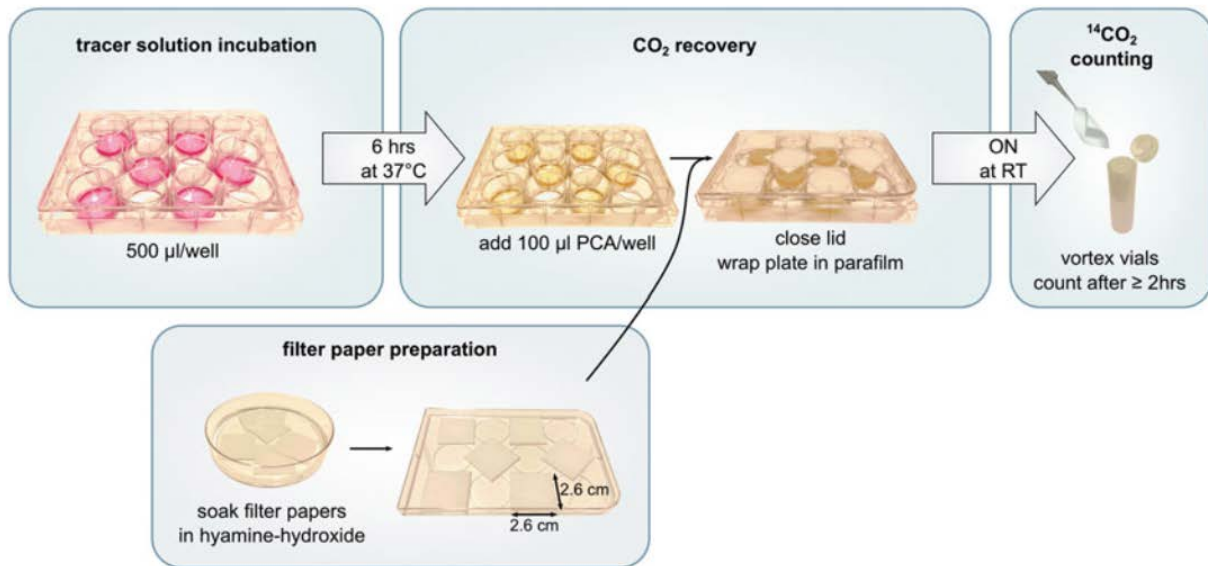


Figure 7. $^{14}\text{CO}_2$ recovery set-up (glucose oxidation/oxPPP/glutamine oxidation). Cells are seeded in a 12-well plate in a staggered configuration and are incubated with the labeling solution. After the addition of the strong acid, perchloric acid (PCA), every metabolic reaction is stopped and $^{14}\text{CO}_2$ is gradually released from the cells and medium. Thereafter, CO_2 is captured overnight by the hyamine-soaked papers upon closure of the well plate with its lid. These papers are either applied to the inside of the well plate lid as indicated here or are placed on the top of the well immediately after adding PCA. Finally, filter papers are transferred into scintillation vials containing scintillation cocktail and incubated at room temperature before counting

2.6. Notes

2.6.1. $^3\text{H}_2\text{O}$ Recovery for Fatty Acid β -Oxidation and Glycolysis Measurement

1. The assay can easily be scaled down to smaller well formats to lower cell numbers which allows the utilization of primary isolated ECs for flux assays. The volume of the labeling solution needs to be adjusted to the size of the well. When using lower cell numbers, researchers should consider increasing the labeling time to get more reliable results.
2. For FAO: in order to be able to calculate the FAO flux, the exact concentration of nonradioactive fatty acids (palmitate) has to be known. Since the precise concentration of fatty acids in the serum is usually not known, either serum-free or fatty acid-free medium has to be used.
3. Nonradioactive palmitate precipitates upon long-term storage in - 20 C. Make sure it is completely solubilized before adding it to the labeling solution.
4. Handle the glass vials gently, as any spillover of labeling solution onto the filter paper will lead to contamination of the $^3\text{H}_2\text{O}$ signal with the labeled substrate tracer signal. For the same reason, avoid contact between the glass vial and the hanging well. Contamination with labeling medium is apparent by the presence of erroneously high values in the dataset.
5. Check national biosafety regulations concerning the proper disposal of radioactive waste. To remove any leftover radioactivity from the glass vials, aspirate the labeling solution and wash 3 times with tap water, before autoclaving the glass vials. Wash the rubber stoppers and hanging wells with a detergent and wash two times in tap water before leaving them out to air dry. It is important that the medium inside the glass vials is free of bacterial and fungal contaminations, to prevent organisms from metabolizing the substrate tracers to $^3\text{H}_2\text{O}$, which would give rise to flux overestimation.
6. Calculations: To report FAO or glycolytic flux as nmol/h/ μg protein, we suggest running a BCA assay to determine protein content or correct the measurement by the number of cells. Also, since only a part of the total $^3\text{H}_2\text{O}$ produced by the cells is captured in the filter paper, the measured dpm values only represent partial $^3\text{H}_2\text{O}$ recovery and not the total $^3\text{H}_2\text{O}$ -production. Therefore, a correction factor has to be applied in order to calculate total $^3\text{H}_2\text{O}$ production. The percentage of $^3\text{H}_2\text{O}$ -recovery can be easily determined through the use of a known amount of commercially available

$^3\text{H}_2\text{O}$ (e.g., 2 $\mu\text{Ci}/\text{mL}$ culture medium). As long as all experimental conditions are kept identical, it is sufficient to calculate one correction factor for all future FAO or glycolysis experiments. However, since this factor is influenced by the amount of radioactive metabolite, equilibration time, incubation temperature, volume of the glass vial, and volume of the medium, a new correction factor needs to be developed and applied every time specific conditions are changed. Lastly, background signal is subtracted from corrected dpm values and the resulting net dpm values are converted to nmol substrate. Once the substrate concentration (nmol glucose or palmitate per μL) and the radioactivity concentration are known (dpm per μL ; typically, duplicates of 2 μL and 4 μL from the labeling solution are measured by liquid scintillation counting), the specific activity (dpm/nmol) can be calculated for its use as a conversion factor.

2.6.2. $^{14}\text{CO}_2$ Recovery for Glucose, Glutamine Oxidation, and oxPPP Measurement

7. Cells are seeded in a staggered configuration to prevent possible cross-contamination of label from neighboring wells during the CO_2 capturing process.

8. Do not inhale radioactive $^{14}\text{CO}_2$; therefore, always work under the hood while performing $^{14}\text{CO}_2$ recovery experiments.

9. Because CO_2 rapidly dissipates when plates are removed from the incubator, both the addition of the strong acid and the placement of the hyamine-soaked paper on the top of the well have to be done fast. It is strongly advised to work plate by plate; this will minimize CO_2 loss and variability within the technical replicates.

10. Gently handle the plates, since direct contact between the radioactive medium and filter paper will contaminate the $^{14}\text{CO}_2$ signal with the labeled substrate tracer signal and will give rise to erroneous data.

11. Calculations: Experimentally, correction factors have not been determined for $^{14}\text{CO}_2$ recovery. Therefore, this method is always a semiquantitative measure in which absolute oxidation rates (e.g., nmol/h) cannot be calculated. However, the dpm-values should be comparable between replicates, and relative between treatments, within an experiment. A parallel experiment should be run to correct for protein or DNA content; alternatively, number of cells can also be used.

3. Endothelial cells secrete GDF15 in a FOXO1 dependent manner

Running title: Vascular control of GDF15.

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Status: In preparation

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3.1. Abstract

Endothelial cells are important gatekeepers of organ homeostasis and metabolism. It has long been determined that impaired vascular function is associated with tissue dysregulation and metabolic disorders, but whether endothelial cells contribute to metabolic homeostasis beyond ensuring tissue perfusion is poorly described. Vascular expression of FOXO1, master regulator of endothelial quiescence, is often found to be upregulated in such pathological conditions. Using an unbiased proteomics approach to analyze the endothelial secretome, we found that endothelial cells secrete growth differentiation factor (GDF15). Moreover, FOXO1 activation reduced while inhibiting FOXO1 increased *gdf15* expression. In vivo, endothelial specific deletion of GDF15 did not reduce circulating GDF15 levels. However, deletion of FOXO1 (*Foxo1* Δ EC) increased *gdf15* expression (>4 fold) in endothelial cells isolated from different tissues and upregulated GDF15 in urine and serum. Furthermore, *Foxo1* Δ EC mice showed a significant decrease in food intake, suggesting that physiological changes in circulating GDF15 suffice to control food intake. This work identifies FOXO1 as a direct regulator of GDF15 in ECs and highlights the critical role of endothelial cells in whole body metabolic control

3.2. Authors contributions

The general content, perspective and supervision of this manuscript chapter was proposed by Prof. Dr. Katrien De Bock. The design, execution, and analysis of all the experiments, writing the full manuscript content, and creation of all figures was carried out by Abdiel Alvarado Diaz. Over the course of this project, several contributors have significantly supported its realization in forms of assistance during the execution and/or analysis of some experiments or figures. Moheb Ghobrial supported the isolation of primary mouse endothelial cells associated with figures 9 and 11, furthermore, he assisted in the design and formatting of all figures, and correction of manuscript. Dr. Fateme Jaleh supported the methodological approach, design, execution, and analysis of proteomic characterization of endothelial cell secretome associated with figure 8, additionally, she supported the qPCR screening that revealed GDF15 as a candidate of interest for this project and technically assisted with experiments associated with figure 10. Dr. Zheng Fan supported with valuable input in the design of the all the experiment, donated material such as lentiviral construct for ATF4 knockdown experiments associated with extended data figure 2 and proved valuable throughout the duration of this chapter. Dr. Adhideb Ghosh supported the bioinformatic proteomic analysis associated with figure 8 and extended data figure 1. Guillermo Turiel also supported the bioinformatic analysis associated with figure 8. Dr. Evi Masschelein, as our animal experimental specialist gave technical assistance to all the *in vivo* experiments associated to figures 9 and 11. Dr. Tatiane Gorski assisted technical support manipulating the cell sorter associated to figures 9 and 11. Dr. Paola Gilardoni provided technical assistance in the conduct of all experiments. The general supervision and correction of this manuscript was done by Abdiel Alvarado Diaz and Prof. Dr. Katrien De Bock.

3.3. Introduction

The physiological role of blood vessels goes beyond acting as simple blood conduits that passively regulate the distribution of blood throughout the entire organism. The endothelium can control tissue function and organ homeostasis, and this is particularly true for the regulation of tissue metabolic control (Graupera and Claret 2018, Pi, Xie et al. 2018, Hasan and Fischer 2021): blood vessels adapt oxygen and nutrient delivery to the metabolic requirements of the tissue by regulating of tissue perfusion via altering capillary recruitment or by forming of new blood vessels through angiogenesis. Moreover, endothelial cells can actively control the transport of nutrients, particularly lipids, from the blood to the parenchymal tissues. The signals that mediate this crosstalk between endothelial cells and their microenvironment are increasingly be studied but still remain poorly understood. For instance, the release of 3-hydroxybutyrate from skeletal muscle enhances endothelial lipid transport (Jang, Oh et al. 2016). On the other hand, endothelial cells also release angiocrine factors, such as nitric oxide (NO), which determines amongst others vascular tone (Pi, Xie et al. 2018). Another example is the secretion and deposition of laminins by islet ECs in the pancreas to stimulate β -cell proliferation and insulin production (Nikolova, Jabs et al. 2006).

Metabolic homeostasis and energy balance are dependent on a tightly orchestrated crosstalk between different highly vascularized metabolic tissues such as muscle, liver, adipose tissue and pancreas on one side, and nutrient intake on the other side. It is therefore not surprising that pathological processes in many of these tissues critically contribute to the development of metabolic disorders such as obesity. Endothelial dysfunction has been linked to metabolic diseases, and vascular complications are a major cause of morbidity and mortality in obese and diabetic patients (King, Aubert et al. 1998). Hyperlipidemia, hyperglycemia and other metabolic stressors result into endothelial dysfunction. However, emerging evidence indicates that endothelial cells do not solely act as passive responders to metabolic stress, but directly contribute to the development of metabolic disorders. For instance, obesity in high-fat diet fed mice (Nwadozi, Roudier et al. 2016) as well as in humans (Karki, Farb et al. 2015) has been associated with increased endothelial expression and activity of the transcription factor forkhead box O-1 (FOXO1), a master regulator of both quiescence and metabolism in endothelial cells (Wilhelm, Happel et al. 2016). Also, endothelial FOXO1 levels are higher in a mouse model of type I diabetes characterized by high glucose levels (Shi,

Fan et al. 2018). Interestingly, reducing endothelial FOXO1 can alleviate metabolic dysfunction and vascular dysfunction caused by obesity. Loss of endothelial FOXO1 reduced weight gain upon the intake of a high-fat diet and improved metabolic control (Rudnicki, Abdifarkosh et al. 2018). These metabolic alterations were attributed to enhanced vascular density in white adipose tissue combined with increased glycolytic activity of the endothelial cells. Moreover, FOXO1 inhibition improved impaired revascularization and wound closure in a mouse model for type 1 diabetes by improving endothelial function (Shi, Fan et al. 2018). However, whether FOXO1 also controls the expression of angiocrine signals that can modulate systemic metabolism and energy balance was not investigated.

Here, we used unbiased proteomics to identify angiocrine factors. We identify the growth and differentiation factor 15 (GDF15; also known as macrophage inhibitory cytokine-1) as one of the most abundantly secreted angiocrine factors. Administration of GDF15 has recently been proposed as an anti-obesity drug due to its ability to suppress appetite in mice and non-human primates (Mullican, Lin-Schmidt et al. 2017, Xiong, Walker et al. 2017). We show that endothelial deletion of *gdf15* does not affect circulating GDF15 levels and food intake under physiological conditions. However, we found that endothelial *gdf15* expression in a quiescent endothelium is repressed by FOXO1. Finally, deletion of FOXO1 in the endothelium enhances circulating GDF15 and reduces food intake. Our data suggest that the endothelium can control systemic metabolism through the modulation of food intake.

3.4. Results

3.4.1. Proteomic characterization of the endothelial secretome

To identify which proteins are secreted by ECs proteins, we decided to use E4ORF1⁺ HUVECs (Seandel, Butler et al. 2008). Lentiviral introduction of *E4ORF1* in ECs allows the long-term survival and growth of ECs under serum free conditions, which allows the analysis of the secretome in the absence of cellular stress induced by serum starvation. We thus generated conditioned medium (CM) from E4ORF1⁺ HUVECs cultured under serum free conditions and subjected it to proteomic analysis (**Figure 8A**). Non-conditioned (NC) medium was used as a control, this is, fresh medium that has not been in cultured in the presence of cells. Protein identification and quantification were performed using MaxQuant and Andromeda search engine (Cox and Mann 2008, Cox and Mann 2011, Cox, Neuhauser et al. 2011). In total, 818 proteins were detected during the analysis. Sample clustering clearly differentiated between CM and NC, implying a high correlation between the samples which enables the identification of relevant protein targets for further screening (**Figure 8B**). Furthermore, we found that 227 proteins were significantly upregulated and abundant in CM whereas 20 proteins were more abundant in NC (**Figure 8C, Supplemental table 1**). Out of the 227 proteins upregulated in CM, 116 proteins were uniquely found in CM (**Figure 8C**, black rectangle), which means that they were secreted by ECs and not detected in NC medium. Taking into consideration uniquely identified proteins, top 40 most abundant proteins are shown (**Extended data figure 1A**). Subsequently, we used the bioinformatic tools SignalP5 (<http://www.cbs.dtu.dk/services/SignalP/>), which identifies proteins with a signal peptide, and Secretome 2.0 (<http://www.cbs.dtu.dk/services/SecretomeP/>) which predicts non-classical protein secretion, to assemble a *bona fide* endothelial cell-secreted protein repository containing 113 proteins (**Figure 8D**). The remaining 114 proteins were mapped against the databases ExoCarta (Keerthikumar, Chisanga et al. 2016) and Vesiclepedia (Kalra, Simpson et al. 2012) to determine whether they have been reported as being secreted through non-classical pathways such as microvesicles and exosomes. Indeed, 105 proteins were overlapping with ExoCarta and 110 against vesiclepedia (**Figure 8D**). Finally, we performed cellular component categorization analysis using GSEA on all 227 proteins, yielding a total of 229 identifiers from which 226 were unambiguously mapped, which showed that main categories were related to vesicle, extracellular

space, membrane, and membrane enclosed lumen, confirming their secretory nature (**Figure 8E**).

Finally, to determine which proteins could exert angiocrine signaling to surrounding tissues, we focused on the proteins that were previously identified as ligands. We used the Cellphone DB repository of curated receptors, ligands, and their interactions to delimitate cell-to-cell interactions via factor secretion. From 227 predicted to have a signal peptide or undergo non-classical secretion, the list was reduced to 29 proteins that were identified either as ligands, extracellular matrix (ECM) proteins or receptors (**Figure 8F**). Of them, 16 proteins were known curated ligands (**Figure 8G**). Many of those have a known role in endothelial biology: For instance, thrombospondin 1 (TSP1) is a negative regulator of angiogenesis (Lawler and Lawler 2012) which is also involved platelet adherence to vasculature (Bonney, Daenens et al. 2006). The secretion of matrix metalloproteases (MMPs) such as MMP2 by EC occurs via vesicle packaging and is stimulated by VEGFA (Taraboletti, D'Ascenzo et al. 2002). Also, Von Willebrand factor (VWF) plays a crucial role in hemostasis, inflammation, permeability and angiogenesis both inside and outside the EC (Jaffe, Hoyer et al. 1974, Randi, Smith et al. 2018). Activated ECs also release intercellular adhesion molecules (Videm and Albrigtsen 2008) which normally play a crucial role in leukocyte transendothelial migration. Finally, we decided to focus on growth and differentiation factor 15 (GDF15; also known as macrophage inhibitory cytokine-1).

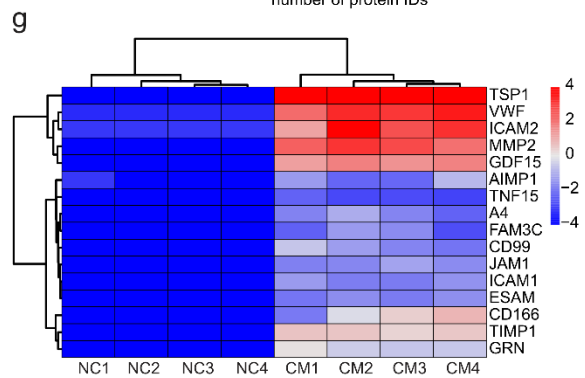
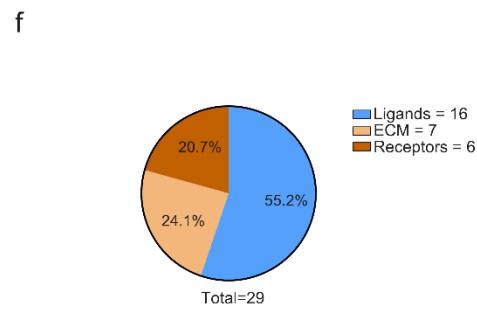
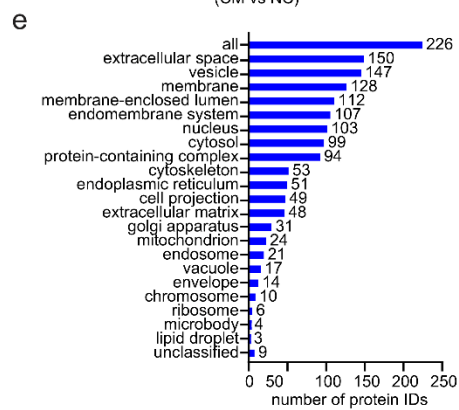
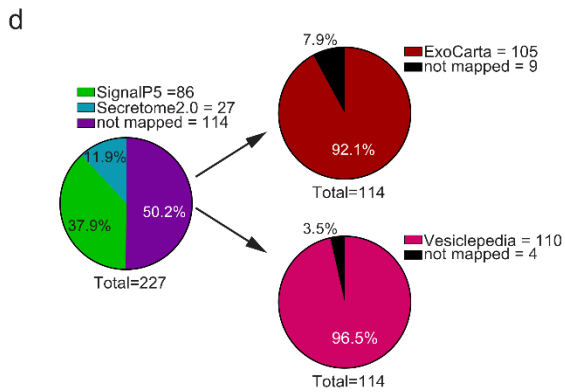
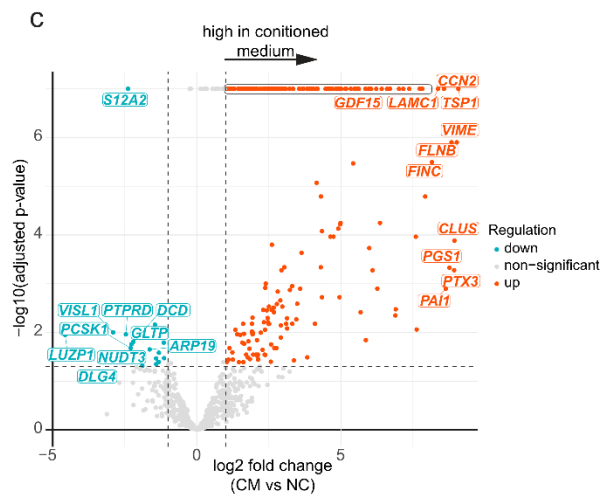
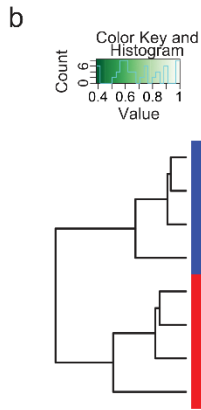
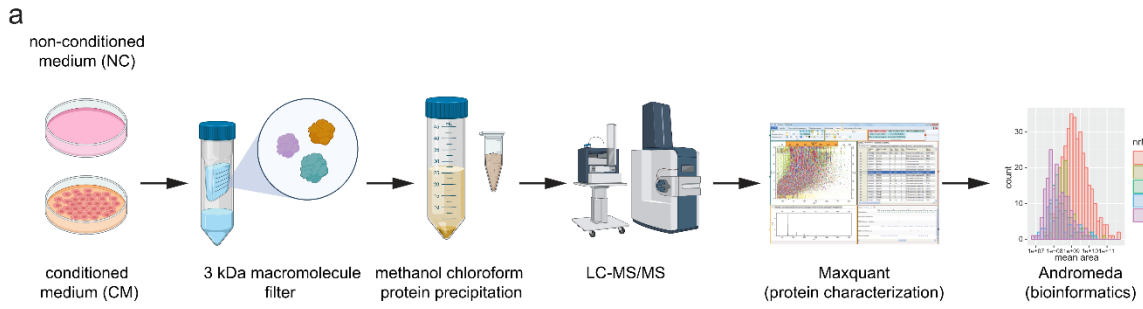


Figure 8. Workflow of generation and quantification of secretome of HUVECs produced in serum-free conditions **a.** Culturing conditions of E4ORF1 HUVECs, conditioned medium processing and LC-MS/MS workflow. **b.** Clustering depending on sample correlation and protein expression profile. Side colors on the left side of the heatmap indicate groupings. **c.** Volcano plot showing adjusted p- values computed from moderated t-test between all quantified proteins in conditioned media and non-conditioned control in endothelial secretome. Noted in a rectangle, all uniquely identified proteins in CM. **d.** Pie chart (left) shows overlap between proteins found in CM and proteins to be predicted for secretion either by the predicted presence of signal peptides or to undergo non-classical secretion. Pie charts (right) shows overlap between proteins not mapped to be secreted in endothelial secretome and proteins that have been identified in isolated vesicles (top) and exosomes (bottom), in the vesiclepedia and exocarta datasets respectively. **e.** Categorization of identified EC secreted proteins according to their cellular localization. **f.** Distribution of protein classes among those that have an interactor role according to Cellphone DB in endothelial secreted proteins. **g.** Heatmap of relative abundance determined by escalated expression in identified secreted ligand proteins according to Cellphone DB

3.4.2. Endothelial deletion of GDF15 does not contribute to circulating GDF15 under physiological conditions.

GDF15 is a stress responsive cytokine and a distant member of the transforming growth factor β superfamily (Bootcov, Bauskin et al. 1997, Tsai, Husaini et al. 2018). GDF15 signals through its receptor glial-derived neurotrophic factor receptor alpha-like (GFRAL), which is highly localized in brain areas which are involved in appetite control (Emmerson, Wang et al. 2017, Hsu, Crawley et al. 2017, Mullican, Lin-Schmidt et al. 2017, Yang, Chang et al. 2017). Due to its ability to suppress appetite in mice and non-human primates, pharmacological administration of GDF15 has been proposed as an anti-obesity drug (Mullican, Lin-Schmidt et al. 2017, Xiong, Walker et al. 2017). Under physiological conditions, *gdf15* is modestly expressed in many tissues and fairly high baseline circulating levels have been detected (Fairlie, Moore et al. 1999, Tsai, Husaini et al. 2018). In addition, *gdf15* can be induced in many tissues in response to specific stressors. Interestingly, some *in vitro* studies have suggested that ECs also efficiently upregulate *gdf15* in response to stressors such as high glucose (Li, Yang et al. 2013) or during senescence (Ha, De Torres et al. 2019), but a contribution of ECs to GDF15 physiology *in vivo* has not been shown. We first confirmed using an ELISA that on low passage WT HUVECs cultured under angiogenic conditions secrete GDF15 (**Extended data figure 1B**). To investigate the *in vivo* relevance of endothelial GDF15 secretion, we subsequently decided to generate endothelial specific *gdf15* knock-out mice. To do so, we intercrossed *gdf15^{Lox/Lox}* mice with a tamoxifen-inducible endothelial specific *pdgfb-Cre^{ERT2}* (Claxton, Kostourou et al. 2008), leading to the generation of *gdf15^{ΔEC}* mice (**Figure 9A**). Despite efficient deletion of *gdf15* mRNA levels in ECs isolated from different organs (muscle, white adipose tissue, heart), with an average approximate efficiency of 80% (**Figure 9E, F, G**), we did not observe altered GDF15 levels in the blood of *gdf15^{ΔEC}* mice (**Figure 9B**). Moreover, food intake was not affected by loss of endothelial *gdf15* (**Figure 9C**). Interestingly, upon lipopolysaccharide stimulation, but not metformin, circulating GDF15 is decreased in *gdf15^{ΔEC}* mice compared to controls (**Figure 9D**) Altogether, this data indicates that under basal physiological conditions, endothelial cells do not contribute to the maintenance of basal circulating GDF15 levels, but rather contribute in LPS inflammatory response (Luan, Wang et al. 2019).

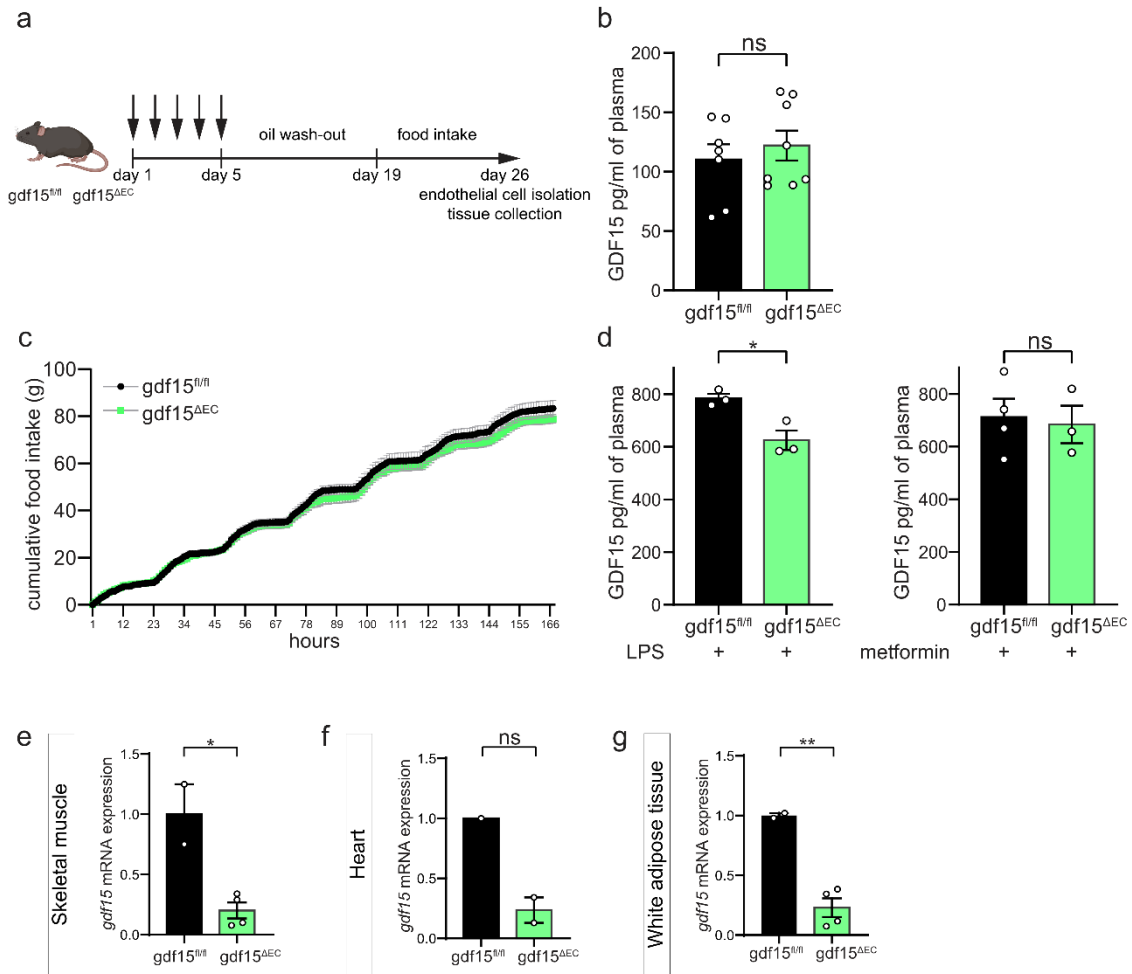


Figure 9. Angiocrine GDF15 does not significantly contribute to circulating levels in basal conditions, only contributes in inflammatory response. **a.** Schematic of tamoxifen injection in *gdf15^{ΔEC}*. **b.** GDF15 ELISA from plasma in *gdf15^{ΔEC}* ($n=8$) mice or controls ($n=7$) (Student-*t* test). **c.** Cumulative food intake over 7 days, 167 h, in *gdf15^{ΔEC}* ($n=7$) mice or controls ($n=6$). (Two-way ANOVA and Sidak's multiple comparison). **d.** GDF15 ELISA after inflammatory challenge with LPS, blood was sampled 2 h after in *gdf15^{ΔEC}* ($n=3$) mice or controls ($n=3$) (Student-*t* test), and GDF15 ELISA after metformin administration 600 mg/kg body weight, blood was sampled 6 h after in *gdf15^{ΔEC}* ($n=3$) mice or controls ($n=4$) (Student-*t* test). Knockdown efficiency in *gdf15^{ΔEC}* compared to controls in freshly isolated endothelial cells from **e** skeletal muscle (controls, $n=2$; *gdf15^{ΔEC}* $n=4$), **f** heart (controls, $n=1$; *gdf15^{ΔEC}* $n=2$), **g**, or white adipose tissue (controls, $n=2$; *gdf15^{ΔEC}* $n=4$). Housekeeping gene 18S was used as control for figure panels **e**, **f**, **g**. Significant values are represented in asterisks as follows * $P<0.05$, ** $P<0.01$, *** $P<0.001$, and **** $P<0.0001$.

3.4.3. Endothelial cells control GDF15 in a FOXO1 dependent manner

To further understand the mechanisms that might control GDF15 release from ECs, we decided to evaluate whether GDF15 is controlled by NOTCH1 and/or Forkhead Box O1 (FOXO1). Both FOXO1 as well as NOTCH are essential regulators of endothelial quiescence and vascular homeostasis (Wilhelm, Happel et al. 2016, Kalucka, Bierhansl et al. 2018). Moreover, increasing evidence also suggests that deregulated FOXO1 and NOTCH1 contribute to endothelial dysfunction during obesity. FOXO1 protein levels are increased in capillary ECs from mice on a high-fat diet (Nwadozi, Roudier et al. 2016) and removing FOXO1 from ECs during high-fat diet prevents the development of insulin resistance (Rudnicki, Abdifarkosh et al. 2018). NOTCH1 activity is also increased in diabetic vessels (Miloudi, Oubaha et al. 2019) and sustained genetic activation of Notch signaling lowers insulin sensitivity (Hasan, Jabs et al. 2020). On the other hand, inhibiting NOTCH1 impairs fatty acid transport into the heart (Jabs, Rose et al. 2018) Whether endothelial FOXO1 or NOTCH1 can control the release of angiocrine factors involved in metabolic control, such as GDF15, is not known.

To address whether NOTCH1 controls *GDF15* expression, we activated NOTCH1 by culturing HUVECs in the presence of the Notch ligand DLL4. DLL4 treatment activated NOTCH1, since the expression of canonical downstream targets such as *HES1*, *HEY1/2*, and *NRARP* was increased, but NOTCH1 activation did not affect *GDF15* expression (**Figure 10A**). Also, inhibition of NOTCH signaling using the gamma-secretase inhibitor DAPT reduced the expression of NOTCH1 targets but did not affect *GDF15* (**Figure 10B**). Second, we also tested whether FOXO1 controls *GDF15*. To do so, we transduced ECs with a Tet-On constitutively active FOXO1A3 expression construct (FOXO1^{CA}) (Potente, Urbich et al. 2005). RT-qPCR quantification of FOXO1 downstream genes *MX11*, *PDK1*, *PDK4*, *CD36* and *ANG2* confirmed activation of FOXO1 in ECs upon doxycycline treatment (**Figure 10C**). Furthermore, we found that FOXO1^{CA} lowered *GDF15* mRNA content in ECs (**Figure 10C**). As a result, FOXO1^{CA} also lowered the secretion of GDF15 into the cell culture medium (**Figure 10D**). We confirmed the regulation of *GDF15* gene expression in E4ORF1⁺ HUVECs (**Figure 10E**). The decrease in *GDF15* in those cells was dose-dependent since increasing doxycycline concentrations further lowered *GDF15* (**Extended data figure 2A**). Subsequently, we also assessed whether inhibition of FOXO1 in ECs would lead to an increased expression and release of GDF15. Inhibition of FOXO1 using the specific

inhibitor AS1842856 (Nagashima, Shigematsu et al. 2010) increased *GDF15* expression (**Figure 10F**) and increased the accumulation of GDF15 in cell culture media (**Figure 10G**). The altered secretion of GDF15 was not secondary to alterations in EC proliferation in response to FOXO1 modulation since treatment with the proliferation inhibitor mitomycin C also resulted into reduced GDF15 secretion upon FOXO1^{CA} versus increased GDF15 upon FOXO1 pharmacological inhibition (**Extended data figure 3A B**). Recently it has shown been shown that FOXO1 inhibition increases ATF4 expression in immune cells (Vallejo-Gracia, Chen et al. 2020). In turn, ATF4 has been reported to be a metabolic rheostat of nutritional stress that ultimately drives the expression of GDF15 in mice and humans (Patel, Alvarez-Guaita et al. 2019). Therefore, we tested whether GDF15 upregulation upon FOXO1 pharmacological inhibition is dependent on ATF4 and thus indirect. Despite that treatment with AS1842856 increased ATF4 expression as previously reported (**Extended data figure 2B**). Efficient knockdown of ATF4 did not blunt GDF15 upregulation upon FOXO1 blockade (**Extended data figure 2CD**). Thus, ECs control GDF15 secretion in a FOXO1 dependent manner and independent of ATF4.

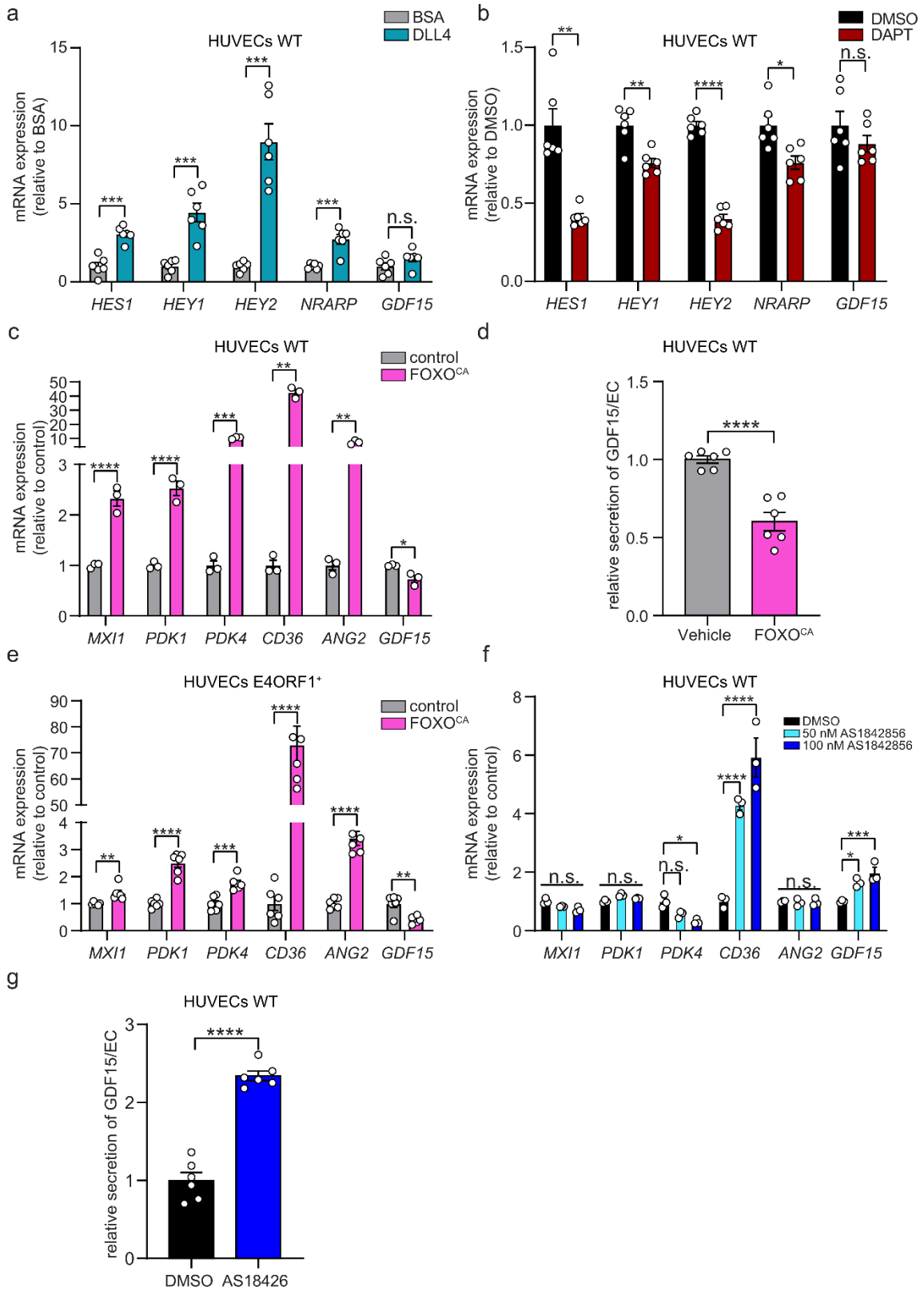


Figure 10 FOXO1 negatively regulates GDF15 expression in E4ORF1⁺ and WT HUVECs. **a.** Gene expression analysis of NOTCH1 target genes (*HES1*, *HEY1*, *HEY2* and *NRARP*) and *GDF15* upon stimulation with notch ligand, DLL4 or vehicle (0.02% BSA), over 24 h in HUVECs. DLL4 did not regulate *GDF15* expression (n= 6) (Student-t test). **b.** Gene expression analysis of NOTCH1 target genes (*HES1*, *HEY1*, *HEY2* and *NRARP*) and *GDF15* upon treatment with gamma-secretase inhibitor, DAPT 10 μ M, over 24 h in HUVECs (n= 6). DAPT did not regulate *GDF15* expression (Student-t test). **c.** Gene expression analysis of FOXO1 target genes (*MXI1*, *PDK1*, *PDK4*, *CD36*, *ANG2*) and *GDF15*, 3hrs after induction of FOXO1^{CA} with 200 ng/ml doxycycline in Tet-On FOXO1^{CA} HUVECs (n=3). (Student-t test) **d.** Secretion of GDF15 in cell culture supernatants 48h after induction of FOXO1^{CA} with 200 ng/ml doxycycline in Tet-On FOXO1^{CA} HUVECs (n=6) (Student-t test). **e.** Gene expression analysis of FOXO1 target genes (*MXI1*, *PDK1*, *PDK4*, *CD36*, *ANG2*) and *GDF15* 24 h after induction of FOXO1^{CA} with 200 ng/ml doxycycline in Tet-On FOXO1^{CA} E4ORF1⁺ HUVECs in serum free conditions (n=6) (Student-t test). **f.** Gene expression analysis of FOXO1 target genes (*MXI1*, *PDK1*, *PDK4*, *CD36*, *ANG2*) and *GDF15*, 24 hrs. after treatment with pharmacological FOXO1 inhibitor, AS1842856 in HUVECs (n=3) (Two-way ANOVA and Dunnet multiple comparison. **g.** Secretion of GDF15 in cell culture supernatants 48h after treatment with pharmacological FOXO1 inhibitor, AS1842856 in HUVECs (n=6) (Student-t test). Housekeeping gene *ACTB* was used as control for figure panels **a**, **b**, **c**, **e**, **f**. Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001

3.4.4. Endothelial deletion of FOXO1 leads to increased circulating GDF15

Since FOXO1 is active in a quiescent endothelium, the expression and release of GDF15 under homeostatic conditions is expected to be low. To study whether endothelial FOXO1 contributes to the regulation of circulating GDF15, we subsequently decided to use an inducible endothelial-specific FOXO1 knock-out mice *foxo1^{ΔEC}* which have been generated previously (Wilhelm, Happel et al. 2016)(**Figure 11A**). Three weeks after the tamoxifen treatment, whole tissue expression analysis of *GDF15* revealed a significant upregulation in skeletal muscle, white adipose tissue and heart. To elucidate EC contribution in those capillary beds, ECs from different tissue beds were isolated for analysis, including the kidney, this to understand whether this is a regional or rather a general regulation. Consistent with previous observations (Rudnicki, Abdifarkosh et al. 2018), we found that *foxo1* mRNA levels were efficiently downregulated in ECs isolated from skeletal muscle, white adipose tissue, kidney and heart (**Figure 11F H J L**). Reduction of *foxo1* coincided with an approximate 4-fold increase in *gdf15* expression in those ECs **Figure 11E G I K**). To test whether increased endothelial *gdf15* expression could affect circulating GDF15 levels, we analyzed serum as well as urine GDF15 which confirmed that circulating GDF15 levels were increased upon endothelial *foxo1* deletion (**Figure 11B**).

Since pharmacological GDF15 delivery, leading to higher increases in GDF15 levels, has been shown to reduce food intake (Emmerson, Wang et al. 2017, Mullican, Lin-Schmidt et al. 2017), we subsequently wondered whether small increments in circulating GDF15 are able to repress food intake in *foxo1^{ΔEC}* mice. Indeed, we found that endothelial deletion of *gdf15* sufficed to reduce food intake by approximately 22.7% (**Figure 11C**) without affecting body weight (**Figure 11M**) nor energy expenditure (**Figure 11N**)

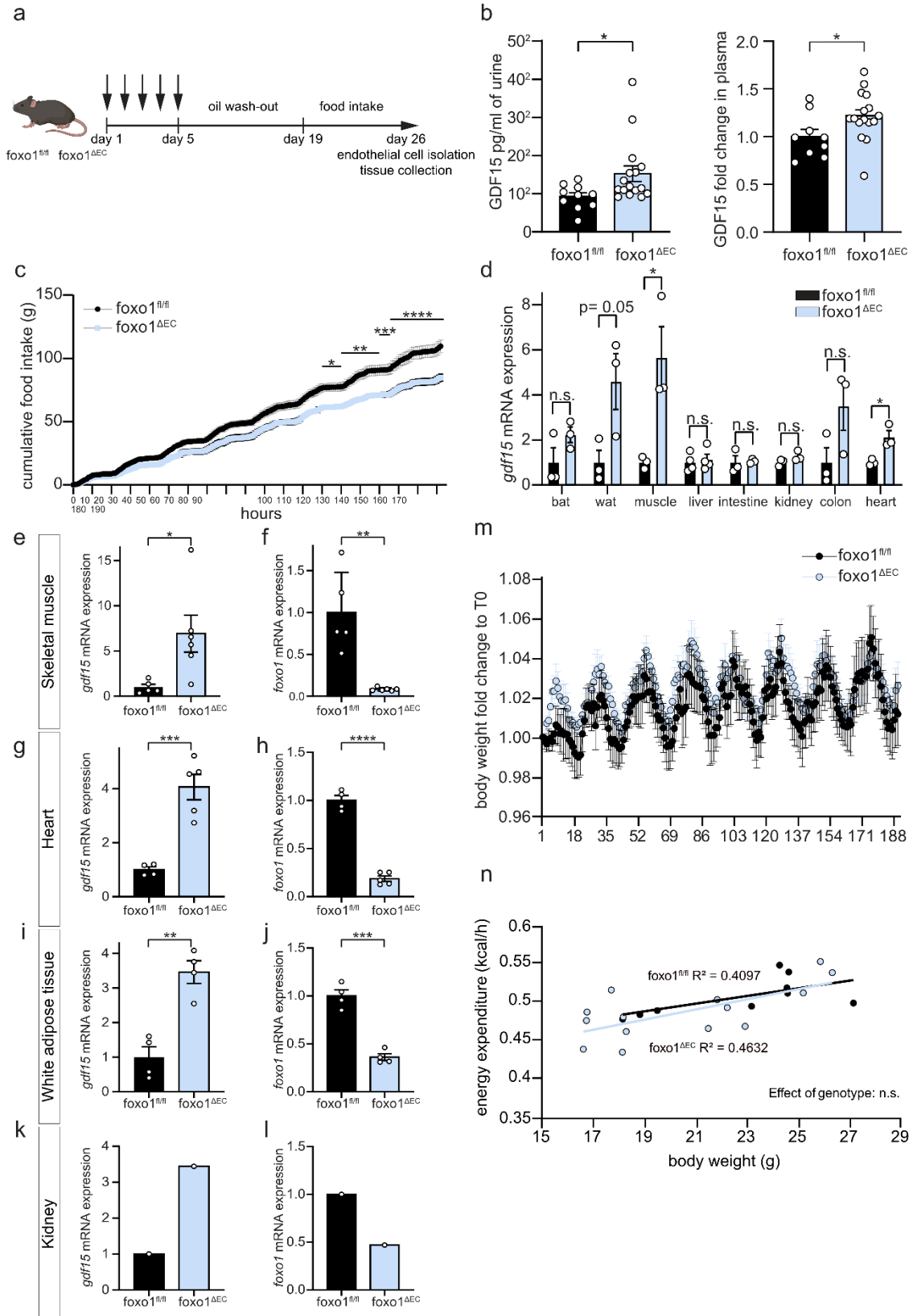


Figure 11 GDF15 is upregulated in $foxo1^{\Delta EC}$ mice and is associated with a decrease in food intake. **a.** Schematic of conditional deletion of FOXO1 in vascular endothelium, $foxo1^{fl/fl}$ mice were crossed with tamoxifen-inducible endothelial specific $pdgfb-CreERT2$ mice. Tamoxifen was administered once daily for 5 consecutive days. Tissues and/or endothelial cell isolation occurred at least 14 days after last tamoxifen dose. **b.** Urine concentrations of GDF15 in control $foxo1^{fl/fl}$ (n= 10) compared to $foxo1^{\Delta EC}$ mice (n= 16). Urine was collected at the beginning of dark cycle. Plasma concentrations of GDF15 in control $foxo1^{fl/fl}$ (n=9) compared to $foxo1^{\Delta EC}$ mice (n=16). (Student-*t* test). **c.** Cumulative food intake in chow fed of control $foxo1^{fl/fl}$ (n= 9) compared to $foxo1^{\Delta EC}$ mice (n= 14). Measurement was done 14 days after last tamoxifen administration and over 8 days (190 h)(Two-way ANOVA and Sidak's multiple comparison). **d.** *gdf15* mRNA expression was measured in whole tissue lysates in brown adipose tissue (BAT), white adipose tissue (WAT), skeletal muscle, liver, intestine, kidney, colon and heart in control $foxo1^{fl/fl}$ (n= 3) compared to $foxo1^{\Delta EC}$ mice (n= 3).(Student-*t* test). **e.** *gdf15* mRNA expression in freshly isolated endothelial cells from skeletal muscle in control $foxo1^{fl/fl}$ (n=5) compared to $foxo1^{\Delta EC}$ mice (n=6). (Student-*t* test) **f.** *foxo1* mRNA expression in freshly isolated endothelial cells from skeletal muscle in control $foxo1^{fl/fl}$ (n=5) compared to $foxo1^{\Delta EC}$ mice (n=6).(Student-*t* test) **g.** *gdf15* mRNA expression in freshly isolated endothelial cells from heart tissue in control $foxo1^{fl/fl}$ (n= 4) compared to $foxo1^{\Delta EC}$ mice (n= 5). (Student-*t* test) **h.** *foxo1* mRNA expression in freshly isolated endothelial cells from heart tissue in control $foxo1^{fl/fl}$ (n= 4) compared to $foxo1^{\Delta EC}$ mice (n= 5) (Student-*t* test) **i.** *gdf15* mRNA expression in freshly isolated endothelial cells from white adipose tissue in control $foxo1^{fl/fl}$ (n= 4) compared to $foxo1^{\Delta EC}$ mice (n= 4). (Student-*t* test) **j.** *foxo1* mRNA expression in freshly isolated endothelial cells from white adipose tissue in control $foxo1^{fl/fl}$ (n= 4) compared to $foxo1^{\Delta EC}$ mice (n= 4) (Student-*t* test). **k.** *gdf15* mRNA expression in freshly isolated endothelial cells from kidney tissue in control $foxo1^{fl/fl}$ (n= 1) compared to $foxo1^{\Delta EC}$ mice (n= 1). **l.** *foxo1* mRNA expression in freshly isolated endothelial cells from kidney tissue in control $foxo1^{fl/fl}$ (n= 3) compared to $foxo1^{\Delta EC}$ mice (n=3). Housekeeping gene 18S was used as control for figure panels from **d** to **l**. **m.** Body mass of control $foxo1^{fl/fl}$ (n= 9) compared to $foxo1^{\Delta EC}$ mice (n= 14). Measurement was done 14 days after last tamoxifen administration and over 8 days (190 h). (Two-way ANOVA and Sidak's multiple comparison) was done **n.** energy expenditure against body weight is shown for $foxo1^{\Delta EC}$ mice (n= 14) or controls (n= 9). (One way ANCOVA). Significant values are represented in asterisks as follows * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$.

3.5. Discussion.

In this paper, we performed unbiased proteomic characterization to identify angiocrine factors that might be involved in tissue metabolic control. To do so, we decided to analyze the secretome of HUVECs expressing the adenoviral protein E4ORF1, which enables EC growth and survival in the absence of serum, growth factors and cytokines (Seandel, Butler et al. 2008). Using this approach, we identified 227 secreted proteins, many of which have previously been validated to be secreted by endothelial cells. Moreover, there was significant overlap between our data and a previously published one where a labeled amino acid approach was used (Burghoff and Schrader 2011). The latter approach revealed 78 secreted proteins, more than half of them, including GDF15, also showed up in our analysis.

We found that ECs secrete significant amounts of GDF15 under proliferating conditions. Moreover, *GDF15* expression was negatively controlled by FOXO1, a known negative regulator of endothelial proliferation through the repression of c-Myc (Wilhelm, Happel et al. 2016). Activation of FOXO1 reduced GDF15 whereas inhibition of FOXO1 increased GDF15. This effect was not caused by changes in proliferation since treating HUVECs with mitomycin C did not affect FOXO1 mediated alterations in GDF15. Furthermore, we assessed the *in vivo* effect of FOXO1 deletion on GDF15 levels less than two weeks after inducing *foxo1* gene deletion. It is unlikely that significant endothelial proliferation occurred at that point. In fact, despite the critical role for FOXO1 in controlling endothelial proliferation during developmental angiogenesis, endothelial specific deletion of FOXO1 does not induce widespread endothelial proliferation (Rudnicki, Abdifarkosh et al. 2018), likely due to redundancy with other FOXO family members (Paik, Kollipara et al. 2007). An increase in vascular density in *foxo1*^{4EC} mice has only been observed in white adipose tissue 16 weeks after recombination. Thus, FOXO1-mediated regulation of GDF15 is proliferation-independent.

It is unclear how FOXO1 controls *gdf15* expression. Among other alternative regulatory pathways, the integrated stress response instructs the upregulation of GDF15 in response to nutritional stress via ATF4/CHOP signaling upon an obesogenic diet (Patel, Alvarez-Guaita et al. 2019). This cellular stress response is replicated upon metformin stimulation (Day, Ford et al. 2019, Coll, Chen et al. 2020). FOXO1 has also been recognized as nutrient sensor (Dong, Copps et al. 2008, Banks, Kim-Muller et al.

2011, Barbato, Tatulli et al. 2013), in which decreased transcriptional activity promotes energy balance and nutrient homeostasis, while increased activity drives maladjustments to nutrient intake and endocrine regulation. The fact that FOXO1 inhibition secretes a nutritional signal to drive energy balance extends the idea that secreted proteins and cytokines, such as GDF15, will continue driving these beneficial effects to whole body through paracrine secretion. We evaluated whether the activity of these aforementioned nutrient sensors were intertwined to mechanistically drive the expression of GDF15 in FOXO1-abrogated endothelial cells, as both, their synergistic interaction (Rached, Kode et al. 2010, Kode, Mosialou et al. 2012, Kode, Mosialou et al. 2012), and negative regulation has been previously described (Wang, Zhou et al. 2014, Chen, Gong et al. 2019, Ma, Su et al. 2020, Vallejo-Gracia, Chen et al. 2020). Despite that pharmacological blockade of FOXO1 upregulates the expression of ATF4, an instructing signal in integrated stress response, ISR, an ATF4 knockdown with a short hairpin did not blunt the upregulation of *GDF15* upon FOXO1 inhibition, which further sustains ATF4 independent mechanisms. Finally, whether the repressor activity is driven by previously described nuclear proteins remains to be elucidated (Nakae, Cao et al. 2012, Langlet, Haeusler et al. 2017). All in all, this evidence suggests that FOXO1 represses expression of GDF15 by directly binding to FOXO1 binding elements in GDF15 promoter and its interaction is independent of ATF4 activity/signaling.

To address the *in vivo* relevance of GDF15 secretion by ECs, we generated endothelial specific *gdf15* knock out mice. Removing *gdf15* from the endothelium however did not affect GDF15 levels. This is not surprising, since FOXO1 is active in a quiescent endothelium and thus should keep *gdf15* expression relatively low under physiological conditions. In contrast, deleting endothelial FOXO1 unleashed *gdf15* resulting into a more than 4-fold upregulation of its gene expression in ECs which sufficed to induce a modest but significant increase in circulating GDF15. Even more, the increase in GDF15 decreased food intake in these mice, underscoring the physiological relevance of our observations. There has been considerable debate about the physiological role of endogenous GDF15 on energy homeostasis in response to different stressors. For instance, vigorous endurance exercise leads to a 4-5 fold increase GDF15 without affecting food intake (Klein, Nicolaisen et al. 2021). On the other hand, similar increases in GDF15 induced by metformin treatment sufficed to reduce food intake and affect energy balance (Day, Ford et al. 2019, Coll, Chen et al. 2020). We found that

upon endothelial deletion, small changes in GDF15 suffice to affect food intake. Our genetic data suggest that even small differences in GDF15 production are enough to sustain a measurable amount of food aversion.

To the best of our knowledge, we could not find any reference discussing the potential contribution of the endothelium to circulating GDF15 levels. In many tissues, the endothelial volume only is a fraction of the total tissue volume. Nonetheless, the total mass of endothelial cells in an adult human body is significant and the localization of the endothelium, as the direct interface between nutrient delivery and nutrient consumption, render it perfectly suited to play a significant role in the regulation of energy balance. Future research needs to unravel the contribution of endothelial cells to whole body metabolic control and energy balance.

In conclusion, we found that endothelial cells are a significant source of GDF15, a main regulator of energy balance and food intake. While endothelial GDF15 does not contribute to circulating GDF15 under physiological conditions, deleting FOXO1 unleashes endothelial *gdf15* expression, leading to increased circulating GDF15 levels and reduced food intake. Our data suggest that endothelial cells can contribute to the control of energy balance.

3.6. Acknowledgments

We thank the Functional Genomics Center Zürich for technical support. This project was funded by the European Research Council (ERC) Starting Grant (716140) to KDB. AAD is supported by a PhD fellowship from the National Research and Technology Council of Mexico (CONACyT). KDB is endowed by the Schulthess Foundation.

3.7. Methods

Mice

Wildtype C57BL/6J mice were obtained from The Jackson Laboratory (000664 | B6). Endothelial specific GDF15^{ΔEC} mice were generated by crossing *gdf15^{loxP/loxP}* mice (obtained from Randy Seeley, University of Michigan, USA) with *pdgfb-Cre^{ERT2}* mice, an EC-selective tamoxifen inducible Cre-driver line (Claxton, Kostourou et al. 2008). *Foxo1^{ΔEC}* mice were generated by crossing *foxo1^{loxP/loxP}* mice (*Foxo1^{tm1Rdp/J}*), with *pdgfb-Cre^{ERT2}* mice. *Foxo1^{ΔEC}* and *gdf15^{ΔEC}* were used in C57BL/6 background. Recombination was induced in 8-12 weeks old mice by daily intraperitoneal administration of 1mg tamoxifen (T5648, Sigma-Aldrich) dissolved in 1:10 ethanol: corn oil solution for 5 consecutive days. A wash out period of at least 14 days was allowed before starting the experiments. Tamoxifen-treated Cre-negative littermates were used as control for all experiments. GDF15 blocking antibody and IgG control (against the non-mammalian protein ANTP) were a kind gift from Sebastian Beck Jørgensen (NovoNordisk A/S). Mice were injected intraperitoneally with 10 mg/ kg of body weight at the beginning of dark cycle every 7 days. Metformin (D150959, Sigma Aldrich) was administered diluted in water with a dosing of 600 mg/kg body weight via oral gavage. Thereafter, food was removed for 6h until blood sampling through a small cut in tail vein. Lipopolysaccharides (LPS - 10 mg/ kg of body weight) from *E. coli* O55:B5 (L6529, Sigma-Aldrich) administered intraperitoneally as described previously (Luan, Wang et al. 2019) and mice were euthanized 2 h later. Blood was sampled by cardiac puncture in heparinized tubes.

For metabolic measurements, *foxo1^{ΔEC}* and *gdf15^{ΔEC}* mice were acclimatized to single caging over 24h and put in metabolic cages thereafter (Promethion Cages, Sable Systems International). Food intake, water intake, spontaneous locomotor activity, oxygen (VO₂) consumption and carbon dioxide production (VCO₂), respiratory exchange ratio (RER) and energy expenditure were measured over 7 days.

Mice were randomly allocated to different treatment groups, and the investigator was blinded to the group allocation during the experiment as well as during the analysis. All mice were housed at standard housing conditions (22 °C, 12 h inverted light/dark cycle), with ad libitum access to chow diet (18 % proteins, 4.5 % fibers, 4.5 % fat, 6.3

% ashes, Provimi Kliba SA) and water. Health status of all mouse lines was regularly monitored according to FELASA guidelines. Animal experiments were approved by the local animal ethics committee (Kantonales Veterinärämtesamt Zürich, licenses ZH014/2016 and ZH211/2019), and performed according to local guidelines (TschV, Zurich) and the Swiss animal protection law (TschG). All mice used in this study were between 8-12 weeks old. Both male and female mice were included in the study.

Isolation of primary mouse endothelial cells (ECs)

Primary ECs from skeletal muscle, white adipose tissue, heart and kidney were isolated from adult *foxo1^{ΔEC}* mice as well as from *gdf15^{ΔEC}* and their respective WT littermates 14 days after the first tamoxifen injection. Mice were anesthetized using 117 mg/kg body weight ketamine and 13 mg/kg body weight xylazine. Skeletal muscle from both hind limbs, white adipose tissue, heart and kidney were dissected and put immediately on a dish at 4°C and finely minced with surgical scalpels. Thereafter, the mashed muscle was enzymatically digested in digestion buffer containing 2 mg/ml Dispase II (D4693, Sigma-Aldrich, Steinheim, Germany), 2 mg/ml Collagenase IV (17104019, Thermo Fisher Scientific, Zurich, Switzerland), 2 mM CaCl₂ and 2% BSA in PBS at 37°C for 20 min under continuous gentle shaking. The digestion reaction was stopped by mixing with an equal volume of cold Hank's Balanced Salt Solution (Thermo Fisher 14025-050) containing 10% FBS. The suspension was filtered through a 70 μm cell strainer (#431751, Corning, New York, USA) to remove large cell debris. Cells were centrifuged 500 RCF for 5 min at 4° C. Supernatant was subsequently discarded, while the cell pellets were resuspended in 1 ml hemolytic buffer (NH₄Cl 154 mM, KHCO₃ 10 mM and EDTA 0.1 mM pH 7.35) at room temperature for 3 minutes. To stop the reaction, 30 mL of ice-cold PBS was added to the mixture and centrifuged at 500 RCF for 5 min at 4°C. The cell pellets were resuspended in FACS buffer (PBS + 1% Bovine Serum Albumin) containing anti-CD31 PE conjugated antibody in a concentration of 1:200 (BD Pharmigen 553373) and anti-CD45 PerCP conjugated antibody in a concentration of 1:200 (BD Pharmigen 557235) for 30 min at 4° C, protected from light. Thereafter the cells were washed with 1ml of FACS buffer, centrifuged in a tabletop centrifuge (CAT number) at 500 RCF at 4° C for 5 min. Finally, the cell pellets were resuspend in 1 ml of FACS buffer (PBS + 1% Bovine Serum Albumin) and the suspension was passed through a 35 μm cell strainer of a FACS

sorting tube (352235, Corning). Immediately before sorting, SYTOX™ blue was added in 1:1000 (Thermo Fisher, S34857) to exclude dead cells from further analysis. Viable endothelial cells (CD31⁺, CD45⁻, SYTOX™ blue⁻) were sorted by a FACS Aria III (BD Bioscience) sorter directly into RLT plus RNeasy plus lysis buffer and RNA was extracted using RNeasy Plus Micro Kit (Cat No. 74034, Qiagen, Hilden, Germany). The FACS plots were analyzed with Flowjo (version 10.4.2).

Cell culture.

Human umbilical vein endothelial cells (HUVECs) from pooled donors (C-12203, PromoCell, Heidelberg, Germany) were cultured in M199 ((11150059, Thermo Fisher Scientific) supplemented with 20% fetal bovine serum (FBS) (10270-106, Thermo Fisher Scientific), 30 mg/L endothelial cell growth factor supplements (EGCS) (E2759, Sigma-Aldrich), 10 U/mL heparin (H3149 Sigma-Aldrich) and 1% Penicillin-Streptomycin (10,000 U/mL) (15140122, Thermo Fisher Scientific). Cells were routinely maintained in 5% CO₂ and 95% air at 37 °C and medium was changed every 48 h. HUVECs overexpressing E4ORF1 and E4ORF1 lentiviral particles (Seandel, Butler et al. 2008) were a kind donation of Prof. Shahin Rafii (Weill Cornell School of Medical Science, New York).

***In vitro* analysis**

For DLL4 stimulation, culture plates were coated with 1 µg/mL recombinant human Delta-like ligand 4 (rhDLL4, cat.1506-D4 R&D Systems) in 0.1% gelatin/PBS. The control plates were coated with 0.1% gelatin supplemented with 0.02% BSA. Prior to EC seeding, excess coating solution was removed by aspiration and ECs were seeded at a density of 30,000 cells/cm². Cells were harvested 24 h after seeding. For NOTCH inhibition, cells were treated with the γ -secretase inhibitor N-[N-(3,5-Difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester (DAPT; 10 µM, 5657702, Calbiochem) for 24 h. For proliferation blocking, Mitomycin C (cat 4150. Carl Roth) was added at a final concentration of 1µg/ml in cell culture media for 72 h. Subsequently, cell culture supernatants were taken for GDF15 quantification and cells were stained with Hoechst 33342 (H3570, Invitrogen) to allow normalization for cell numbers using a plate reader (Tecan, Spark). The FOXO1 inhibitor AS1842856 (4265, TOCRIS) was

resuspended in DMSO at a concentration of 50 μ M and kept -20°C. For FOXO1 inhibition, readily attached HUVECs were treated with either 50 or 100 nM for the indicated duration. An equivalent volume of DMSO was used as control.

Sample preparation for proteomic analysis

Conditioned media was generated from E4ORF1 HUVECs seeded at a cell density of 15,000 cells/cm², grown in serum-free M199 (2mM L-glutamine, 30 mg/l endothelial cell growth factor supplements (Sigma E2759), 10 units/ml heparin (Sigma H3149), 50 IU/ml penicillin and 50 μ g/ml) for 48 hours. The media were collected and centrifuged for 10 min at 500 RCF to precipitate cell debris. To facilitate volume handling during protein precipitation and further LC-MS/MS analysis, the conditioned media was subsequently concentrated using Amicon Ultra filters with a molecular weight cut-off value of 3 kDa (Merck-Millipore UFC900296). Proteins in the conditioned media were then precipitated using the chloroform methanol method according to Wessel and Flügge (Wessel and Flugge 1984) to prepare for label free quantification proteomic analysis. Briefly, a mixture of conditioned media, methanol, chloroform, and distilled water was prepared in a ratio of 1:4:1:3, respectively. The mixture was thoroughly vortexed and centrifuged for 10 min, 5,000 RCF at 4°C. The aqueous (upper) phase was removed and the remaining interphase/organic phase was further mixed with 3 parts of methanol. After centrifugation, the supernatant was removed and the resulting pellet was vacuum (5305000304, Eppendorf) dried for 5 min. Dried protein pellets were kept at -80 °C until further processing.

Knock down and overexpression plasmid constructions and lentiviral particle production.

Lentiviral particles were generated by transfection of HEK 293 cells (Cat.# ACC635; EGF, Braunschweig, Germany) with Pmd2 (AddGene, Plasmid #12259), lentiviral envelope plasmid psPAX2 (AddGene, Plasmid #12260) and the plasmids. The pLVX-TetOn-Puro-FLAG-FOXO1A3 (FOXO1^{CA}) plasmid was a kind gift of Michael Potente (Max Planck Institute for Heart and Lung Research, Bad Nauheim, Germany). Additionally, GIPZ lentiviral shRNA targeting *atf4* (V2LHS272113; V3LHS302002; V3LHS302003) were purchased from Dharmacon (Horizon Discovery; Waterbeach,

United Kingdom). A nonsense scrambled shRNA sequence was used as control. Lentiviral particles were generated by transfection of HEK 293 cells with the respective plasmid and pLenti-C-mGFP- PEP-Puro Lentiviral Gene Expression Vector (Cat. #PS100093, Origene). Lipofectamine 2000 (Cat.# 11668030; Thermo Fisher Scientific) was used for transfection. Viral particles were collected at least 48 hours after incubation. Subsequently, HUVECs were transduced with particle containing supernatant (passed through a 0.45 µm filter) for 72 hours in the presence of 8 µg/ml polybrene and re-fed with fresh medium the next day. Transduced HUVECs were subsequently selected with 2 µg/ml of puromycin over 3 days. Puromycin containing medium was changed every 24 h. To induce expression of pLVX-TetOn-Puro-FLAG-FOXO1A3 (FOXO1^{CA}), transfected endothelial cells were treated with the noted concentration of doxycycline for 24 h E4ORF1 HUVECs and 3 h in wild type HUVECs. To generate E4ORF1 expressing HUVECs, we transduced a 30% confluent T75 flask with 15 µl of E4ORF1 lentiviral particles (Seandel, Butler et al. 2008) reaching an approximate MOI of 3. After 48 hours, the medium was removed and cells were selected by culturing them in serum deprived medium for 4 days.

GDF15 enzyme-linked immunosorbent assay ELISA

For determination of GDF15, urine was acquired at the beginning of dark cycle by placing a plastic film under an animal while it was being restrained. Urine (approximately 10 µl) was taken and diluted in 490 µl of calibrator diluent. Blood samples were taken from the tail vein according to approved ethical procedures. Blood was acquired in heparinized tubes (Microvette CB 300 K2E) which were centrifuged at 4 °C for 20 min at 2000 RCF to acquire the plasma. If ELISA was not carried out immediately after, plasma was stored at -80 °C. Cell culture supernatants were centrifuged for 10 min at 500 RCF to eliminate cell debris. 50 µl of diluted urine, serum or cell culture supernatants were used to quantify GDF15 concentrations according to manufacturer instructions (R&D systems, Quantikine Mouse/Rat GDF15 ELISA, MGD150).

RNA extraction and quantitative RT-PCR

RNA of directly FACS sorted mEC was extracted using an RNeasy Plus Micro Kit according to the manufacturer's instructions (QIAGEN, 74034). RNA of cultured HUVECs was extracted using PureLink™ RNA Mini Kit (12183020, Thermo Fischer Scientific). RNA purity and concentration were assed via a spectrophotometer (Tecan, Spark). RNA was reverse-transcribed to cDNA by High Capacity cDNA Reverse Transcription Kit (Thermo Fisher, 43-688-13). A SYBR Green-based master mix (ThermoFisher Scientific, A25778) was used for real-time qPCR analysis with primers listed in Table 1. To compensate for variations in RNA input and efficiency of reverse-transcription, 18S was used as a housekeeping gene for primary isolated EC and Actin Beta for HUVECs. The delta-delta CT method was used to normalize the data.

Proteomic analysis.

Sample preparation

Samples were prepared by using a commercial iST Kit (PreOmics, Germany) with an updated version of the protocol. Briefly, 50 ug of the samples were solubilized in 'Lyse' buffer, boiled at 95°C for 10 minutes and processed with High Intensity Focused Ultrasound (HIFU) for 30s setting the ultrasonic amplitude to 85%. Then the samples were transferred to the cartridge and digested by adding 50ul of the 'Digest' solution. After 60min of incubation at 37°C the digestion was stopped with 100ul of Stop solution. The solutions in the cartridge were removed by centrifugation at 3800g, while the peptides were retained by the iST-filter. Finally the peptides were washed, eluted, dried and re-solubilized in 20ul 'LC-Load' buffer for MS-Analysis.

Liquid chromatography-mass spectrometry analysis

Mass spectrometry analysis was performed on an Orbitrap Fusion Lumos (Thermo Scientific) equipped with a Digital PicoView source (New Objective) and coupled to a M-Class UPLC (Waters). Solvent composition at the two channels was 0.1% formic acid for channel A and 0.1% formic acid, 99.9% acetonitrile for channel B. For each sample 2 µL of peptides were loaded on a commercial MZ Symmetry C18 Trap Column (100Å, 5 µm, 180 µm x 20 mm, Waters) followed by nanoEase MZ C18 HSS T3 Column (100Å, 1.8 µm, 75 µm x 250 mm, Waters). The peptides were eluted at a flow rate of 300 nL/min by a gradient from 5 to 22% B in 80 min, 32% B in 10 min and 95% B in 1 min. Samples were acquired in a randomized order. The mass spectrometer

was operated in data-dependent mode (DDA) acquiring a full-scan MS spectra (300–1'500 m/z) at a resolution of 120'000 at 200 m/z after accumulation to a target value of 500'000. Data-dependent MS/MS were recorded in the linear ion trap using quadrupole isolation with a window of 0.8 Da and HCD fragmentation with 35% fragmentation energy. The ion trap was operated in rapid scan mode with a target value of 10'000 and a maximum injection time of 50 ms. Only precursors with intensity above 5'000 were selected for MS/MS and the maximum cycle time was set to 3 s. Charge state screening was enabled. Singly, unassigned, and charge states higher than seven were rejected. Precursor masses previously selected for MS/MS measurement were excluded from further selection for 30 s, and the exclusion window was set at 10 ppm. The samples were acquired using internal lock mass calibration on m/z 371.1012 and 445.1200. The mass spectrometry proteomics data were handled using the local laboratory information management system (LIMS) (Türker, Akal et al. 2010).

Protein identification and label free protein quantification

The acquired raw MS data were processed by MaxQuant (version 1.6.2.3), followed by protein identification using the integrated Andromeda search engine (Cox and Mann 2008). Spectra were searched against a Swissprot Homo sapiens reference proteome (taxonomy 9606, version from 2019-07-09), concatenated to its reversed decoyed fasta database and common protein contaminants. Carbamidomethylation of cysteine was set as fixed modification, while methionine oxidation and N-terminal protein acetylation were set as variable. Enzyme specificity was set to trypsin/P allowing a minimal peptide length of 7 amino acids and a maximum of two missed-cleavages. MaxQuant Orbitrap default search settings were used. The maximum false discovery rate (FDR) was set to 0.01 for peptides and 0.05 for proteins. Label free quantification was enabled and a 2 minutes window for match between runs was applied. In the MaxQuant experimental design template, each file is kept separate in the experimental design to obtain individual quantitative values. Protein fold changes were computed based on Intensity values reported in the proteinGroups.txt file. A set of functions implemented in the R package SRMSservice (Wolski, Grossmann et al. 2018) was used to filter for proteins with 2 or more peptides allowing for a maximum of 4 missing values, and to normalize the data with a modified robust z-score transformation and to compute

p-values using the t-test with pooled variance. If all measurements of a protein are missing in one of the conditions, a pseudo fold change was computed replacing the missing group average by the mean of 10% smallest protein intensities in that condition.

Figures and diagrams

Figures in this manuscript were created with BioRender.com. or R studio.

Quantification and statistical analysis

All data represent mean \pm SEM. GraphPad Prism software (version 8.0.0) was used for statistical analyses. Investigators were always blinded to group allocation. Unless otherwise indicated, when comparing two group means, Student's *t*-test was used in an unpaired two-tailed fashion. For more than two groups, one-way ANOVA with Tukey's multiple comparisons test was used and for experimental set-ups with a second variable, two-way ANOVA with Sidak's multiple comparisons test was used. The statistical method used for each experiment is indicated in each figure legend. Asterisks in figure legends denote statistical significance. No experiment-wide multiple test correction was applied. $P > 0.05$ is considered non-significant (ns). $P < 0.05$ is considered significant (*).

Table 1. Sequences of primers used for RT-PCR

gene	Forward	Reverse
<i>18s</i> (m.& h.)	AGTCCCTGCCCTTTGTACACA	CGATCCGAGGGCCTCACTA
<i>ATF4</i> (h.)	GTTCTCCAGCGACAAGGCTA	ATCCTGCTTGCTGTTGTTGG
<i>BIP</i> (h.)	TGTTCAACCAATTATCAGCAAACCT	TTCTGCTGTATCCTCTTCACCACT
<i>CHOP</i> (h.)	AGAACCAGGAAACGGAAACAGA	TCTCCTTCATGCGCTGCTTT
<i>HEY1</i> (h.)	TGGATCACCTGAAAATGCTGC	CGAAATCCCAAACCTCCGATAGT
<i>HEY2</i> (h.)	TGGGGAGCGAGAACAATTAC	TCAAAGCAGTTGGCACAAG
<i>HES1</i> (h.)	TGAAGAAAGATAGCTCGCGGC	GGTACTTCCCCAGCACACTT
<i>NRARP</i> (h.)	CGCTGTTGCTGGTGTCTAAA	CATTGACCACGCAGTGTTTTTC
<i>foxo1</i> (m.)	AGTTAGTGAGCAGGCTACATTT	TTGGACTGCTCCTCAGTTCC
<i>CD36</i> (h)	GCTGTTGATTTGTGAATAAGAACC	GCACCTGTTTCTTGCAAACCTCC
<i>MXI1</i> (h.)	GCCAAAGCACACATCAAGAAACT	GCTGTTCCAGTCGCCACTTT
<i>PDK1</i> (h.)	TCTCAGGACACCATCCGTTCA	ACCATGTTCTTCTAGGCCTTTCAT
<i>ANG2</i> (h.)	TGCCACGGTGAATAATTCAG	TTCTTCTTTAGCAACAGTGGG
<i>PDK4</i> (h.)	GTAGCAGTGGTCCAAGATGCC	ACACGATGTGAATTGGTTGGTCT
<i>GDF15</i> (h.)	ATACTCACGCCAGAAGTGCGG	GAACAGAGCCCGGTGAAGGC
<i>gdf15</i> (m.)	GAGCCGAGAGGACTCGAACT	CCCCAATCTCACCTCTGGACT
<i>ACTB</i> (m. h.)	GCTCCTCCTGAGCGCAAG	CATCTGCTGGAAGGTGGACA

4. Angiocrine GDF15 improves glucose homeostasis by direct effects on insulin sensitivity in skeletal muscle and is independent of weight loss.

Running title: GDF15 is a *bonafide* insulin sensitizer.

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4.1. Abstract.

Endothelial cells form the inner lining of blood vessels. Yet, the function of blood vessels lies beyond simple conduits that exchange oxygen and nutrients between blood and the tissues they irrigate. It has been recently suggested that endothelial cells can control the function of metabolic organs and, with this, whole energy homeostasis. Indeed, it has become increasingly clear that endothelial cell dysfunction precedes metabolic disease and diabetes type II. On the other hand, increasing their responsiveness to growth factors, metabolic adaptation to nutrient insults, nitric oxide (NO) and reactive oxygen species signaling (ROS) improves glucose handling and insulin sensitivity. Along with these evidences, in this study we hypothesized that endothelial cells are capable of sensitizing skeletal muscle to the effect of insulin via the secretion of soluble factors. Furthermore, in a previous study we identified GDF15 to be abundantly secreted by endothelial cells *in vivo*. Therefore, we explored whether GDF15 could be a *bonafide* insulin sensitizer by direct actions to skeletal muscle and whole body glucose homeostasis. First, *in vitro*, we found that endothelial cell conditioned media increases insulin sensitivity in human myotubes and that GDF15 plays a significant role in this observation. Furthermore, GDF15 alone increases insulin sensitivity shown by both, insulin signaling and GLUT4 translocation in myotubes. Finally, *in vivo*, mice that were administrated GDF15 display an improved glucose handling in an insulin tolerance test (ITT), increased glucose uptake, and insulin signaling in skeletal muscle. Importantly, these effects were found to be acute and independent of well-established anorectic effects on weight loss.

4.2. Authors contributions

The general content, perspective and supervision of this chapter was proposed by Prof. Dr. Katrien De Bock. The design, execution, and analysis of all the experiments, writing the full manuscript content, and creation of all figures was carried out by Abdiel Alvarado Diaz. Over the course of this project, several contributors have significantly supported its realization in forms of assistance during the execution and/or analysis of some experiments or figures. Dr. Fateme Jaleh supported the design and execution of experiment associated with figures 12, 13 and 14. She also gave valuable input to the design of manuscript. Finally, she helped me solve fundamental technical issues assessing *in vivo* glucose homeostasis experiments. Dr. Gommaar D'Hulst technically supported the execution of experiments associated to figure 13. Moheb Ghobrial assisted with significant input regarding the design of figures 12, 13, 14. Dr. Paola Gilardoni provided technical assistance in the conduct of all experiments. The general supervision and correction of this manuscript was done by Abdiel Alvarado Diaz and Prof. Dr. Katrien De Bock.

4.3. Introduction.

By mass, skeletal muscle harbors the largest amount of vascular endothelial cells (Egginton 2011). The skeletal muscle vasculature consists of an intricate meshwork of endothelial cells in charge of sustaining the high metabolic demands of oxygen and nutrients while enduring the high mechanical stress of its surrounding tissue (Olfert and Birot 2011). However, the importance of adequate vascular function and structure lies beyond this primary function as the balance between perfusion and metabolic demands also needs to match the emerging properties of vascular endothelium. Among these emerging regulatory roles of these muscle-associated vascular endothelial cells, we find that ECs can modulate skeletal muscle regeneration. ECs reside in close proximity to satellite cells (Pax7⁺ and Myf5⁺ expressing muscle progenitor cells) (Christov, Chrétien et al. 2007), this close association suggests that crosstalk may occur via direct or indirect mechanisms, this is, through cell-to-cell contact or through paracrine communication via the secretion of angiocrine factors into interstitial space of musculoskeletal tissue (Verma, Asakura et al. 2018). ECs control skeletal muscle stem cells and its progenitor clonal expansion and differentiation through the secretion of angiocrine IGF1, HGF, FGF-2, VEGFA and homodimers of platelet-derived growth factor β (Arsic, Zacchigna et al. 2004, Bryan, Walshe et al. 2008, Borselli, Storrie et al. 2010). Interestingly, vascular endothelial cells have also been demonstrated to undergo myogenic differentiation *in vivo* (Zheng, Cao et al. 2007), which suggest that skeletal muscle cells and its supplying vasculature, in permissive conditions, have overlapping physiological properties. Furthermore, a reciprocal synergistical contribution between myogenesis and angiogenesis has been previously reported to occur through the secretion of soluble factors *in vitro* (Osaki, Sivathanu et al. 2018). However, whether EC can modulate the other aspects of skeletal muscle biology such as metabolism has not been addressed in proper depth.

There are preliminary indications that indeed EC can control skeletal muscle metabolism, specifically in the context of insulin-mediated control of glucose homeostasis. It has been shown that a dampened insulin signaling in EC leads to a decreased glucose uptake in skeletal muscle (Kubota, Kubota et al. 2011). Additionally, caveolin-1, insulin-receptor and insulin-like growth factor receptor-1 have been identified to be important players in the transendothelial transport of insulin

towards the skeletal muscle (Wang, Liu et al. 2006). This via formation of caveolae, which are membrane invaginations involved in the transport of large macromolecules to interstitial compartment (Schubert, Frank et al. 2001). Furthermore, during exercise, skeletal muscle goes from a basal insulin sensitivity to high insulin sensitivity (Richter, Garetto et al. 1982, Reynolds IV, Brozinick Jr et al. 2000). However, before these metabolic changes happen, a significant increase of skeletal muscle capillarity occurs (Waters, Rotevatn et al. 2004). Furthermore, it has been shown that upon VEGFB stimulation an increase of capillarity in adipose tissue leads to improved glucose metabolism, for which an improved insulin supply is shown to be partially responsible; these changes are also accompanied by weight loss in mice under obesogenic diet (Robciuc, Kivelä et al. 2016). Additionally, it has been previously suggested that endothelial cells present in the skeletal muscle are the primary source for NO and ROS regulation of glucose homeostasis (Sansbury and Hill 2014, Paneni, Costantino et al. 2015, Watt, Gage et al. 2017). Yet, the regulation of skeletal muscle glucose homeostasis by direct secretion of angiocrine factors is not yet established, despite growing evidence that crosstalk between endothelial cells and other metabolic organs plays a fundamental role in metabolic organs homeostasis and ultimately whole energy control (Nolan, Ginsberg et al. 2013, Robciuc, Kivelä et al. 2016, Potente and Mäkinen 2017).

Under the light that endothelial cells control tissue homeostasis through the secretion of angiocrine factors, the closely related physical and physiological relationship between endothelial cells and skeletal muscle cells and the importance of endothelial cells in muscle adaptations exercise, the aim of the present work is to identify whether soluble factors secreted by endothelial cells improve insulin sensitivity and glucose homeostasis in skeletal muscle. Previously, we identified GDF15 to be abundantly secreted by endothelial cells. GDF15 is a distant member of the transforming growth factor B family (TGF- β), which has been shown to modulate food intake via interaction with its receptor, GFRAL, in the area postrema of the hindbrain (Emmerson, Wang et al. 2017, Hsu, Crawley et al. 2017, Mullican, Lin-Schmidt et al. 2017, Yang, Chang et al. 2017). Besides the well-established central nervous system (CNS) anorectic effects in food intake and the consequential metabolic benefits of a reducing adipose mass in the body such as improved insulin sensitivity and overall glucose homeostasis, it has

been suggested that GDF15 could also act locally to reduce cytokine-induced β -cell apoptosis and resolve endoplasmic reticulum (ER) stress in diabetes type I model of human pancreata (Nakayasu, Syed et al. 2020) or to modulate the macrophage metabolism to drive systemic insulin actions (Jung, Choi et al. 2018). Altogether this suggest that GDF15 may have a rather pleiotropic influence in the control of glucose homeostasis in mammals. The identification of secreted GDF15 in culturing media from EC, led us to hypothesize that GDF15 could be a potential angiocrine factor that can be considered as *bonafide* insulin sensitizer to the skeletal muscle and thus improving whole body glucose homeostasis.

4.4. Results

4.4.1. Endothelial cell conditioned medium sensitizes human myotubes to insulin.

Recent studies have pointed out that improving endothelial function has a positive impact in insulin sensitivity and glucose homeostasis (Robciuc, Kivelä et al. 2016) . Yet, mechanistic approaches are largely lacking. To explore the hypothesis that ECs control muscle insulin sensitivity via secretion of soluble factors, also known as angiocrine factors, rather than cell-to-cell contact, we stimulated human myotubes with endothelial cell conditioned medium (CM), and then assessed their insulin responsiveness. We first generated CM from HUVECs in standard culturing conditions, this is, with medium supplemented with fetal bovine serum and endothelial growth factors. As non-conditioned (NC) control, we used fresh media incubated at 5% of CO₂ in without cells for the same amount of time. Fully differentiated human myotubes (HSkM) were stimulated in a CM or NC / differentiation medium ratio of 3:7 for 48 h (**Figure 12A**). Subsequently, myotubes were stimulated with 100 nM of insulin, and tissues were harvested 30 min later. We observed that myotubes stimulated with endothelial CM, but NC control, upregulated targets of the insulin signaling cascade (**Figure 12B**) such as increased phosphorylation in AS160 T642 (**Figure 12C**), AKT T308 (**Figure 12D**) and AKT S473 (**Figure 12E**).

However, some of the limitations of addition of fetal serum in standard culturing conditions is the presence of overt amounts of growth factors and cytokines that are exogenous to ones produced by ECs (Hannoun, Fletcher et al. 2010, Jeon, Lim et al. 2010, Karnieli, Friedner et al. 2017). To exclude a potential interference from the serum-associated cytokines and growth factors in media, we subsequently used E4ORF1⁺ ECs, which were a kind donation from Prof. Shahin Raffi (Butler, Nolan et al. 2010). E4ORF1 is an adenoviral protein that keeps AKT constitutively active in cells expressing it, thus ensuring survival in the absence of serum, which allow for a long term culture. CM was produced from E4ORF1⁺ HUVECs serum-free , again a ratio 3:7 in differentiation media was applied on human myotubes, as shown before (**Figure 12F**).CM stimulation in serum-free conditions also proved to increase insulin signaling (**Figure 12G**) as shown by increased phosphorylation of AS160 T642 (**Figure 12H**), AKT T308 (**Figure 12I**) and AKT S473 (**Figure 12J**) compared to controls.

Finally, we excluded that increased insulin sensitivity responses in myotubes stimulated with CM compared to control are caused by differences in relative concentration of nutrients. To do so, we used serum-free conditioned medium taken from E4ORF1⁺ HUVECs and concentrated it 40x by filtering it through a filter with cut-off value of 3 kDa, which means that only macromolecules with a molecular weight value superior of 3kDa were taken to stimulate human myotubes with an approximate ratio of 1:9 in skeletal muscle differentiation media (**Figure 12K**). Concentrated CM provoked an increased in insulin cascade (**Figure 12L**) as shown by increased phosphorylation of AS160 T642 (**Figure 12M**), AKT T308 (**Figure 12N**), AKT S473 (**Figure 12O**), ERK 1/2 p42/44 (**Figure 12P**). This upregulation of insulin sensitivity was not caused by energy stress as assessed by phosphorylation of AMPK T172 (**Figure 12Q**). Furthermore, increased insulin sensitivity in human myotubes was accompanied by enhanced glycolytic flux (**Figure 12R**) and insulin dependent glucose translocation of glucose transporter 4 (GLUT4) (**Figure 12S**)

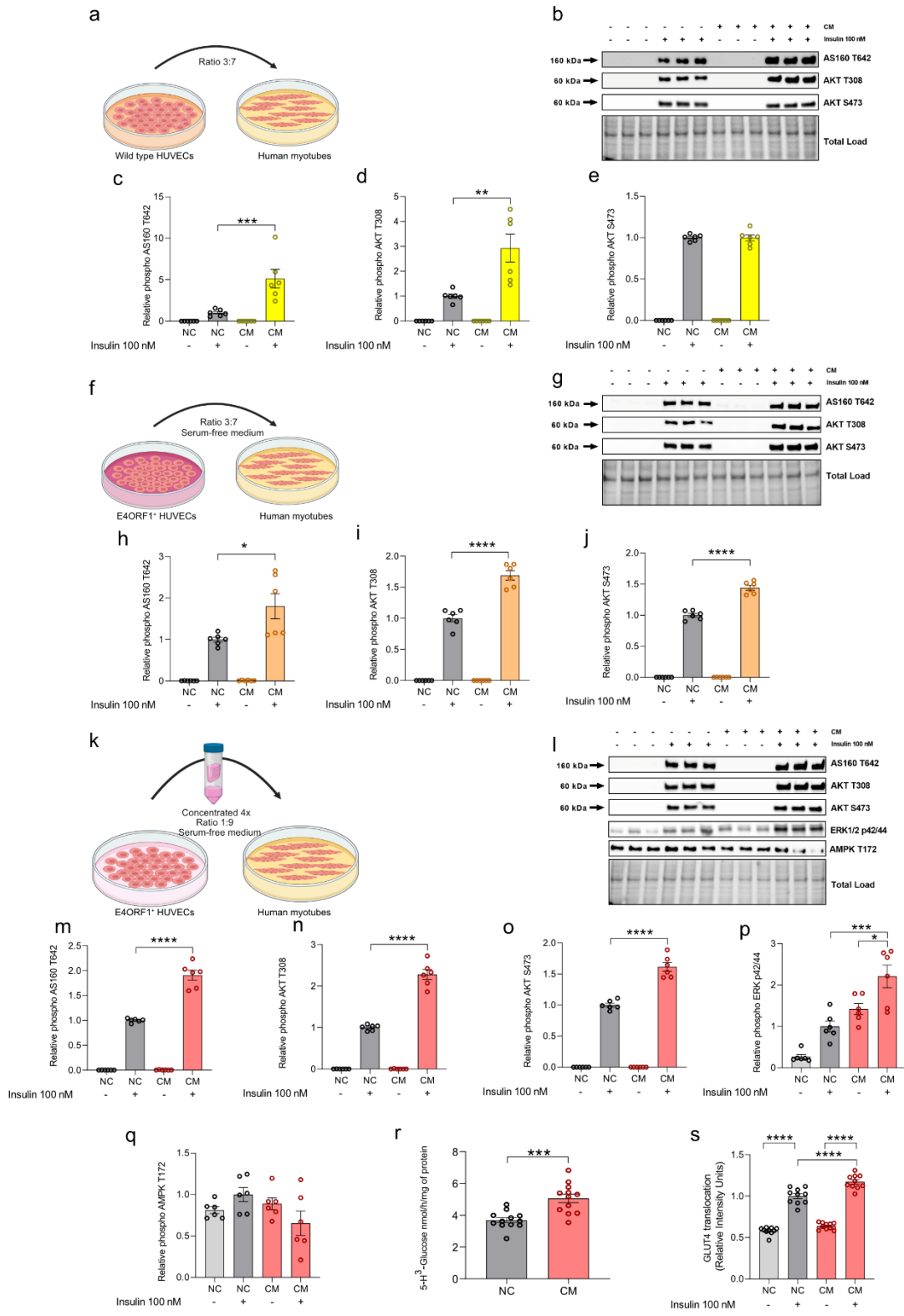


Figure 12 Endothelial cell secretome sensitizes skeletal muscle to the effects of insulin in vitro. **a.** Schematic of conditioned media generation from wild type HUVECs in the presence of serum and applied in a 3:7 ratio to human myotubes. **b.** Representative blot of insulin signaling from human myotubes that have been stimulated with CM or NC from WT HUVECs over 48 h. **c.** CM treated myotubes increase responsiveness to insulin assessed by phosphorylation of AS160 T642 (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **d.** CM treated myotubes increase responsiveness to insulin assessed by phosphorylation of AKT T308 (n= 6). **e.** CM treated myotubes does not respond differently to insulin when compared to control assessed by phosphorylation of AKT S473. (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **f.** Schematic of CM generation from E4ORF1+ HUVECs in the absence of serum and applied in a 3:7 ratio to human myotubes. **g.** Representative blot of insulin signaling from human myotubes that have been stimulated with CM or NC from E4ORF1+ HUVECs over 48 h. **h.** CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AS160 T642 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **i.** CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AKT T308 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **j.** CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AKT S473 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **k.** Schematic of 40x concentrated CM generation from E4ORF1+ HUVECs in the absence of serum and applied in a 1:10 ratio to human myotubes. **l.** Representative blot of insulin signaling from human myotubes that have been stimulated with 4x CM or NC from E4ORF1+ HUVECs over 48 h. **m.** 4x concentrated CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AS160 T642 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **n.** 4x concentrated CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AKT T308 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **o.** 4x concentrated CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of AKT S473 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **p.** 4x concentrated CM generated in serum-free conditions increase responsiveness to insulin assessed by phosphorylation of ERK1/2 p42/44 in human myotubes (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **q.** Insulin sensitizing effects of conditioned media are not mediated by differences in energy stress as assessed by AMPK T172 in human myotubes stimulates with 4x concentrated CM produced in serum-free conditions (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **r.** Human myotubes treated with 4x concentrated CM generated in serum-free conditions have increased glycolytic flux (n= 12) (Student-t test). **s.** Rat L6 myotubes treated with 4x concentrated CM generated in serum-free conditions display a greater GLUT4 translocation upon insulin stimulation (n= 10) (Two-way ANOVA and Tukey's multiple comparison). In figures **b**, **g** and **l**, the total load shows comparable amounts of protein loaded in the representative blot. Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001.

4.4.2. GDF15 significantly contributes to insulin sensitizing effects of conditioned media

From a previous study where we analyzed endothelial secretome *in vitro*, we identified GDF15 to be highly abundant and upregulated in endothelial cell secretome. GDF15 is a distant member of the TGF- β family. Upon binding to its receptor, GFRAL, in the area postrema of the hind brain, it mediates aversion to food intake, and thus ultimately leads to reduction of body mass (Emmerson, Wang et al. 2017, Hsu, Crawley et al. 2017, Mullican, Lin-Schmidt et al. 2017, Yang, Chang et al. 2017). . Despite that it has been suggested that GDF15 acts locally in peripheral tissue, no receptor outside CNS has been identified so far. Along with findings that GDF15 has a holistic influence on glucose homeostasis, we evaluated whether its able to synergistically act with insulin. To assess whether angiocrine GDF15 plays a significant role in the insulin sensitizing effects of endothelial CM, GDF15 was knocked down in E4ORF1 ECs, and CM was produced culturing these cells in in serum free conditions over 48 h as shown previously. with an efficiency of 78% (**Figure 13A**). CM from α -GDF15 E4ORF1 failed to increase insulin sensitivity as compared to sh-scrambled control (**Figure 13B**) as phosphorylated AKT308 (**Figure. 13C**) and AS160 T642 (**Figure 13D**) were not significantly different from scrambled control. Altogether this suggests that GDF15 alone could be a *bonafide* insulin sensitizer. To further test this proposition *in vitro*, we stimulated fully differentiated human myotubes with GDF15 from commercial suppliers. Treatment increased insulin signaling (**Figure 13E**), as shown by increased phosphorylation AS160 T642 (**Figure 13F**), AKT T308 (**Figure 13G**) insulin receptor β Y1150/1151 (**Figure 13I**), but not ERK 1/2 p42/44 (**Figure 13H**). Furthermore, GDF15 treatment also increases glycolytic flux (**Figure 13J**), glucose uptake (**Figure 13K**) and insulin mediated GLUT4 translocation (**Figure 13L**). Altogether this suggests that GDF15 could directly act on skeletal muscle.

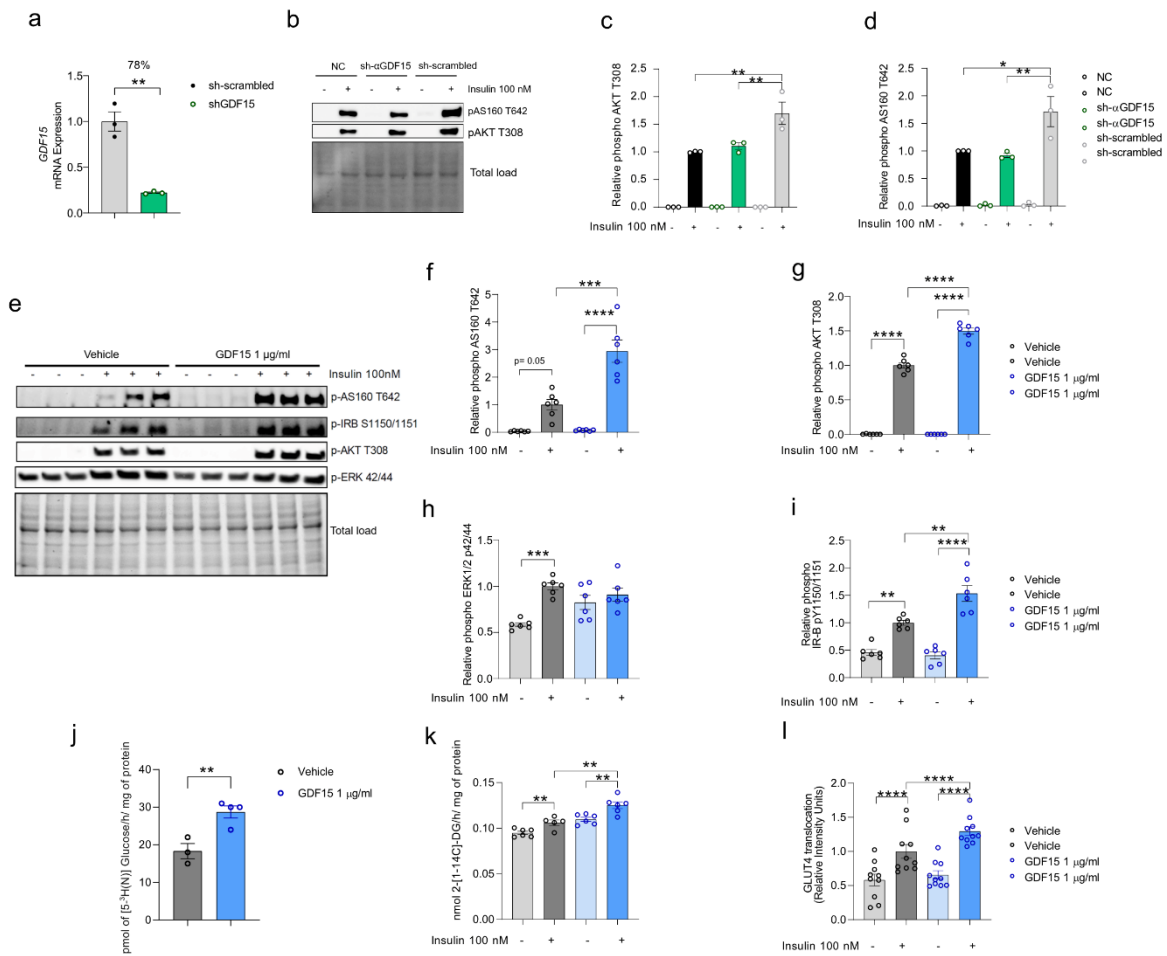


Figure 13 GDF15 stimulation increases insulin sensitivity and glucose handling in human myotubes. **a.** *GDF15* knockdown efficiency in E4ORF1 HUVECs using sh- α GDF15 lentiviral viral particles. (n= 3)(Student-*t* test). **b.** Insulin signaling cascade from human myotubes that have been stimulated over 48 h with CM from E4ORF1 in serum free conditions or control. CM was either produced from sh-scrambled or sh- α GDF15 endothelial cells. (n= 3). **c.d.** Human myotubes treated with CM from sh- α GDF15 failed to increase insulin sensitivity as shown by quantification of downstream targets AKT T308 and AS160 T642 ((Two-way ANOVA and Tukey's multiple comparison). **e.** Representative blot from human Myotubes were treated for 48 h with rhGDF15 and then stimulated with insulin 100 nM. In figures **b** and **e**, the total load shows comparable amounts of protein loaded in the representative blot. **f. g. i.** rhGDF15 increases insulin sensitivity in human myotubes *in vitro* by increasing phosphorylation of downstream targets AS160 T642, AKT T308, IR- β Y1150/1151 upon maximal insulin stimulation 100 nM. (n= 6)(Two-way ANOVA and Tukey's multiple comparison). **h.** rhGDF15 stimulation of human myotubes does not increases ERK1/2 p42/44 as compared to other downstream targets (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **j.** rhGDF15 stimulation in human myotubes increases glycolytic flux (n= 4)(Student-*t* test). **k.** rhGDF15 stimulation in human myotubes increases insulin mediated glucose uptake assessed by radioactive 2-deoxy glucose incorporation (n= 6) (Two-way ANOVA and Tukey's multiple comparison). **l.** 48 h stimulation of rhGDF15 (kindly donated by Group Leader Dr. Sebastian Jørgensen, Novonordisk) increases insulin-mediated GLUT4 translocation in rat L6-GLUT4-MYC myotubes (n= 10). (Two-way ANOVA and Tukey's multiple comparison). Significant values are represented in asterisks as follows * P <0.05, ** P <0.01, *** P <0.001, and **** P <0.0001.

4.4.3. GDF15 administration improves glucose homeostasis and insulin response *in vivo*.

Despite the growing evidence suggesting that GDF15 could also act locally (Chung, Ryu et al. 2017, Jung, Choi et al. 2018, Nakayasu, Syed et al. 2020), the expression of the only known receptor, GFRAL, can be found exclusively in the area postrema of the hindbrain (Emmerson, Wang et al. 2017, Hsu, Crawley et al. 2017, Mullican, Lin-Schmidt et al. 2017, Yang, Chang et al. 2017). and other areas of the central nervous system (CNS), however, is absent in peripheral tissues (Li, Wang et al. 2005, Hsu, Crawley et al. 2017, Emmerson, Duffin et al. 2018, Nakayasu, Syed et al. 2020) Nonetheless these observations may be contradictory, a plausible scenario is that GDF15 acts through a yet unknown receptor that has not been described in periphery. The *in vitro* data in this work, so far suggests that this unknown mechanism or receptor could be present in skeletal muscle to partially mediate insulin sensitivity and improved glucose homeostasis.

Insulin sensitivity and improved glucose homeostasis have been described in literature upon administration of GDF15 *in vivo*. However, they are often referred as to be secondary to the effects of reduced body weight in animals fed an obesogenic diet. Therefore, to test whether GDF15 has insulin sensitizing properties that are primary to its stimulation and not secondary to weight loss, we decided to administer GDF15 acutely 3 times every other day. Assessment of their body weight indicated that there were no significant differences between treated rhGDF15 animals and controls after 5 days (**Figure 14A**). Furthermore, insulin signaling in skeletal muscle showed increased phosphorylation in insulin signaling cascade (**Figure 14B**) as shown by AKT T308 (**Figure 14C**) and AS160 T642 (**Figure 14D**). Yet, when assessing insulin responsiveness in an insulin tolerance test (ITT), GDF15 treated mice showed lower glucose levels in different time points compared to control mice (**Figure 14E**). However, no differences were found in a glucose tolerance test (GTT) (**Figure 14F**), which suggest that endogenous levels of insulin release are not sufficient to elicit changes in glucose levels in chow fed mice. Finally, when assessing glucose uptake in a radioactive assay *in vivo* we observe that different muscle like tibialis anterior (Ta), extensor digitorum longus (Edl), soleus and oxidative gastrocnemius (Rgas)(**Figure 14G**). Despite these interesting findings, further experiments are still necessary to

elucidate the molecular mechanisms in charge of GDF15 insulin sensitizing effects on skeletal muscle *in vivo* and *in vitro*.

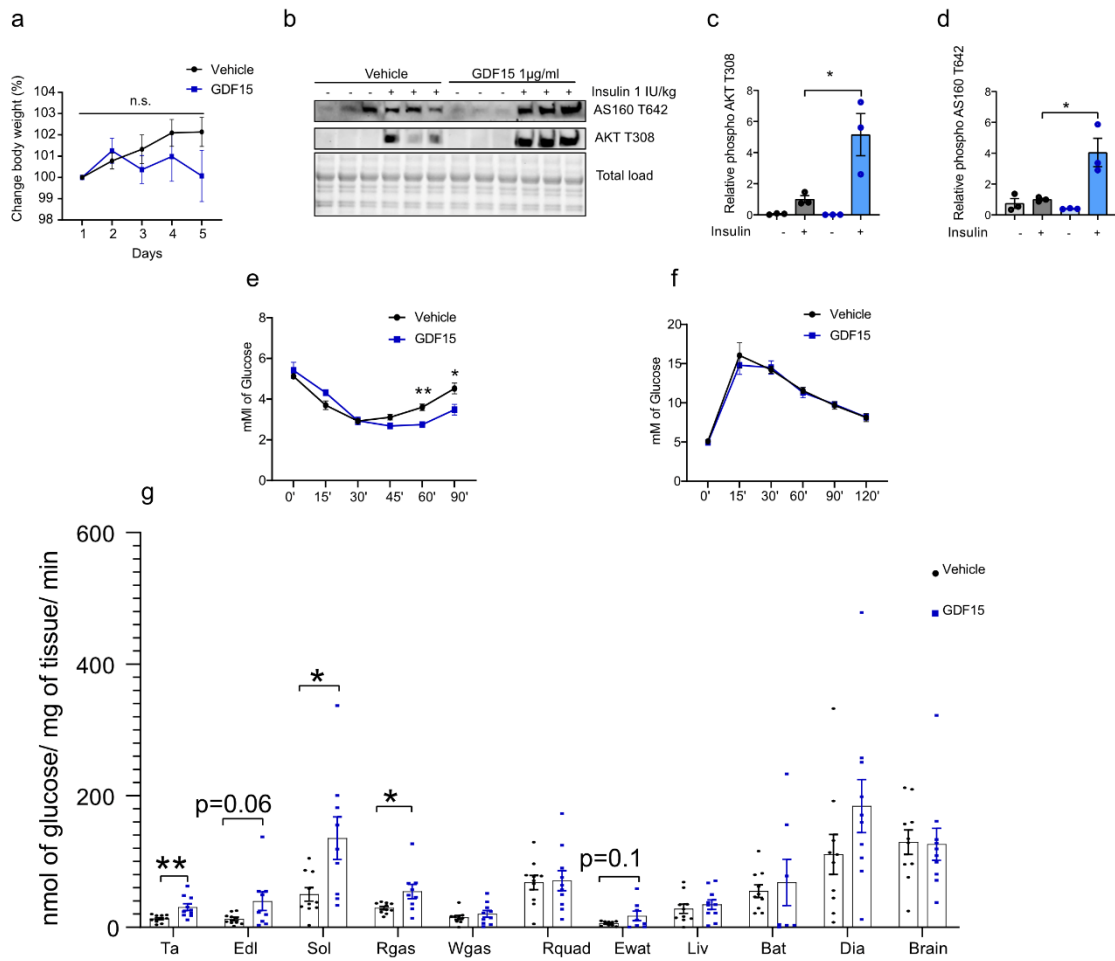


Figure 14 Administration of rhGDF15 promotes increased insulin sensitivity and greater glucose uptake in skeletal muscle in mice fed chow diet. Mice were administered 8 nmol of rhGDF15 at the beginning of their dark cycle. **a** There are no differences in changes in body weight after 5 days between controls (n= 6) and GDF15 treated animals (n= 6) (Two-way ANOVA with Sidak's multiple comparison). **b** GDF15 treated mice show an increased insulin signaling upon stimulated with 1 IU/kg body weight insulin for 15 min, the total load shows comparable amounts of protein loaded in the representative blot. **c. d.** Downstream insulin signaling was increased as measured by phosphorylation of AKT T308 and AS160 T642(Two-way ANOVA and Tukey's multiple comparison). **e.** rhGDF15 treatment improves insulin sensitivity as shown by ITT (vehicle, n= 4, rhGDF15, n= 3) (Two-way ANOVA and Sidak's multiple comparison) **f.** but treatment does not influence glucose handling as assessed in ipGTT (n= 4) (Two-way ANOVA and Sidak's multiple comparison). **g.** *In vivo* insulin-stimulated radioactive 2-Deoxy glucose uptake of several tissue (Student-t test). *Ta*, tibialis anterior; *Edl*, extensor digitorum longus; *Sol*, soleus; *Rgas*, red gastrocnemius; *Wgas*, white gastrocnemius; *Rquad*, red quadriceps; *Ewat*, epididymal white adipose tissue; *Liv*, liver; *Bat*, brown adipose tissue; *Dia*, diaphragm. Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001

4.5. Discussion

Skeletal muscle constitutes approximately 45% of total body mass in humans (Spargo, Pratt et al. 1979), which makes it the largest metabolic organ in the adult and a major site of glucose uptake (Kim and Kim 2020). Indeed, skeletal muscle is recipient of up to 80% glucose uptake postprandially (Thiebaud, Jacot et al. 1982), which makes it a central component in whole body energy homeostasis and its dysregulation leads to metabolic disease and diabetes. However, the extent to which it responds to insulin greatly varies depending on the health status (Wu and Ballantyne 2017, Schwartsburd 2019), age (Gupte, Bomhoff et al. 2008), and physical activity of an individual (Richter and Hargreaves 2013). Hence, skeletal muscle shows metabolic plasticity that is adapted to meet physiological demands of the body. Importantly, this metabolic plasticity also shows circadian rhythmicity, during active cycle skeletal muscle is more responsive to the effects of insulin, in both, signaling and glucose clearance potential; while in the inactive cycle the signaling induction is dampened (Basse, Dalbram et al. 2018). Furthermore, skeletal muscle insulin sensitivity can be modulated by the stimulation of several growth factors, cytokines, hormones and metabolites. In this work, we present angiocrine GDF15 as a *bonafide* insulin sensitizer in skeletal muscle, which independent of weight loss.

Previously, published literature suggests that an increased in muscle capillarization enhances insulin sensitivity after exercise and detraining in senile human subjects independently from known exercise-induced benefits to glucose handling (Prior, Goldberg et al. 2015), this vascular control of glucose homeostasis has also been replicated in other target organs such as adipose tissue (Robciuc, Kivelä et al. 2016), pancreas (Duvillié 2013, El-Gohary and Gittes 2018, Obata, Kimura et al. 2019) and liver (Tsuchiya and Accili 2013). We show that vascular derived secreted factors can modulate skeletal muscle insulin sensitivity. In an *in vitro* model system, we stimulated human skeletal muscle myotubes with conditioned media from HUVECs in serum free conditions, ultimately allowing us to delimit endothelial cell secretome insulin sensitizing capabilities in human skeletal muscle myotubes. Indeed, we found that CM from endothelial cells promotes increase insulin signaling by the increased phosphorylation of AKT, AS160 and ERK1, and increased GLUT4 translocation in rat L6 myotubes.

We explored whether the consequential enhanced glucose handling *in vivo* is independent of anorectic effects arising from the central nervous system (CNS). We found that human recombinant stimulation of GDF15 on human myotubes *in vitro* mimic the improved metabolic properties of endothelial CM, which leads to an increased insulin signaling increased phosphorylation, increased GLUT4 translocation to cell membrane. Thus, suggesting that GDF15 stimulation in skeletal muscle could go via an independent mechanism of GFRAL

Skeletal muscle is subject to the action of different cytokines that modulate its insulin sensitivity and glucose handling potential. To exemplify this, adipose tissue has been identified to be a metabolic organ with secretory capacity that signals to muscle. It modulates the expression of positive and negative signaling molecules. Fibroblast Growth Factor 21 (FGF 21) is among the molecules that exert an improved glucose handling phenotype. It acts on skeletal muscle by preventing insulin resistance upon high fat diet through peripheral and CNS (BonDurant, Ameka et al. 2017) , mediated effects in preclinical models of diabetes in rats (Sarruf, Thaler et al. 2010) and rhesus monkeys (Kharitononkov, Wroblewski et al. 2007). Closely associated to FGF21, adiponectin is yet another adipokine that improves GLUT4 translocation in muscle through the activation of its receptor adiponectin receptor 1 (AdipoR1) and adiponectin receptor 2 (AdipoR2) (Ceddia, Somwar et al. 2005). Importantly, signaling via adiponectin receptor is blunted upon overexpression of a dominant negative AMP kinase (AMPK), which supports the notion that an adequate response against energy stress is fundamental in regulation of glucose homeostasis(Yamauchi, Kamon et al. 2002). Furthermore, skeletal muscle stimulation with adiponectin increases fatty acid uptake and oxidation via mitochondrial biogenesis and suppresses fatty acid synthesis through the actions of AMPK, p38 and PPAR α (Fruebis, Tsao et al. 2001, Yamauchi, Kamon et al. 2002, Yoon, Lee et al. 2006, Ritchie and Dyck 2012).

We identified that GDF15 interacts directly with skeletal muscle. Seminal publications identified the direct relationship between GDF15 and skeletal muscle metabolic benefits by improving lipid mobilization and oxidative metabolism (Chung, Ryu et al. 2017). Beyond the skeletal muscle, GDF15 has been demonstrated to have metabolic benefits in liver(Kim, Kim et al. 2018), adipose tissue (Chrysovergis, Wang et al. 2014), macrophages(Jung, Choi et al. 2018) and pancreas (Nakayasu, Syed et al. 2020). We showed increased insulin signaling and increased glucose uptake in several muscle

like TA, EDL, RGAS and soleus. These improved glucose handling was accompanied by an increased insulin sensitivity as shown by ITT.

In conclusion, we present evidence that endothelial cell secretome is able to stimulate skeletal muscle to have a greater responsiveness to insulin. Furthermore, that endothelial cells secrete GDF15 in pharmacologically relevant concentrations. *In vitro*, GDF15 was able to increase insulin signaling and GLUT4 translocation. *In vivo* GDF15 was able to induce an increase insulin sensitivity, an increased skeletal muscle insulin sensitivity and glucose uptake in muscle like TA, EDL, RGAS and soleus. Importantly, these insulin sensitizing effects are independent of weight loss, which furthers GDF15 as a pleiotropic metabolic regulator against deleterious effects of obesity and diabetes.

4.6. Methods

Cell culture.

HUVECs and E4ORF1 HUVECs were cultured on gelatin coated plates and M199 medium (1 mg/ml D-glucose) supplemented with 20% fetal bovine serum (FBS), 2mM L-glutamine, 30 mg/l endothelial cell growth factor supplements (Sigma E2759), 10 units/ml heparin (Sigma H3149), 50 IU/ml penicillin and 50 µg/ml streptomycin. Cells were routinely maintained in 5% CO₂ and 95% air at 37 °C and medium changed every 48 h. E4ORF1 HUVECs were a kind donation from the lab Prof. Shahin Rafii at the Weill Cornell School of Medical Science, New York. CM was produced culturing EC over 48 h in the aforementioned conditions. CM was subsequently concentrated using Amicon Ultra filters with a molecular weight cut-off value of 3 kDa (Merck-Millipore UFC900396).

Culture and differentiation of human myoblasts

Human myoblasts from healthy donors were obtained from Cook Myosite. Myoblasts were cultured in proliferation media (Cook Myosite MB-2222) avoiding confluence greater than 60% to ensure cell-cell contact and thus spontaneous differentiation. For differentiation, human myoblasts were seeded in collagen coated well-plates and medias was replaced for differentiation media (Cook Myosite MD-5555). Briefly, for the collagen coating, 10% v/v of collagen solution (Gibco A10483-01) was prepared in PBS. Coating solution was incubated on plates at 37 °C for 4 h or overnight at 4 °C. Solution was discarded, and plates dried for 2 h. Myoblasts differentiated for 7 days before using in assays. *In vitro*, human myoblasts were stimulated with rhGDF15 (R&D Systems, 957-GD-025, source unless otherwise noted) for 48 h at a concentration of 1 µg/ml in differentiation media (Cook Myosite MD-5555).

Glycolysis Flux

Determination of glycolytic flux was performed as described before (Veys, Alvarado-Diaz et al. 2019). Briefly, cells were incubated for 2 h in culture medium containing D-[5-³H(N)]-glucose (NET53100, PerkinElmer,) at a final concentration of 0.4 µCi/mL medium. The supernatant was then transferred into glass vials sealed with rubber stoppers, and ³H₂O was captured in hanging wells using a H₂O-soaked Whatman paper for 48 h at 37°C to reach saturation. Radioactivity in ³H-labeled paper was

determined by liquid scintillation counting (LSC) and the glycolytic flux was measured by the rate of $3\text{H}_2\text{O}$ production

Radioactive Glucose Uptake *in vitro*.

Fully differentiated human myotubes were incubated for 3 h in no glucose Krebs-Ringer-Buffer for 3 h to starve them. Myotubes were incubated for 30 min in 100 nM insulin (Actrapid, Novonordisk) in Krebs-Ringer-Bicarbonate buffer. Myotubes were washed quickly with clean Krebs-Ringer-Bicarbonate buffer. Tracer was added 1 μCi 2-[1- ^{14}C]-Deoxy Glucose/ml (Perkin Elmer NEC495A001MC) in Krebs Ringer Buffer. Labeling occurred over 1 h. Immediately after, cells were wash twice with ice cold PBS. Then, they were lysed with 1 000 μl of 1 M NaOH at room temperature for 30 min.

Colorimetric GLUT4 translocation assay.

L6-GLUT4-MYC rat myoblasts were a kind donation from Prof. Amira Klip, University of Toronto, Hospital for Sick Children, Toronto, Canada. L6-Glut4-Myc cells were grown in α -Minimal Essential Medium (α -MEM) (Gibco, 12571-06) supplemented with 10% FBS 50 IU/ml penicillin and 50 $\mu\text{g}/\text{ml}$ streptomycin until confluency and differentiated in α -MEM supplemented with 2% horse serum (HS) 50 IU/ml penicillin and 50 $\mu\text{g}/\text{ml}$ streptomycin over 5 days. For insulin stimulation, serum-starve cells (3 hours, 37°C) prior to experimentation. Insulin (Actrapid, Novonordisk) was added at a concentration of 100 nM for 20 minutes and incubated at 37°C . Some cells were left untreated for measurements of basal GLUT4-MYC density and background immunoreactivity, which was used to further calculate relative intensity units, basal GLUT4-MYC background was considered with a relative intensity unit of 1. To end the incubation, place cells were placed on ice and quickly washed twice with ice-cold PBS (1 mL per wash). Cells were incubated with cold 3% paraformaldehyde (PFA) in PBS for 15 min at 4°C . PFA was aspirated, cells were washed twice with PBS and incubated with 0.1 M glycine in PBS at 4°C for 10 min to quench trace formaldehyde. Cells were blocked with 5% goat serum in PBS for 15 min at 4°C , approx. 1 ml per well). Cells were incubated with 1 mg/ml anti-MYC polyclonal antibody containing 5% goat serum for 60 min at 4°C , 250 μl approximately. For background staining, two wells were left untreated without the primary antibody per plate. Absorbance determinations give the

background and should be subtracted from each condition. Cells were washed six times with PBS at 4°C with aspiration, avoid disturbing attached cells between washes. Cells were incubated with a 1:1000 dilution of HRP-conjugated goat anti-rabbit IgG in PBS containing 5% goat serum for 45 min at 4°C. Incubation can also be performed for 30 min at room temperature. Cells were washed six times with PBS. Cells were incubated with 1 ml per well of OPD peroxidase reagent (Sigma, P9187) at room temperature for up to 30 min protected from light. Stop the reaction with the addition of 250 µl of 3M HCl per well and gentle mixing. Absorbance was read at 492 nm in a multiplate reader (Tecan, Spark)

Mice

Wildtype C57BL/6J mice were obtained from The Jackson Laboratory (000664 | B6). Mice were randomly allocated to different treatment groups, and the investigator was blinded to the group allocation during the experiment as well as during the analysis. All mice were housed at standard housing conditions (22 °C, 12 h inverted light/dark cycle), with ad libitum access to chow diet (18 % proteins, 4.5 % fibers, 4.5 % fat, 6.3 % ashes, Provimi Kliba SA) and water. Health status of all mouse lines was regularly monitored according to FELASA guidelines. Animal experiments were approved by the local animal ethics committee (Kantonales Veterinärämtesamt Zürich, licenses ZH014/2016 and ZH211/2019), and performed according to local guidelines (TschV, Zurich) and the Swiss animal protection law (TschG). All mice used in this study were between 8-12 weeks old. Both male and female mice were included in the study. rhGDF15 (R&D Systems, 957-GD-025; or donated by Group Leader Dr. Sebastian Jørgensen, Novonordisk) was administered intravenously

Insulin Tolerance Test (ITT)

Mice were food starved during 6 h at the beginning of the dark cycle. Mice body mass was determined prior the beginning of the assay. Insulin (Actrapid, Novonordisk) was diluted in saline and administered intraperitoneally at dose of 0.65 IU/ kg of body weight for females and 0.75 IU/kg body weight for males. Blood was samples by a small cut in the tail vein in the following timepoints: 0 min, 15 min, 30 min, 45 min, 60 min and 90 min with a portable glucose meter.

Glucose Tolerance Test (GTT)

Mice were food starved during 6 h at the beginning of the dark cycle. Mice body mass was determined prior the beginning of the assay. D-Glucose (Sigma G8270) was diluted 10% w/v in saline. Mice body mass was determined prior the beginning of the assay. Dose of glucose was 1 g/ kg of body weight and administered intraperitoneally. Blood was samples by a small cut in the tail vein in the following timepoints: 0 min, 15 min, 30 min, 60 min, 90 min and 120 with a portable glucose meter (AccuCheck AVIVA, 06870317001).

Skeletal muscle *in vivo* insulin stimulation.

Mice were food starved during 6 h at the beginning of the dark cycle, while in starvation they had free access to water. Mice body mass was determined prior the beginning of the assay. Insulin (Actrapid, Novonordisk) was diluted in saline and administered intraperitoneally at dose 1 IU/kg of body weight and skeletal muscle was taken 15 min later. Animals were anesthetized 5 min before muscle was dissected with pentobarbital. Muscle samples were snap frozen and stored at -80 °C until further processing.

***In vivo* radioactive glucose uptake.**

Determination of radioactive glucose uptake *in vivo* has been described before (10.1038/s41598-017-15548-6). Briefly, mice were food starved during 6 h at the beginning of the dark cycle, while in starvation they had free access to water. Mice body mass was determined prior the beginning of the assay. Mice were injected 5 μ Ci of 2-[1-¹⁴C]-Deoxy Glucose intravenously (Perkin Elmer NEC495A001MC). After 5 min, mice were administered 1 IU/kg body weight of insulin (Actrapid, Novonordisk) intraperitoneally and tissues were taken 15 min later. Of note, mice were euthanized while in anesthesia with pentobarbital 5 min before tissue harvesting. Glucose levels were measured by a small cut in the tail vein in the following timepoints considering insulin administration as zero: -5, 0, 7.5 and 15 min with a portable glucose meter (AccuCheck AVIVA, 06870317001) and blood was sampled to calculate circulating specific activity from plasma. Tissues were quickly dissected, weighted, and digested overnight with 0.5 ml of 1 M NaOH at 65 °C. Next day, reaction was neutralized by adding 0.5 ml of 1M HCl. Each sample was split in two parts: 200 μ L of lysate should

be mixed with 0.5 ml ZnSO₄ and 0.5 ml Ba (OH)₂; while another 200 µL of lysate with 1 ml of 6% perchloric acid. (Sigma 244252). Both samples groups were vortexed thoroughly and centrifuged 13 000 RCF for 2 min. A volume of 800 µl supernatants were transferred to 5 ml of scintillation liquid (Perkin Elmer Ultima Gold 6013329) and counted with liquid scintillator (Tri-Carb 2000CA). The subtraction of values of Ba(OH)₂ + ZnSO₄ from perchloric acid gives a quantitative value of phosphorylated 2DG and thus glucose uptake.

Transfection of packaging cells and lentiviral transduction of HUVEC's.

Briefly, on the first, 2 x 10⁶ HEK 293T cells were plated on 10 cm dishes. On the second day, HEK293T cells were transfected with 3.4 µg of pMD2.G (AddGene, Plasmid #12259), 3.4 µg of psPAX2 (AddGene, Plasmid #12260) and 5.4 µg of GIPZ lentiviral shRNA to target *gdf15* (RHS4531) were acquired from Dharmacon (Horizon Discovery; Waterbeach, United Kingdom). A nonsense scrambled shRNA sequence was used as control. Plasmids used for transfection were used with Lipofectamine 2000 (Thermo Fisher 11668019) according to instructions of manufacturer in Opti-MEM (Thermo Fisher 31985070) and incubated for 4h, cell medium was changed to medium containing 10% serum. E4ORF1 HUVEC's or WT HUVEC's were used for lentiviral transduction, they were seeded one day before at a confluence of 8 x 10⁵ cells in a 10 cm dish. Viral particle- containing supernatant were filtered through a 0.45 µm filter mouter syringe. Polybrene was used at a concentration of 8 µg/ml for three consecutive days with viral particle containing supernatants. Transduced HUVEC's were selected with 2 µg/ml of puromycin over 3 days. Puromycin containing medium was changed every 24 h.

RNA extraction and quantitative RT-PCR

RNA isolate from cell cultured HUVECs was extracted using PureLink™ RNA Mini Kit (12183020, Thermo Fischer). RNA purity and concentration were assed via a spectrophotometer (Tecan, Spark). RNA was reverse-transcribed to cDNA by High Capacity cDNA Reverse Transcription Kit (Thermo Fisher, 43-688-13). A SYBR Green-based master mix (Thermo Fisher Scientific, A25778) was used for real-time qPCR analysis with primers listed in Table 1. To compensate for variations in RNA input and

efficiency of reverse-transcription, beta actin was used as a housekeeping gene. The delta-delta CT method was used to normalize the data.

Immunoblotting.

Cells were collected and lysed with [50 mM Tris–HCl pH 7.0, 270 mM sucrose, 5 mM EGTA, 1 mM EDTA, 1 mM sodium orthovanadate, 50 mM glycerophosphate, 5 mM sodium pyrophosphate, 50 mM sodium fluoride, 1 mM DTT, 0.1% Triton-X 100 and a complete protease inhibitor tablet (Roche Applied Science)]. Lysates were centrifuged at 10,000 RCF for 10 min at 4°C. Supernatant was collected and protein concentration was measured using the DC protein assay kit (5000116, Bio-rad). 5-10 µg of total protein was loaded in a 10-well pre-casted gradient gel (456-8086, Bio-Rad). After electrophoresis, a picture of the gel was taken under UV-light to determine protein loading using stain-free technology. Proteins were transferred onto a PVDF membrane (Bio-rad, 170-4156) with a semi-dry system and subsequently blocked for 1 h at room temperature with 5% milk in 0.1% TBS-Tween. Membranes were incubated overnight at 4 °C with primary antibodies see table below. The appropriate HRP-linked secondary antibodies were used for chemiluminescent detection of proteins. Membranes were scanned with a Chemidoc imaging system (Bio-rad) and quantified using Image Lab 6 software (Bio-rad). Normalization of data was carried out dividing chemiluminescent intensity of a given band by the one of their respective total protein load.

Figures and Diagrams

Figures in this manuscript were created with BioRender.com.

Quantification and statistical analysis

The images presented in the manuscript are representative of the data (quantification of image is approximately the group average) and the image/staining quality. All data represent mean \pm SEM. GraphPad Prism software (version 8.0.0) was used for statistical analyses. Investigators were always blinded to group allocation. Unless otherwise indicated, when comparing two group means, Student's t-test was used in an unpaired two-tailed fashion. For more than two groups, one-way ANOVA with

Tukey's multiple comparisons test was used and for experimental set-ups with a second variable, two-way ANOVA with Sidak's multiple comparisons test was used. The statistical method used for each experiment is indicated in each figure legend. Asterisks in figure legends denote statistical significance. No experiment-wide multiple test correction was applied. $P > 0.05$ is considered non-significant (ns). $P < 0.05$ is considered significant (*).

List of primers

GDF15 (h.)	ATACTCACGCCAGAAGTGCGG	GAACAGAGCCCGGTGAAGGC
ACTB(m. h.)	GCTCCTCCTGAGCGCAAG	CATCTGCTGGAAGGTGGACA

List of antibodies

p-AKT (Thr 308)	13038S	Cell signaling
p-AKT (Ser 473)	4060S	Cell signaling
p-AS160 (Thr 642)	8881S	Cell signaling
p-ERK1/2 (p42/44)	9101S	Cell signaling
pAMPK (T172)	2531L	Cell signaling
pIR- β (Tyr 1150/1151)	3024L	Cell signaling

5. Concluding remarks

The secretory properties of ECs in modulation of tissue metabolism is still a developing area. The prevailing view is that metabolism is mostly controlled unidirectionally from signals arising in tissues towards vascular endothelium. While this has been long validated, seminal work has suggested that ECs, as well, exert an influence on their surrounding tissue that may counteract the commencement, progression and even reversal of metabolic disease. In other words, they suggest that the metabolic communication between endothelial cells and tissues is bidirectional instead. However, mechanistic approaches are lacking that may shed light on the molecules, receptors and signaling driving these findings.

In this project, through an unbiased approach, we have revealed the identity of novel angiocrine protein, GDF15. Furthermore, we identified a previously unrecognized property on glucose homeostasis. GDF15 has insulin sensitizing properties, which are acute and independent of body weight loss. Interestingly, by modulating the activity of transcriptional master regulators in ECs, we showed that FOXO1 negatively controls GDF15 expression. This is, FOXO1 activity decreases, while FOXO1 abrogation increase expression of GDF15. This opens a new paradigm in the angiocrine control of tissue metabolism, as angiocrine expression of GDF15 potentially links immunity, inflammation response, regulation of food intake and body mass, adaptations to exercise, glucose handling and insulin sensitivity to vascular biology. Nonetheless, proper experimental examination are necessary to test the limits of these new propositions.

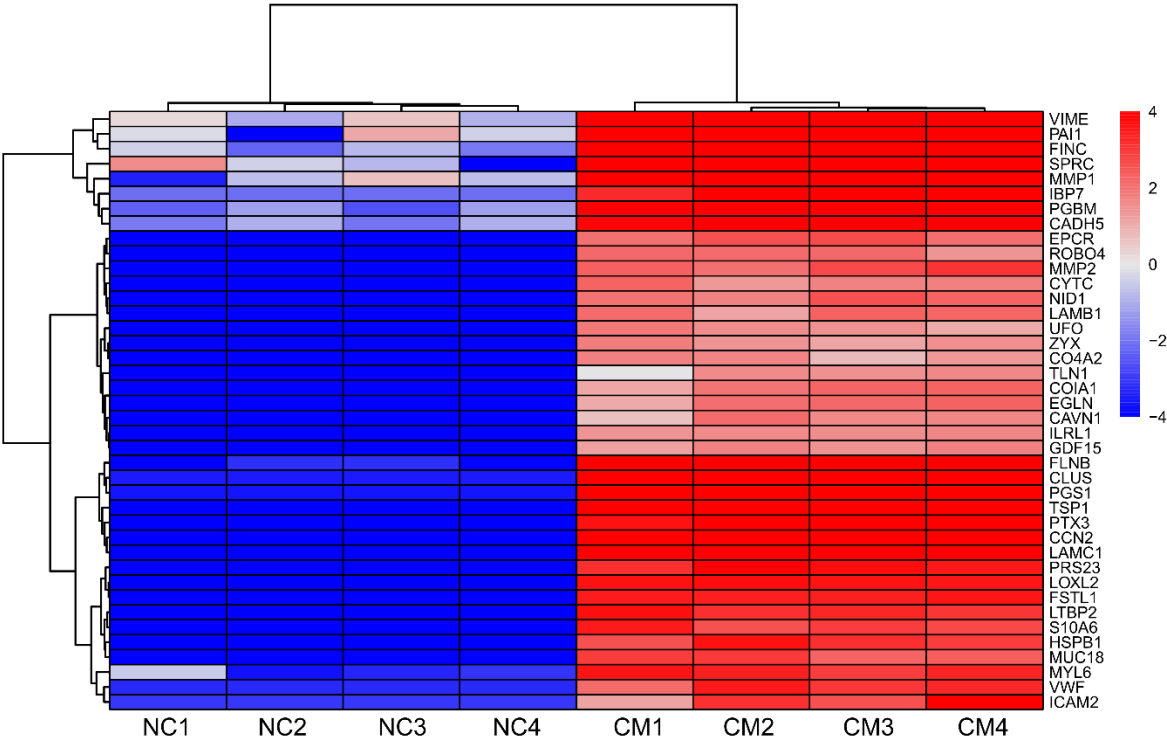
One of the questions that remain to be answered is whether a reduction of food intake is the main purpose of angiocrine GDF15 upon exiting quiescence transcriptional program when FOXO1 is abrogated. This, due to the small yet significant increase in circulating GDF15. It is unlikely there is a non-pathological condition which would lead to a general vascular exit of quiescence that would parallel the genetic model we have used in this project. Rather, the integration of all this knowledge would point towards a

local influence on cells and tissue in close vicinity to endothelial cells secreting GDF15. Along the same train of thought, another question that remains to be answered is whether angiocrine GDF15 would also promote metabolic beneficial effects over longer periods of time, for instance during aging or to even extend life span of an organism.

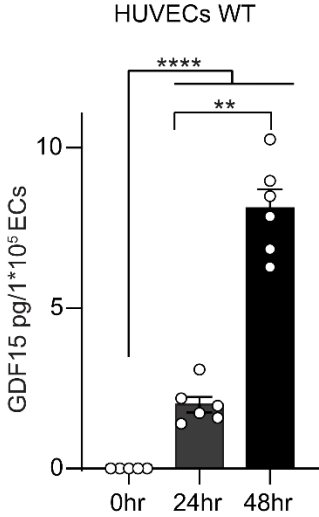
This project has contributed to the exciting new avenues of research and potential treatments that are being explored where angiocrine secretion of proteins/metabolites produced in different stages of vascular development are taking the central stage.

6. Extended data figures

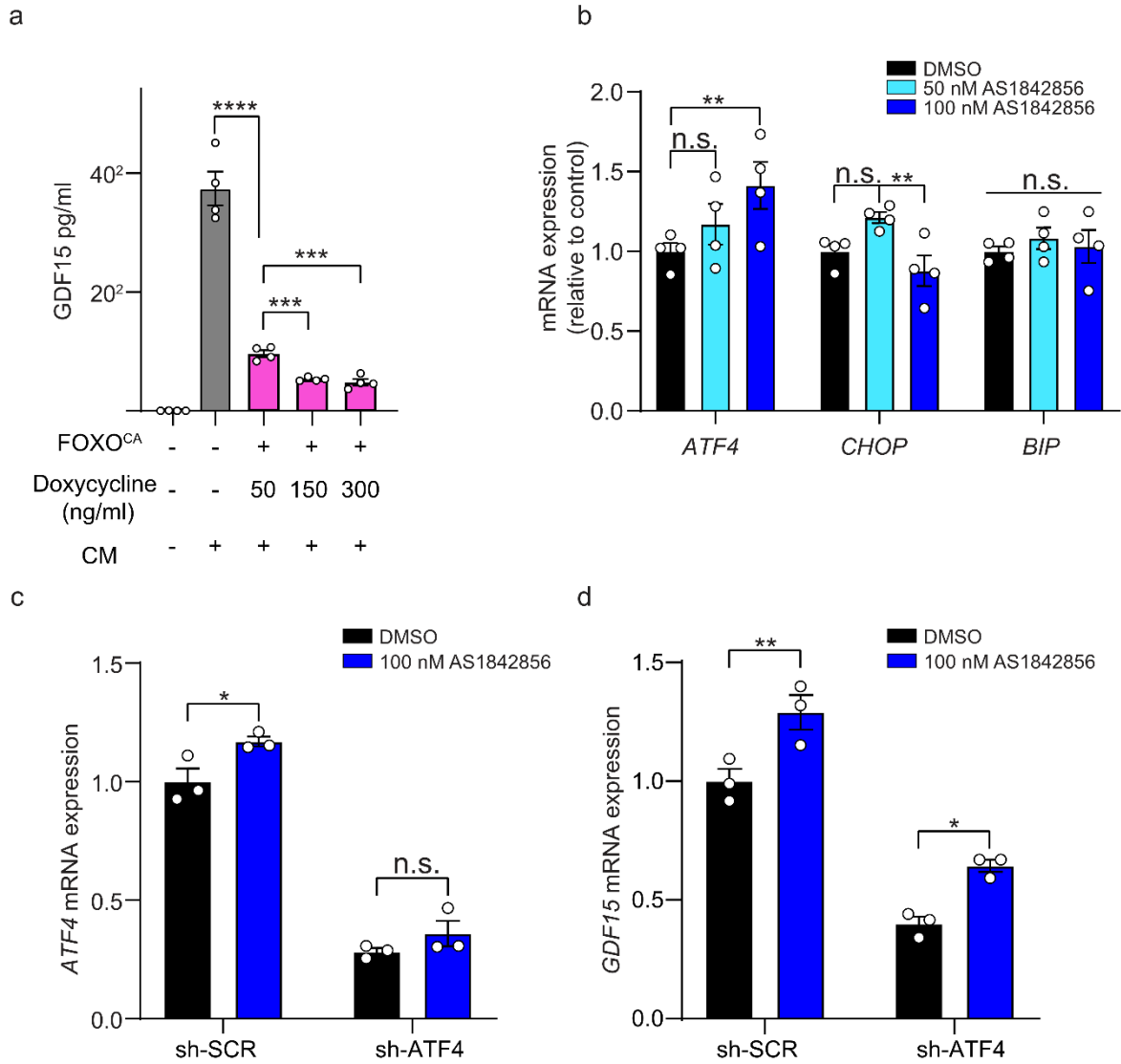
a



b

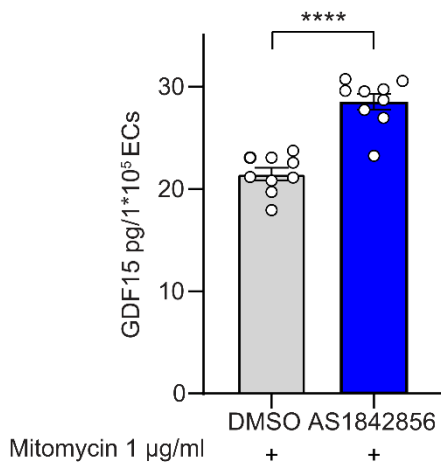


Extended data figure 1 Workflow of generation and quantification of secretome of HUVECs produced in serum-free conditions. **a.** Heatmap of the top 40 most upregulated proteins present in CM. Uniquely identified proteins were included in the analysis, relative abundance is shown as escalated expression compared to control. **b** GDF15 accumulates in HUVECs culture supernatants in standard culturing conditions over time (0 h, 24 and 48 h), GDF15 ELISA. (Student-*t* test) **P*<0.05, ***P*<0.01, ****P*<0.001, and *****P*<0.0001

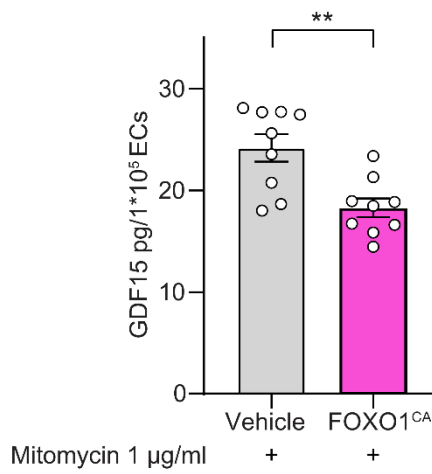


Extended data figure 2 FOXO1, but not NOTCH1, regulates expression of GDF15 in E4ORF1 and WT HUVECs. **a**. Dose response secretion of GDF15, 48 hrs. after induction of FOXO1CA with 50, 150 and 300 ng/ml doxycycline in Tet-On FOXO1CA E4ORF1 HUVECs in serum free conditions (n=4) (Two-way ANOVA and Tukey's multiple comparison). **b** FOXO1 inhibition, AS1842856, increases in a dose response expression of *ATF4* but not *CHOP* (*CHOP* was upregulated with 50 nM but not 100 nM AS1842856 treatment) or *BIP* (Two-way ANOVA with Tukey's multiple comparison). **c**. Knockdown of *ATF4* in HUVECs (n=3) **d** did not blunt *GDF15* upregulation upon FOXO1 pharmacological blockade (n=3) (Student-*t* test). Housekeeping gene *ACTB* was used as control for figure panels **b**, **c**, **d**. Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001

a



b



Extended data figure 3 A block of proliferation by Mitomycin C treatment does not blunt FOXO1 negative regulation on GDF15 in vitro. **a.** GDF15 secretion of HUVECs that were treated with FOXO1 inhibitor, AS1842856, 100 nM (n= 9), or an equivalent volume of vehicle, DMSO (n= 9), both in the presence of Mitomycin C 1 µg/ml. Cell culture supernatants were collected after 72 hours. (Student-t test) **b.** GDF15 secretion of FOXO1CA HUVECs (n= 9) or controls cells (n= 9) that were treated with Mitomycin C 1 µg/ml. Cell culture supernatants were collected after 72 hours. (Student-t test). Significant values are represented in asterisks as follows *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001

7. Supplemental Tables.

Supplemental table 1. Full list of identified proteins in proteomic analysis. In yellow are highlighted uniquely identified proteins.

1	ProteinName	# Pepti des	Ctrl	Group	log2FC	CL.L	CL.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M.3.Co ntrol.ra w	M.2.Co ntrol.ra w	M.4.Co ntrol.ra w	M.1.Co ntrol.ra w	E4.2.Gr oup_1r aw	E4.4.Gr oup_1r aw	E4.3.Gr oup_1r aw	E4.1.Gr oup_1r aw	M.3.C ontr oltransf	M.2.C ontr oltransf	M.4.C ontr oltransf	M.1.C ontr oltransf	E4.2. Group _1tran sf	E4.4. Group _1tran sf	E4.3. Group _1tran sf	E4.1.G roup_1 transf	pseud o.Ctrl	pseud o.Group pl	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val	
2	O75369FLNB	77	-3.72	5.12	8.84	7.93	9.75	0.70	2183	2.70E-09	1.25E-06	11.29	0	3.07E+06	2.59E+06	2.20E+06	3.68E+06	3.98E+09	3.26E+09	3.67E+09	9.07E+08	-3.906	-3.175	-4.640	-3.158	5.228	5.395	5.309	4.541	-3.720	5.118	8.838	2.70E-09	1.25E-06	
3	P08670VIMB	42	-0.29	8.73	9.02	8.06	9.98	4.22	2111	3.67E-09	1.25E-06	11.06	0	2.62E+07	1.21E+07	6.43E+07	5.93E+07	3.32E+10	2.79E+10	3.46E+10	2.07E+10	-0.937	-1.052	0.245	0.589	8.394	8.732	8.712	9.086	-0.289	8.731	9.020	3.67E-09	1.25E-06	
4	P02751FINC	46	-1.33	6.83	8.16	7.15	9.17	2.75	1819	1.42E-08	3.21E-06	10.03	0	1.33E+07	5.01E+06	4.18E+07	2.14E+07	1.20E+10	9.04E+09	1.18E+10	3.25E+09	-1.878	-2.263	-0.379	-0.788	6.871	6.981	7.081	6.397	-1.327	6.832	8.159	1.42E-08	3.21E-06	
5	P21333FLNA	61	-0.27	5.16	5.43	4.73	6.12	2.45	1752	1.99E-08	3.39E-06	9.75	0	3.96E+07	3.09E+07	3.85E+07	2.78E+07	3.84E+09	3.19E+09	3.96E+09	9.97E+08	-0.966	0.237	-0.499	-0.436	5.177	5.363	5.425	4.679	-0.266	5.161	5.427	1.99E-08	3.39E-06	
6	Q02818NLCB1	23	1.10	5.25	4.16	3.95	4.76	3.18	1544	6.22E-08	8.46E-06	8.79	0	1.46E+08	4.50E+07	1.17E+08	8.63E+07	4.07E+09	2.80E+09	3.69E+09	1.51E+09	1.444	0.752	1.104	1.093	5.262	5.158	5.318	5.281	1.098	5.255	4.156	6.22E-08	8.46E-06	
7	P98160PGBM	56	-2.47	5.45	7.92	6.96	8.88	2.81	1941	1.54E-07	1.62E-05	7.32	2	NA	NA	1.08E+07	5.58E+06	5.24E+09	3.56E+09	4.99E+09	1.13E+09	NA	NA	-2.341	-2.598	5.639	5.534	5.775	4.863	-2.469	5.453	7.922	1.54E-07	1.62E-05	
8	Q9UBPAIDKK3	5	-1.38	2.93	4.31	3.61	5.01	0.78	1382	1.67E-07	1.62E-05	7.92	0	1.42E+07	1.00E+07	2.13E+07	1.74E+07	7.51E+08	5.31E+08	7.70E+08	4.31E+08	-1.784	-1.310	-1.358	-1.064	2.743	2.575	2.941	3.460	-1.379	2.930	4.309	1.67E-07	1.62E-05	
9	P33151CAOH5	14	-1.92	4.44	6.36	5.40	7.31	2.32	1555	7.48E-07	5.65E-05	6.24	2	NA	NA	1.49E+07	8.92E+06	2.51E+09	2.34E+09	2.08E+09	5.72E+08	NA	NA	-1.875	-1.965	4.545	4.878	4.447	3.872	-1.920	4.436	6.356	7.48E-07	5.65E-05	
10	Q8NBS9TND5	7	-1.37	3.62	4.99	4.13	5.84	1.48	1339	6.70E-07	5.65E-05	6.60	1	2.26E+07	7.70E+06	2.23E+07	NA	1.72E+09	1.18E+09	1.95E+09	2.91E+08	-1.143	-1.673	-1.290	NA	3.980	3.817	3.791	2.891	-1.369	3.620	4.969	6.70E-07	5.65E-05	
11	P02545ILMNA	21	-1.52	3.46	4.98	4.09	5.86	1.33	1292	8.88E-07	6.04E-05	6.36	1	1.72E+07	NA	1.21E+07	2.03E+07	1.09E+09	1.05E+09	9.39E+08	4.96E+08	-1.516	NA	-2.179	-0.859	3.302	3.628	3.241	3.663	-1.518	3.458	4.976	8.88E-07	6.04E-05	
12	Q5V7EQIEF1A3	7	-3.01	1.90	4.91	4.00	5.81	-0.21	1244	1.20E-06	7.42E-05	6.09	1	4.94E+06	3.09E+06	NA	4.59E+06	4.47E+06	4.80E+08	4.53E+08	8.28E+07	-3.245	-2.928	NA	-2.860	1.969	2.419	2.137	1.062	-3.011	1.897	4.908	1.20E-06	7.42E-05	
13	P07737PPOF1A	6	-0.02	4.32	4.34	3.44	5.25	2.15	1079	1.47E-06	8.33E-05	5.89	0	2.79E+07	3.29E+07	9.59E+07	2.88E+07	2.27E+09	1.90E+09	2.10E+09	5.75E+08	-0.848	0.319	0.822	-0.384	4.392	4.556	4.459	3.879	-0.023	4.321	4.344	1.47E-06	8.33E-05	
14	P19022CADH2	11	-3.96	0.78	4.74	3.91	5.57	-0.80	1332	2.23E-06	1.08E-04	5.39	2	2.82E+06	NA	3.66E+06	NA	1.95E+08	1.81E+08	2.10E+08	5.64E+07	-4.023	NA	-3.903	NA	0.733	0.902	0.970	0.504	-3.963	0.777	4.740	2.23E-06	1.08E-04	
15	P62857RS28	2	-2.52	2.12	4.64	3.82	5.45	0.57	1334	2.22E-06	1.08E-04	5.40	2	9.54E+06	NA	8.36E+06	NA	4.78E+08	3.94E+08	4.34E+08	1.82E+08	-2.335	NA	-2.708	NA	2.071	2.110	2.073	2.207	-2.072	2.115	4.637	2.22E-06	1.08E-04	
16	Q12841FSTL1	4	-4.10	3.49	7.60	6.46	8.74	1.97	1612	2.39E-06	1.08E-04	4.60	3	NA	NA	NA	1.82E+06	1.18E+09	9.34E+08	1.21E+09	4.40E+08	NA	NA	NA	-4.105	3.412	3.452	3.621	-4.105	3.494	7.599	2.39E-06	1.08E-04		
17	P10909CLLUS	12	-3.51	5.43	8.94	7.54	10.33	3.64	1548	3.07E-06	1.30E-04	4.46	3	NA	NA	4.80E+06	NA	3.97E+09	4.45E+09	4.72E+09	1.17E+09	NA	NA	-3.511	NA	5.226	5.879	5.690	4.907	-3.511	5.425	8.937	3.07E-06	1.30E-04	
18	P09382LEG1	4	2.55	5.15	2.61	2.00	3.22	3.85	9.62	3.95E-06	1.58E-04	4.93	0	2.90E+08	1.37E+08	3.33E+08	3.27E+08	3.38E+09	2.73E+09	3.41E+09	1.54E+09	2.390	2.285	2.625	2.888	4.985	5.121	5.198	5.312	2.547	5.154	2.607	3.95E-06	1.58E-04	
19	P06601MYL6	5	-2.65	3.33	5.98	4.54	7.42	0.34	935	5.00E-06	1.86E-04	4.69	0	5.62E+06	1.78E+06	8.80E+07	3.21E+06	8.59E+08	9.11E+08	1.02E+09	4.68E+08	-3.067	-3.685	-0.517	-3.345	3.414	3.373	3.580	-2.853	3.327	5.981	5.00E-06	1.86E-04		
20	zzlY-FGC2Cont004401	26	-2.43	4.82	7.25	5.64	8.87	1.71	1032	5.18E-06	1.86E-04	4.73	1	6.53E+06	NA	3.73E+07	2.15E+06	2.78E+09	2.48E+09	2.87E+09	1.01E+09	-2.860	NA	-0.545	-3.886	4.696	4.970	4.936	4.694	-2.430	4.824	7.254	5.18E-06	1.86E-04	
21	Q14181DAG1	10	-2.26	1.37	3.63	2.80	4.47	-0.19	996	6.83E-06	2.32E-04	4.47	1	7.68E+06	3.89E+06	1.87E+07	NA	2.99E+08	2.41E+08	2.92E+08	9.69E+07	-2.635	-2.610	-1.541	NA	1.372	1.349	1.469	1.291	-2.262	1.370	3.632	6.83E-06	2.32E-04	
22	P68371TBB4B	8	-4.09	0.22	4.30	3.31	5.30	-1.22	1016	1.47E-05	4.59E-04	3.76	2	2.12E+06	NA	NA	2.35E+06	1.02E+08	1.13E+08	1.30E+08	6.42E+07	-4.416	NA	NA	-3.762	-0.239	0.166	0.241	0.692	-4.089	0.215	4.304	1.47E-05	4.59E-04	
23	P68404IGABT	2	-2.73	0.31	3.04	2.26	3.81	-1.00	900	1.49E-05	4.59E-04	3.71	1	6.44E+06	2.84E+06	1.13E+07	NA	1.36E+08	1.01E+08	1.58E+08	5.59E+07	-2.880	-3.044	-2.271	NA	0.193	0.000	0.537	0.492	-2.731	0.305	3.037	1.49E-05	4.59E-04	
24	P21810PGS1	10	-3.57	5.20	8.76	6.97	10.56	3.45	1183	1.58E-05	4.66E-04	3.40	3	NA	NA	4.62E+06	NA	5.32E+09	3.52E+09	4.25E+09	6.63E+08	NA	NA	-3.566	NA	5.664	5.515	5.530	4.085	-3.566	5.199	8.765	1.58E-05	4.66E-04	
25	P26022PTX3	11	-4.13	4.80	8.93	7.02	10.83	3.01	1136	2.02E-05	5.29E-04	3.22	3	NA	NA	NA	1.79E+06	3.52E+09	3.74E+09	2.79E+09	4.96E+08	NA	NA	NA	NA	5.045	5.607	4.886	3.662	-4.127	4.800	8.927	2.02E-05	5.29E-04	
26	P29401TKT	9	-1.71	1.23	2.94	2.16	3.71	-0.03	865	2.00E-05	5.29E-04	3.42	1	NA	NA	8.84E+06	1.82E+07	8.39E+06	2.50E+08	2.00E+08	2.22E+08	1.33E+08	-1.484	-1.584	-2.048	1.102	1.056	1.014	1.748	-1.705	1.230	2.935	2.00E-05	5.29E-04	
27	P305301UFO	3	-4.61	1.49	6.10	4.80	7.39	0.27	1139	1.99E-05	5.29E-04	3.23	3	NA	NA	NA	1.26E+06	3.22E+08	2.72E+08	2.22E+08	1.46E+08	NA	NA	NA	NA	4.127	5.045	5.607	4.886	3.662	-4.127	4.800	8.927	2.02E-05	5.29E-04
28	zzlY-FGC2Cont000871	15	-3.09	4.00	7.08	5.22	8.95	1.64	891	3.58E-05	9.01E-04	2.93	2	NA	NA	2.15E+06	1.26E+07	1.60E+09	1.33E+09	1.68E+09	6.20E+08	NA	NA	-4.671	-1.503	3.874	4.007	4.121	3.987	-3.087	3.997	7.084	3.58E-05	9.01E-04	
29	Q149791HNRDL	3	-3.23	0.06	2.39	1.69	3.09	-0.96	786	4.09E-05	9.94E-04	2.70	1	NA	4.27E+06	1.10E+07	7.59E+06	1.55E+08	1.09E+08	1.01E+08	3.70E+07	NA	NA	-2.484	-2.314	-2.183	0.386	0.109	-0.137	-0.107	-2.327	0.063	2.390	4.08E-05	9.94E-04
30	Q15904IVAS1	4	-1.10	2.22	3.31	2.26	4.36	0.56	711	4.76E-05	1.12E-03	2.42	0	4.87E+07	1.69E+07	2.12E+07	6.74E+06	5.01E+06	4.61E+08	4.85E+08	1.74E+08	-0.080	-0.598	-1.360	-2.343	2.139	2.357	2.238	2.139	-1.095	2.218	3.313	4.76E-05	1.12E-03	
31	P319481STP1	14	1.45	3.83	2.38	1.61	3.14	2.64	701	5.36E-05	1.22E-03	2.30	0	1.77E+08	4.93E+07	2.07E+08	9.86E+07	1.16E+09	1.35E+09	1.27E+09	7.13E+08														

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN	M_3_Co ntrol.ra w	M_2_Co ntrol.ra w	M_4_Co ntrol.ra w	M_1_Co ntrol.ra w	E4.2_Gr oup_1r aw	E4.4_Gr oup_1r aw	E4.3_Gr oup_1r aw	E4.1_Gr oup_1r aw	M_3_C ontr.transf	M_2_C ontr.transf	M_4_C ontr.transf	M_1_C ontr.transf	E4.2_Gr oup_1r aw	E4.4_Gr oup_1r aw	E4.3_Gr oup_1r aw	E4.1_Gr oup_1r aw	pseud o.Ctrl	pseud o.Group pl	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val		
49	P35241FADI	8	-3.74	-1.31	2.43	1.52	3.34	-2.35	6.11	0.000247	3.50E-03	0.87	3	3.28E+06	1.31E+06	1.70E+07	3.33E+06	3.07E+07	5.56E+07	NA	4.16E+08	4.12E+08	4.63E+08	1.06E+08	-0.302	0.036	NA	1.862	2.182	2.186	1.444	-0.013	1.919	1.932	3.80E-04	4.97E-03
50	P61091UBC12	4	-3.06	-0.39	2.68	1.61	3.74	-1.73	5.66	0.000277	3.84E-03	0.61	0	2.94E+06	2.06E+06	1.70E+07	3.77E+06	8.79E+07	7.83E+07	7.29E+07	3.79E+07	-3.962	-3.463	-1.682	-3.126	-0.455	-0.388	-0.635	-0.074	-3.063	-0.368	2.676	2.77E-04	3.84E-03		
51	Q9Y490TLN1	13	-4.52	1.16	5.68	3.79	7.58	0.02	7.25	0.000283	3.95E-03	1.01	3	NA	NA	NA	1.34E+06	3.24E+08	2.79E+08	3.26E+08	3.82E+07	NA	NA	NA	-4.523	1.489	1.578	1.637	-0.060	-4.523	1.161	5.684	2.63E-04	3.95E-03		
52	P09486ISFRC	6	-1.13	5.76	6.89	4.19	9.59	2.81	5.95	0.000336	4.46E-03	0.56	1	2.61E+06	NA	1.60E+08	2.09E+07	7.37E+09	5.57E+09	6.61E+09	8.57E+08	-4.130	NA	1.562	-0.819	6.149	6.228	6.201	4.459	-1.123	5.759	6.888	3.36E-04	4.46E-03		
53	P61978HHPFK	9	-0.01	1.92	1.93	1.16	2.70	1.09	5.74	0.00039	4.97E-03	0.43	1	4.15E+07	3.07E+07	5.56E+07	NA	4.16E+08	4.12E+08	4.63E+08	1.06E+08	-0.302	0.036	NA	1.862	2.182	2.186	1.444	-0.013	1.919	1.932	3.80E-04	4.97E-03			
54	Q9Y220SSG T1	2	-2.40	0.06	2.46	1.47	3.44	-0.99	5.71	0.000394	5.05E-03	0.39	1	8.22E+06	3.77E+06	1.37E+07	NA	1.07E+08	7.63E+07	9.83E+07	8.07E+07	-2.541	-2.654	-1.932	NA	-0.158	-0.440	-0.161	1.024	-2.396	0.062	2.457	3.94E-04	5.05E-03		
55	Q94996ICSTN1	7	-1.32	1.81	3.19	1.87	4.39	0.47	5.69	0.000406	5.11E-03	0.36	3	3.49E+07	8.89E+06	1.42E+07	NA	5.66E+08	4.27E+08	4.55E+08	5.75E+07	-0.541	-1.478	-1.945	NA	2.323	2.237	2.143	3.460	0.533	-1.321	1.909	3.130	4.06E-04	5.11E-03	
56	Q9B9E3ITBA1C	6	-0.59	2.96	3.45	1.99	4.91	1.19	5.32	0.000435	5.36E-03	0.14	0	5.58E+07	2.35E+07	5.80E+07	6.30E+06	8.06E+08	1.09E+09	1.20E+08	0.110	-0.139	0.095	-2.435	3.148	3.224	3.460	1.601	-0.952	2.958	3.450	4.35E-04	5.36E-03			
57	Q96124IFTN1	5	-2.00	0.47	2.47	1.40	3.54	-0.77	5.20	0.000508	6.16E-03	-0.02	0	1.26E+07	4.32E+06	2.22E+07	6.99E+06	2.58E+08	2.30E+08	1.02E+09	2.96E+07	-1.955	-2.467	-1.295	-2.295	1.150	1.273	-0.131	-0.432	-2.003	0.465	2.468	5.08E-04	6.16E-03		
58	Q14352ICALLU	8	-3.67	-1.34	2.33	1.35	3.30	-2.34	5.49	0.000519	6.17E-03	0.11	2	2.53E+06	1.61E+06	1.77E+06	NA	6.19E+07	4.34E+07	5.39E+07	1.02E+07	-4.172	-3.908	-2.931	NA	-0.977	-1.315	-1.092	-1.985	-3.670	-1.342	2.329	5.19E-04	6.17E-03		
59	P14628IKPYM	14	2.09	4.00	1.91	1.07	2.76	3.04	5.08	0.000598	6.66E-03	-0.19	0	1.74E+08	1.21E+08	4.29E+08	1.23E+08	1.39E+09	1.16E+09	1.91E+09	7.25E+08	1.695	2.111	2.965	1.569	3.662	3.810	4.316	4.215	2.088	4.001	1.913	5.59E-04	6.66E-03		
60	P23226I0A2	6	-2.25	0.26	2.51	1.39	3.63	-0.99	5.06	0.000617	6.66E-03	-0.22	0	1.64E+07	3.28E+06	2.24E+07	3.38E+06	1.69E+08	1.64E+08	1.22E+09	3.12E+07	-1.588	-2.848	-1.282	-3.275	0.509	0.746	0.146	-0.365	-2.248	0.262	2.510	6.17E-04	6.66E-03		
61	P30101P0A3	6	-3.95	-0.63	3.31	1.91	4.72	-2.05	5.40	0.000676	6.66E-03	0.00	1	1.52E+06	1.74E+06	5.79E+06	NA	1.19E+08	9.93E+07	8.34E+07	9.63E+06	-4.981	-3.720	-3.240	NA	-0.014	-0.031	-0.430	-2.064	-3.947	-0.635	3.312	5.76E-04	6.66E-03		
62	Q96AE4I0UBP1	7	-4.00	0.10	4.10	2.43	5.77	-1.26	5.75	0.000608	6.66E-03	0.12	2	6.74E+06	6.00E+06	NA	NA	1.16E+08	1.65E+08	1.38E+08	2.56E+07	-2.916	-5.179	NA	NA	-0.038	0.759	0.329	-0.644	-3.999	0.102	4.099	6.08E-04	6.66E-03		
63	Q9Y798ACDN	12	1.89	4.99	3.10	1.73	4.47	3.44	5.09	0.000591	6.66E-03	-0.17	0	2.64E+08	1.16E+08	5.16E+08	3.83E+07	3.09E+09	2.88E+09	3.39E+09	1.03E+09	2.262	2.036	3.258	0.000	4.834	5.205	5.184	4.729	3.989	3.099	5.91E-04	6.66E-03			
64	Q12904IAMP1	3	-4.14	-1.65	2.49	1.38	3.60	-2.89	5.05	0.000629	6.69E-03	-0.24	0	2.25E+06	7.18E+05	2.83E+06	4.10E+06	2.72E+07	6.02E+07	2.52E+07	1.59E+07	-4.336	-4.932	-4.276	-3.014	-2.206	-0.810	-2.247	-1.336	-4.140	-1.650	2.490	6.29E-04	6.69E-03		
65	Q16811ITSN1	5	-0.48	-1.93	-1.45	-2.11	-0.80	-1.20	-5.01	0.000665	6.96E-03	-0.30	0	2.79E+07	1.81E+07	3.76E+07	3.79E+07	2.73E+07	3.12E+07	3.47E+07	1.06E+07	-0.849	-0.503	-0.534	-0.015	-2.200	-1.824	-1.759	-1.928	-0.475	-1.928	-1.452	6.65E-04	6.96E-03		
66	Q196FK9ICAB45	10	-0.61	-2.11	2.92	1.77	4.06	1.53	6.17	0.000697	7.18E-03	0.17	3	NA	1.44E+07	NA	NA	4.95E+08	4.04E+08	4.73E+08	1.54E+08	NA	-0.810	NA	NA	2.122	2.162	2.201	1.963	-0.810	2.109	2.919	6.97E-04	7.18E-03		
67	P69459ICAF1	2	-5.47	-2.34	3.13	1.87	4.40	-2.96	6.00	0.000815	8.27E-03	0.01	3	NA	NA	1.24E+06	NA	2.91E+07	1.70E+07	2.39E+07	9.42E+06	NA	NA	-5.469	NA	-2.157	-2.770	-2.324	-2.096	-5.469	-2.337	3.132	8.16E-04	8.27E-03		
68	Q162701BP7	6	-2.08	5.55	7.63	4.51	10.74	4.02	9.33	0.000868	8.69E-03	-0.05	3	NA	NA	8.20E+06	8.25E+09	6.41E+09	6.65E+09	3.67E+08	NA	NA	NA	-2.080	6.317	6.445	6.210	3.225	-2.080	5.549	7.629	8.69E-04	8.69E-03			
69	P10909ICHO	14	3.00	4.34	1.94	0.71	1.97	3.67	4.79	0.000908	8.82E-03	-0.62	0	4.02E+08	1.99E+08	5.90E+08	3.38E+08	1.89E+09	1.70E+09	1.97E+09	8.67E+08	2.844	2.766	3.452	2.934	4.123	4.381	4.364	4.475	2.999	4.336	1.337	9.08E-04	8.82E-03		
70	P28838IAMPL	5	-2.84	-0.48	2.37	1.29	3.44	-1.49	5.04	0.000906	8.82E-03	-0.47	1	7.81E+06	1.70E+06	1.21E+07	NA	7.03E+07	6.87E+07	1.30E+08	2.35E+07	-2.612	-3.746	-2.170	NA	-0.788	-0.602	0.247	-0.767	-2.843	-0.477	2.365	9.06E-04	8.82E-03		
71	P38646IGRP75	24	2.74	4.65	1.91	1.00	2.82	3.70	4.72	0.001004	8.82E-03	-0.72	0	5.05E+08	1.03E+08	6.26E+08	2.24E+08	2.76E+09	2.58E+09	2.30E+09	7.60E+08	3.156	1.984	3.138	2.379	4.683	5.034	4.603	4.284	2.739	4.641	1.912	1.00E-03	9.62E-03		
72	P62760V1SL1	3	-1.22	-4.12	-2.90	-4.20	-1.60	-2.67	-5.23	0.00107	9.97E-03	-0.49	2	2.18E+07	1.27E+07	1.98E+07	NA	1.32E+07	8.85E+06	3.40E+06	NA	-1.195	-0.991	-1.463	NA	-3.295	-3.788	-5.284	NA	-1.216	-4.119	-2.902	1.07E-03	9.97E-03		
73	Q130521LPP	2	-0.82	-2.26	-1.44	-2.11	-0.77	-1.44	-4.91	0.001069	9.97E-03	-0.64	4	3.08E+07	1.96E+07	2.54E+07	2.17E+07	2.19E+07	2.59E+07	2.55E+07	NA	-0.716	-1.028	-1.099	-0.766	-2.174	-2.377	-2.226	NA	-0.821	-2.159	-1.439	1.07E-03	9.97E-03		
74	Q143391TFD54	6	-1.98	-0.06	1.91	0.99	2.84	-1.02	4.66	0.001094	1.01E-02	-0.81	0	1.98E+07	6.92E+06	1.48E+07	4.51E+06	9.27E+07	1.32E+08	7.44E+07	4.95E+07	-1.325	-1.821	-1.877	-2.886	-0.376	0.411	-0.604	0.315	-1.977	-0.063	1.914	1.09E-03	1.01E-02		
75	P20908IC05A1	12	2.55	4.47	2.22	1.22	3.22	1.73	5.19	0.001105	1.01E-02	-0.50	2	NA	NA	5.14E+07	5.89E+07	6.78E+08	6.28E+08	5.90E+08	1.48E+08	NA	NA	-0.080	0.577	2.591	2.835	2.538	1.909	0.248	2.468	2.200	1.12E-03	1.01E-02		
76	Q9Y524IHEBP2	5	-3.59	-1.64	1.95	1.06	2.83	-2.29	5.16	0.001154	1.03E-02	-0.54	2	4.50E+06	NA	NA	2.27E+06	3.86E+07	3.57E+07	3.07E+07	1.60E+07	-3.375	NA	NA	-3.808	-1.682	-1.618	-1.948	-1.322	-3.992	-1.643	1.949	1.15E-03	1.03E-02		
77	O14737PDCD5	7	0.35	1.99	1.64	0.83	2.44	1.17	4.59	0.001208	1.07E-02	-0.91	0	5.04E+07	2.06E+07	1.02E+08	7.18E+07	4.86E+08	3.32E+08	3.42E+08	1.63E+08	-0.031	-0.325	0.916	0.846	2.095	2.105	1.711	2.050	0.352	1.990	1.638	1.21E-03	1.07E-02		
78	O75874IDHC	7	0.36	1.76	1.40	0.71	2.09	1.06	4.95	0.001282	1.09E-02	-0.97	0	5.29E+07	3.12E+07	1.09E+08	4.20E+07	3.67E+08	2.98E+08	4.10E+08	1.27E+08	0.036														

ProteinName	# Peptid	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrAs	M3_Co ntrol.ra w	M2_Co ntrol.ra w	M4_Co ntrol.ra w	M1_Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3_C ontrol. transf	M.2_C ontrol. transf	M.4_C ontrol. transf	M.1_C ontrol. transf	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	pseud o. Ctrl	pseud o. Group pt	pseud o. log2 FC	pseud o. P. Value	pseud o. adj .P.Val
P28072PFSB6	2	-0.91	0.23	1.14	0.50	1.78	-0.34	4.00	0.00293	2.10E-02	-1.83	0	2.30E+07	1.24E+07	2.66E+07	2.73E+07	1.19E+08	1.12E+08	1.33E+08	5.60E+07	-1.18	-1.017	-1.032	-0.456	-0.001	0.150	0.278	0.494	-0.906	0.230	1.136	2.93E-03	2.10E-02
Q9LHG2PFSK1	2	-4.29	-6.59	-2.30	-3.48	-1.11	-5.21	-4.69	0.002965	2.10E-02	-1.36	3	1.49E+06	NA	3.85E+06	1.78E+06	1.69E+06	NA	1.24E+06	NA	-4.92	NA	-3.829	-4.138	-6.359	NA	-6.817	NA	-4.290	-6.598	-2.298	2.96E-03	2.10E-02
Q1571EALVY1	3	-4.43	-2.60	1.83	0.84	2.81	-3.21	4.35	0.003068	2.15E-02	-1.55	2	2.64E+06	NA	2.05E+06	NA	2.76E+07	2.14E+07	1.57E+07	5.86E+06	-4.712	NA	-4.744	NA	-2.104	-2.415	-3.024	-2.795	-4.428	-2.502	1.826	3.07E-03	2.15E-02
P86771AFRP1	3	2.21	0.67	-1.64	-2.57	-0.70	1.39	-3.94	0.003211	2.23E-02	-1.52	2	2.68E+08	8.04E+07	3.13E+08	2.39E+08	1.17E+08	9.99E+07	2.14E+08	1.00E+08	2.279	1.548	2.534	2.461	-0.035	-0.026	1.001	1.339	2.205	0.570	-1.636	3.21E-03	2.23E-02
F06733IENDA	15	2.68	4.97	2.28	0.97	3.60	3.82	3.92	0.003314	2.29E-02	-1.95	0	3.94E+08	1.95E+08	9.66E+08	8.15E+07	3.65E+09	3.62E+09	1.03E+09	2.790	2.764	4.855	1.017	5.099	4.756	5.290	4.779	2.692	4.566	2.384	3.31E-03	2.29E-02	
zz1-FGC2Cant00285f	21	2.95	0.68	-2.27	-3.58	-0.95	1.82	-3.89	0.003495	2.37E-02	-2.00	0	2.87E+08	2.77E+08	1.02E+08	7.87E+07	4.22E+08	1.03E+08	2.375	3.217	3.535	2.675	-0.230	-0.452	2.028	1.380	2.950	0.682	-2.263	3.49E-03	2.37E-02		
P37802TLAGL2	9	3.89	5.13	1.23	0.51	1.96	4.51	3.83	0.003615	2.57E-02	-2.10	0	9.59E+08	4.96E+08	1.09E+09	3.95E+08	3.71E+09	3.00E+09	3.85E+09	1.27E+09	4.046	4.347	4.338	3.145	5.126	5.269	5.064	5.031	3.894	5.126	1.244	3.82E-03	2.57E-02
Q75044ISRGP2	2	-1.69	-3.01	-1.32	-2.08	-0.55	-2.44	-3.96	0.003907	2.60E-02	-1.98	1	1.69E+07	6.50E+06	1.77E+07	NA	1.39E+07	1.13E+07	1.74E+07	6.65E+06	-1.542	-1.906	-1.621	NA	-3.212	-3.403	-2.803	-2.603	-1.690	-3.005	-1.316	3.91E-03	2.60E-02
REV_Q8TUCU4IALMS1	2	-1.84	-3.87	-2.04	-3.25	-0.83	-2.71	-3.86	0.004529	2.86E-02	-2.14	1	2.17E+07	3.21E+06	1.79E+07	1.11E+07	NA	5.39E+06	7.52E+06	5.12E+06	-1.199	-2.874	-1.609	-1.666	NA	-4.562	-4.079	-2.983	-1.837	-3.674	-2.037	4.53E-03	2.96E-02
Q14019COTL1	4	0.12	2.18	2.05	0.81	3.30	1.15	3.73	0.004446	2.86E-02	-2.25	0	1.97E+07	4.82E+07	1.41E+08	2.82E+07	4.79E+08	4.12E+08	4.57E+08	1.94E+08	-1.333	0.647	1.391	-0.413	2.071	2.182	2.148	2.300	1.202	2.175	2.055	4.45E-03	2.96E-02
Q16775IGLD2	3	-0.77	0.94	1.71	0.67	2.75	0.08	3.72	0.004534	2.86E-02	-2.27	0	4.79E+07	7.95E+06	5.32E+07	1.43E+07	2.09E+08	3.01E+08	1.72E+08	5.95E+07	-0.102	-1.629	-0.030	-1.325	0.840	1.693	0.666	0.557	-0.771	0.939	1.710	4.53E-03	2.96E-02
Q82747AFAC1A	2	-2.92	-0.56	2.35	1.04	3.67	-1.03	4.33	0.004438	2.86E-02	-1.67	3	NA	NA	NA	7.03E+07	6.16E+07	1.09E+08	2.52E+07	NA	-2.915	NA	NA	-0.798	-0.772	-0.025	-0.664	-2.915	-0.562	2.353	4.44E-03	2.96E-02	
Q82777SYN2	3	-3.83	-1.71	2.12	0.89	3.35	-2.77	4.05	0.00446	2.86E-02	-1.98	2	1.77E+06	1.36E+06	7.99E+06	NA	3.71E+07	3.13E+07	3.94E+07	5.99E+07	-4.667	-4.056	-2.774	NA	-1.741	-1.823	-1.569	NA	-3.832	-1.711	2.121	4.46E-03	2.96E-02
Q82945FLBP2	9	-2.17	-0.22	1.95	0.78	3.13	-1.19	3.75	0.004343	2.86E-02	-2.23	0	2.46E+07	2.43E+06	7.68E+06	1.20E+07	8.80E+07	7.98E+07	7.80E+07	4.57E+08	-1.026	-3.257	-2.831	-1.570	-0.453	-0.371	-0.531	0.491	-2.171	-0.216	1.955	4.34E-03	2.96E-02
Q19Y209IAKAP2	5	-4.04	-1.67	2.38	1.04	3.72	-2.14	4.29	0.004632	2.89E-02	-1.71	3	NA	NA	3.33E+06	NA	3.08E+07	3.17E+07	5.31E+07	1.22E+07	NA	NA	-0.401	NA	-2.017	-1.807	-1.115	-1.721	-4.041	-1.665	2.376	4.63E-03	2.89E-02
P62820RAB1A	2	-3.28	-1.18	2.09	0.90	3.29	-2.02	4.25	0.004822	2.98E-02	-1.87	3	NA	2.29E+06	3.53E+06	6.15E+07	NA	6.98E+07	NA	-3.941	NA	-3.215	-0.989	NA	-0.701	-1.860	-2.718	-1.183	2.095	4.82E-03	2.98E-02		
P30046IDCPD	4	-4.86	-1.03	3.83	1.60	6.06	-1.80	4.17	0.005339	3.24E-02	-1.85	3	NA	NA	1.89E+06	NA	8.02E+07	8.86E+07	6.56E+07	7.02E+06	NA	NA	-4.859	NA	-0.591	-0.207	-0.793	-2.524	-4.859	-1.029	3.830	5.34E-03	3.24E-02
Q13163PAK1	13	0.84	2.60	1.76	0.67	2.86	1.72	3.62	0.005298	3.24E-02	-2.43	0	1.20E+08	4.78E+07	3.81E+07	1.54E+08	5.84E+08	7.08E+08	4.85E+08	2.42E+08	1.165	0.833	-0.512	1.876	2.538	3.022	2.239	2.621	0.841	2.605	1.764	5.30E-03	3.24E-02
zz1-FGC2Cant00466f	5	-1.83	-3.67	-1.83	-2.94	-0.73	-2.75	-3.91	0.005419	3.26E-02	-2.18	2	1.33E+07	5.35E+06	1.99E+07	NA	1.29E+07	5.59E+06	1.35E+07	NA	-1.873	-2.174	-1.495	NA	-3.310	-4.506	-3.190	NA	-1.834	-3.659	-1.835	5.42E-03	3.26E-02
zz1-FGC2Cant00037f	9	2.38	3.72	1.33	0.50	2.17	3.05	3.59	0.005561	3.29E-02	-2.49	0	3.36E+08	9.66E+07	4.46E+08	1.81E+08	1.99E+09	1.20E+09	1.25E+09	3.49E+08	2.594	1.801	3.048	2.095	4.195	3.845	3.680	3.154	2.395	3.718	1.334	5.66E-03	3.29E-02
zz1-FGC2Cant00365f	15	-0.37	-0.99	1.36	0.51	2.22	0.31	3.59	0.005545	3.29E-02	-2.48	0	5.43E+07	2.50E+07	3.95E+07	1.76E+07	3.01E+08	1.17E+08	2.24E+08	9.71E+07	0.072	-0.056	-0.460	-1.046	1.380	0.229	1.066	1.294	-0.372	0.922	1.364	5.55E-03	3.29E-02
P78352DLD4	2	-2.02	-2.84	-2.72	-4.33	-1.12	-1.21	-4.10	0.005734	3.29E-02	-2.05	3	7.83E+07	1.00E+07	7.71E+07	NA	1.99E+07	NA	1.52E+07	NA	0.578	-1.915	0.989	NA	-2.670	NA	-3.012	NA	-0.176	-2.841	-2.725	5.74E-03	3.36E-02
Q12M20AAK1	9	2.35	1.21	-1.14	-1.85	-0.42	1.78	-3.56	0.005797	3.37E-02	-2.52	0	4.26E+08	1.12E+08	2.32E+08	2.23E+08	2.09E+08	2.06E+08	3.03E+08	1.04E+08	2.923	2.004	2.102	2.371	0.839	1.102	1.527	1.390	2.300	1.214	-1.135	5.80E-03	3.37E-02
Q17451MYLK	4	0.02	-1.39	-1.41	-2.29	-0.53	-0.79	-3.68	0.005893	3.40E-02	-2.41	1	5.25E+07	2.65E+07	NA	3.88E+07	2.77E+07	5.62E+07	4.8E+07	2.01E+07	0.024	0.024	NA	0.008	-2.176	-0.916	-1.487	-0.998	-0.219	-1.394	-1.413	5.80E-03	3.40E-02
P18065IBP2	2	-1.42	0.71	2.13	0.81	3.44	0.00	3.80	0.006199	3.53E-02	-2.28	2	NA	NA	1.06E+07	2.69E+07	2.11E+08	1.46E+08	1.50E+08	7.63E+07	NA	NA	-2.360	-0.475	0.853	0.964	0.461	0.956	-1.417	0.708	2.126	6.20E-03	3.53E-02
Q19BPX9IARF5L	3	-1.74	0.11	1.84	0.68	3.00	-0.68	3.64	0.00623	3.53E-02	-2.46	1	1.67E+07	NA	NA	1.07E+07	1.47E+07	1.72E+08	7.89E+07	2.61E+07	-1.558	NA	-2.356	-1.296	0.547	1.016	-0.522	-0.617	-1.736	0.106	1.842	6.20E-03	3.53E-02
P14550IAKA1	3	-3.72	-1.60	2.12	0.84	3.41	-2.02	3.99	0.006546	3.62E-02	-2.06	3	NA	NA	4.44E+06	NA	4.90E+07	3.91E+07	4.10E+07	9.50E+06	NA	NA	-3.724	NA	-1.327	-1.480	-1.505	-2.084	-3.724	-1.599	1.255	6.55E-03	3.62E-02
Q14193IDPYL3	19	2.78	4.03	1.25	0.44	2.06	3.40	3.49	0.006551	3.62E-02	-2.65	0	4.57E+08	1.56E+08	6.00E+08	1.90E+08	2.08E+09	1.44E+09	1.68E+09	4.74E+08	3.020	2.462	3.477	2.154	4.259	4.128	4.127	3.599	2.778	4.028	1.250	6.55E-03	3.62E-02
Q16643DREB	3	-2.17	-1.08	1.10	0.40	1.80	-1.95	3.60	0.006548	3.62E-02	-2.52	1	1.11E+07	6.23E+06	1.01E+07	NA	4.80E+07	5.53E+07	6.50E+07	2.03E+07	-2.124	-1.964	-2.474	-1.356	-0.939	-1.035	-0.980	-2.175	-1.078	1.097	6.55E-03	3.62E-02	
P19139IPUR8	5	-4.91	-1.54	3.37	1.25	5.49	-2.66	3.73	0.006819	3.74E-02	-2.37	2	NA	NA	4.00E+06	4.31E+05	5.91E+07	5.49E+07	6.34E+07	4.06E+06	NA	NA	-3.774	-6.050	-1.046	-0.952	-0.846	-3.319	-4.912	-1.541	3.371	6.82E-03	3.74E-02
P67936ITPM4	5	-2.88	-0.31	2.57	0.97	4.17	-0.82	3.90	0.007326	3.99E-02	-2.18	3	NA	NA	7.45E+06	NA	5.84E+07	1.26E+08	1.09E+08	2.88E+07	NA	NA	-2.876	NA									

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrNAs	M3_Co ntrol.ra w	M2_Co ntrol.ra w	M4_Co ntrol.ra w	M1_Co ntrol.ra w	E4.2_Gr oup_1r aw	E4.4_Gr oup_1r aw	E4.3_Gr oup_1r aw	E4.1_Gr oup_1r aw	M3_C ontr.tra nsf	M2_C ontr.tra nsf	M4_C ontr.tra nsf	M1_C ontr.tra nsf	E4.2_Gr oup_1tran sf	E4.4_Gr oup_1tran sf	E4.3_Gr oup_1tran sf	E4.1_Gr oup_1tran sf	pseud o.Ctrl	pseud o.Group pl	pseud o.log2 FC	pseudo.P. Value	pseudo.ad j.P.Val	
143	Q9P265DIFP2B	2	-1.08	-3.04	-1.96	-3.35	-0.58	-1.73	-3.33	0.01886	5.69E-02	-2.95	2	2.89E+07	1.41E+07	4.34E+07	6.77E+06	NA	1.45E+07	1.47E+07	NA	NA	-0.801	-0.844	-0.325	-2.337	NA	-3.023	-3.058	NA	-1.077	-3.041	-1.964	1.19E-02	5.69E-02
144	Q96G7W7FPGCB	3	-0.28	-2.01	-1.73	-2.96	-0.51	-1.15	-3.32	0.01213	5.77E-02	-3.02	2	5.35E+07	1.59E+07	4.71E+07	NA	6.16E+07	2.12E+07	1.96E+07	NA	0.051	-0.689	-0.207	NA	0.239	-0.985	-2.429	-2.625	NA	-0.282	-2.013	-1.731	1.21E-02	5.77E-02
145	P0DF25ICALM3	5	2.24	0.61	-1.64	-2.84	-0.44	1.42	-3.07	0.012311	6.05E-02	-3.34	0	3.20E+08	5.95E+07	5.58E+08	1.62E+08	1.40E+08	1.11E+08	1.41E+08	1.25E+08	2.526	1.134	3.371	1.941	0.239	0.145	0.371	1.665	2.243	0.605	-1.638	1.29E-02	6.05E-02	
146	Q96YHHS6L1	3	-0.02	-1.48	-1.46	-2.51	-0.41	-0.75	-3.27	0.012867	6.05E-02	-3.08	2	5.33E+07	2.30E+07	5.65E+07	NA	7.21E+07	3.87E+07	2.50E+07	NA	0.045	-0.168	0.057	NA	-0.751	-1.495	-2.199	NA	-0.022	-1.482	-1.460	1.29E-02	6.05E-02	
147	Q96CVI1H4P	8	-1.94	0.28	2.21	0.80	3.62	-0.67	3.15	0.01303	6.07E-02	-3.22	1	3.15E+07	NA	2.13E+07	2.33E+06	1.55E+08	1.33E+08	8.59E+07	5.29E+07	-0.653	NA	-1.353	-3.776	0.400	0.423	-0.365	0.663	-1.937	0.275	2.213	1.30E-02	6.07E-02	
148	Q96FT31M1EP	2	1.60	0.61	-0.98	-1.71	-0.26	1.11	-3.04	0.013461	6.10E-02	-3.38	0	1.46E+08	8.29E+07	1.57E+08	1.48E+08	1.43E+08	1.15E+08	1.74E+08	5.79E+07	1.444	1.589	1.537	1.824	0.273	0.200	0.662	1.304	1.598	0.615	-0.963	1.35E-02	6.10E-02	
149	Q96L89DCHH	2	-1.13	-2.31	-1.18	-2.03	-0.33	-1.72	-3.24	0.013398	6.10E-02	-3.12	2	3.11E+07	8.77E+06	2.39E+07	NA	2.59E+07	2.13E+07	2.59E+07	NA	-0.701	-1.495	-1.190	NA	-2.295	-2.424	-2.208	NA	-1.129	-2.306	-1.177	1.34E-02	6.10E-02	
150	Q9UK49UJECIL2	8	-0.13	-1.94	-1.81	-3.14	-0.49	-1.17	-3.13	0.013414	6.10E-02	-3.25	1	5.50E+07	2.67E+07	3.79E+07	NA	4.59E+07	3.63E+07	4.99E+07	3.90E+06	0.098	0.035	-0.522	NA	-1.442	-1.593	-1.210	-3.533	-0.133	-1.945	-1.812	1.34E-02	6.10E-02	
151	zz1Y_FGC2Ccont00495l	13	1.76	3.98	2.12	0.55	3.68	2.92	3.04	0.013428	6.10E-02	-3.38	0	7.26E+07	2.65E+08	2.67E+08	6.59E+07	2.79E+08	1.40E+09	5.10E+08	8.43E+08	0.474	3.185	2.305	1.067	-1.442	4.078	2.317	4.435	1.763	3.878	2.115	1.34E-02	6.10E-02	
152	zz1Y_FGC2Ccont00256l	5	3.12	4.09	0.97	0.25	1.69	3.61	3.04	0.013632	6.10E-02	-3.38	0	5.33E+08	3.61E+08	4.94E+08	2.36E+08	1.57E+08	1.58E+08	1.70E+08	6.77E+08	3.232	3.613	3.195	2.452	3.844	4.270	4.145	4.116	3.123	4.094	0.971	1.35E-02	6.10E-02	
153	Q14011C1PBP	2	-2.41	-0.90	1.51	0.43	2.59	-1.50	3.38	0.013907	6.18E-02	-2.97	3	6.74E+06	NA	1.13E+07	NA	6.94E+07	6.28E+07	5.29E+07	NA	-2.917	NA	-2.002	NA	-0.830	-0.743	-1.124	NA	-2.409	-0.989	1.571	1.39E-02	6.18E-02	
154	Q9LKK3NIDJ15	3	4.43	-3.12	1.31	0.34	2.28	-3.68	3.10	0.01404	6.24E-02	-3.30	1	2.58E+06	1.12E+06	1.93E+06	NA	2.63E+07	1.04E+07	1.01E+07	4.88E+06	-4.145	-4.320	-4.828	NA	-2.253	-3.531	-3.639	-3.057	-4.431	-3.120	1.311	1.40E-02	6.24E-02	
155	Q75083wDR1	4	-3.59	-1.34	2.26	0.57	3.92	-2.31	3.08	0.014544	6.34E-02	-3.33	1	2.25E+06	1.59E+06	9.03E+06	NA	9.87E+07	2.00E+07	9.79E+07	7.79E+06	-4.336	-3.838	-2.597	NA	-0.283	-2.517	-0.192	-2.373	-3.591	-1.341	2.249	1.45E-02	6.34E-02	
156	P09937C05A2	10	-0.53	1.47	2.00	0.95	3.44	1.07	3.34	0.014618	6.34E-02	-2.88	3	NA	NA	3.77E+07	NA	2.67E+08	3.24E+08	2.04E+08	1.52E+08	NA	NA	-0.527	NA	1.204	1.805	0.923	1.942	-0.527	1.468	1.936	1.46E-02	6.34E-02	
157	P22314UBA1	9	-1.16	2.05	3.22	0.80	5.63	0.44	2.99	0.014959	6.34E-02	-3.46	0	1.70E+07	3.14E+07	1.28E+08	1.24E+06	4.42E+08	3.02E+08	5.52E+08	1.59E+08	-1.537	0.259	1.246	-4.626	1.953	1.701	2.541	2.011	-1.165	2.051	3.216	1.46E-02	6.34E-02	
158	Q9UPP5K1107	3	1.94	-0.25	-2.19	-3.75	-0.64	0.84	-3.57	0.014645	6.34E-02	-2.84	4	1.13E+08	1.70E+08	NA	NA	8.77E+07	9.83E+07	NA	NA	1.300	2.578	NA	NA	-0.458	-0.047	NA	NA	1.939	-0.263	-2.181	1.46E-02	6.34E-02	
159	Q16352AINX	4	-1.62	-3.09	-1.46	-2.95	-0.37	-2.46	-3.06	0.014987	6.41E-02	-3.35	1	1.89E+07	3.94E+06	NA	1.99E+07	1.41E+07	1.19E+07	1.17E+07	7.52E+06	-1.388	-2.593	NA	NA	-0.893	-3.186	-3.328	-3.403	-2.424	-1.625	-3.085	-1.460	1.49E-02	6.41E-02
160	Q6N7S1I1FP2B	2	0.48	-0.96	-1.04	-1.85	-0.24	-0.04	-2.92	0.016445	6.99E-02	-3.58	0	6.75E+07	2.76E+07	6.78E+07	9.98E+07	6.33E+07	6.37E+07	7.03E+07	4.27E+07	0.373	0.081	0.317	1.148	-0.944	-0.720	-0.690	0.101	0.479	-0.563	-1.042	1.64E-02	6.99E-02	
161	zz1Y_FGC2Ccont00195l	5	1.03	2.36	1.33	0.31	2.35	1.70	2.93	0.016355	6.99E-02	-3.57	0	1.27E+08	7.98E+07	1.78E+08	3.16E+07	6.75E+08	5.34E+08	5.50E+08	1.41E+08	1.246	1.431	1.720	-0.260	2.584	2.584	2.430	1.841	1.034	2.860	1.326	1.64E-02	6.99E-02	
162	P026881MEP	3	2.25	1.28	-0.97	-1.71	-0.22	1.77	-2.91	0.016661	6.99E-02	-3.59	0	2.44E+08	1.01E+08	3.21E+08	2.23E+08	2.32E+08	2.23E+08	1.51E+08	NA	2.150	1.866	2.723	2.411	0.854	1.288	1.060	1.935	2.250	1.284	2.960	1.67E-02	7.04E-02	
163	zz1Y_FGC2Ccont00307l	2	-0.49	-2.21	-1.72	-3.02	-0.41	-1.35	-3.09	0.016801	7.05E-02	-3.35	2	1.92E+07	1.62E+07	7.95E+07	NA	2.33E+07	2.02E+07	3.64E+07	NA	-1.369	-0.656	0.552	NA	-2.434	-2.504	-1.690	NA	-0.492	-2.209	-1.718	1.68E-02	7.05E-02	
164	P08123C01A2	15	6.79	5.74	-1.00	-1.85	-0.23	6.26	-2.90	0.016793	7.06E-02	-3.61	0	7.01E+09	2.63E+09	7.19E+09	6.59E+09	3.83E+09	3.58E+09	4.80E+09	3.59E+09	6.798	6.339	7.076	6.937	5.173	5.542	5.716	6.541	6.786	5.743	-1.043	1.70E-02	7.06E-02	
165	P46821MAMP1B	14	3.78	3.00	-0.79	-1.40	-0.18	3.39	-2.90	0.017028	7.06E-02	-3.61	0	8.91E+08	4.36E+08	6.24E+08	6.37E+08	7.07E+08	7.42E+08	9.24E+08	3.19E+08	3.944	3.870	3.533	3.787	2.652	3.096	3.217	3.022	3.784	2.997	0.787	1.70E-02	7.06E-02	
166	Q94760D0AH1	10	4.26	5.15	0.89	1.20	1.59	4.70	2.88	0.017516	7.22E-02	-3.64	0	1.13E+09	3.98E+08	1.47E+09	8.94E+08	3.63E+09	3.45E+09	3.12E+09	1.21E+09	4.270	3.744	4.771	4.245	5.094	5.484	5.063	4.960	4.258	5.500	0.892	1.75E-02	7.22E-02	
167	P005581PGK1	14	3.24	4.41	1.17	0.25	2.09	3.82	2.86	0.018202	7.37E-02	-3.68	0	5.27E+08	2.39E+08	1.05E+09	2.30E+08	2.23E+09	1.59E+09	2.59E+09	7.17E+08	3.216	3.042	2.280	2.416	4.366	4.279	4.776	4.200	3.238	4.405	1.167	1.82E-02	7.37E-02	
168	Q96J9E3MAP6	7	2.19	1.17	-1.02	-1.83	-0.22	1.68	-2.96	0.018108	7.37E-02	-3.68	0	1.95E+08	1.09E+08	2.44E+08	3.02E+08	2.52E+08	1.46E+08	2.40E+08	1.39E+08	1.842	1.969	2.175	2.782	1.118	0.570	1.176	1.805	2.192	1.167	-1.025	1.81E-02	7.37E-02	
169	Q9Y213IGUAD	4	-0.24	0.69	0.94	0.20	1.67	0.23	2.86	0.018069	7.37E-02	-3.67	0	4.26E+07	1.95E+07	7.12E+07	2.87E+07	1.90E+08	1.29E+08	2.25E+08	6.16E+07	-0.264	-0.701	0.391	-0.391	0.691	0.376	1.077	0.632	-0.241	0.694	0.935	1.81E-02	7.37E-02	
170	Q75061A0J1	6	0.46	-0.45	-0.90	-1.62	-0.19	0.01	-2.84	0.018921	7.61E-02	-3.72	0	6.84E+07	2.24E+07	8.60E+07	7.96E+07	8.29E+07	8.16E+07	8.39E+07	2.86E+07	0.392	-0.209	0.665	0.965	-0.542	-0.336	-0.423	-0.483	0.458	-0.446	-0.904	1.89E-02	7.61E-02	
171	P099721ALDCC	16	4.91	5.72	0.81	0.16	1.46	5.31	2.79	0.020635	8.12E-02	-3.80	0	2.07E+09	7.18E+08	2.07E+09	1.25E+09	4.49E+09	4.29E+09	4.99E+09	2.27E+09	5.109	4.957	5.269	4.701	5.410	5.823	5.763	5.873	4.909	5.717	0.808	2.05E-02	8.12E-02	
172	P529071CAZ41	7	-1.55	1.29	2.84	0.95	5.13	-0.13	2.79	0.020497	8.12E-02	-3.80	0</																						

Table with columns: ProteinName, # Pepti des, Ctrl, Group, log2FC, Cl.L, Cl.R, Ave Expr, P.Value, adj.P.Val, B, M3.Co ntr.ro, M2.Co ntr.ro, M4.Co ntr.ro, M1.Co ntr.ro, E4.2.Gr oup.Lr aw, E4.4.Gr oup.Lr aw, E4.3.Gr oup.Lr aw, E4.1.Gr oup.Lr aw, M3.C ontr.transf, M2.C ontr.transf, M4.C ontr.transf, M1.C ontr.transf, E4.2. Group.Ltr ansf, E4.4. Group.Ltr ansf, E4.3. Group.Ltr ansf, E4.1.G roup.Ltr ansf, pseud.o.Ctr l, pseud.o.Grou p, pseud.o.log2 FC, pseudo.P. Value, pseudo.adj .P.Val

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrAs	M3_Co ntrol.rat w	M2_Co ntrol.rat w	M4_Co ntrol.rat w	M1_Co ntrol.rat w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3_C ontrl.transf	M.2_C ontrl.transf	M.4_C ontrl.transf	M.1_C ontrl.transf	E4.2_Group _1tran sf	E4.4_Group _1tran sf	E4.3_Group _1tran sf	E4.1_Group _1tran sf	pseud o.Ctrl	pseud o.Group pt	pseud o.log2 FC	pseud o.P.Value	pseud o.adj .P.Val
237	P21291VATB2	9	0.14	1.14	1.01	-0.03	2.05	0.64	2.18	0.056336	1.62E-01	-4.79	6.61E+07	1.75E+07	1.42E+08	2.38E+07	2.59E+08	1.68E+08	2.74E+08	9.47E+07	0.343	-0.547	1.391	-0.642	1.156	0.790	1.375	1.257	0.136	1.144	1.008	5.63E-02	1.62E-01	
238	Q9LWQ0LIMC1	2	-2.02	-3.43	-1.41	-2.88	0.06	-2.37	-2.43	0.056588	1.62E-01	-4.14	4.10E+07	5.07E+06	1.78E+07	NA	1.20E+07	NA	NA	NA	-2.189	-2.248	-1.617	NA	-3.430	NA	NA	NA	-2.018	-3.430	-1.412	5.66E-02	1.62E-01	
239	P07205PGK2	6	-2.42	-1.67	0.75	-0.03	1.53	-2.05	2.25	0.057436	1.64E-01	-4.59	2.759E+06	4.74E+06	1.13E+07	NA	3.92E+07	3.75E+07	3.36E+07	NA	-2.651	-2.341	-2.272	NA	-1.660	-1.546	-1.890	NA	-2.422	-1.672	0.750	5.74E-02	1.64E-01	
240	P18659IPGAM1	10	3.43	4.44	1.01	-0.04	2.06	3.94	2.16	0.058077	1.65E-01	-4.81	4.432E+08	3.04E+08	1.49E+09	2.35E+08	2.35E+09	1.90E+09	2.05E+09	7.89E+08	3.123	3.376	4.789	2.441	4.441	4.558	4.423	4.338	3.432	4.440	1.008	5.87E-02	1.65E-01	
241	Q13409DC12	2	-0.93	-1.94	-1.01	-2.07	0.05	-1.60	-2.23	0.059213	1.68E-01	-4.57	2.19E+07	1.96E+07	NA	2.21E+07	3.32E+07	2.66E+07	2.1E+07	1.69E+08	NA	-1.113	NA	-0.743	-1.909	-2.078	-2.511	-1.250	-0.928	-1.937	-1.009	5.92E-02	1.68E-01	
242	P601741P15	15	6.88	7.93	1.05	-0.05	2.16	7.40	2.14	0.059779	1.69E-01	-4.84	9.69E+09	2.95E+09	1.14E+10	3.34E+09	2.76E+10	2.67E+10	2.20E+10	4.61E+09	7.246	6.495	7.739	6.020	8.119	8.666	8.028	6.904	6.875	7.929	1.054	5.98E-02	1.69E-01	
243	P04561PFD	2	-1.04	-2.12	-1.07	-2.21	0.06	-1.59	-2.29	0.060280	1.69E-01	-4.48	3.249E+07	NA	NA	1.72E+07	4.05E+07	2.68E+07	1.90E+07	NA	-1.106	NA	-1.078	-1.609	-2.066	-2.670	NA	-1.042	-2.116	-1.073	6.03E-02	1.69E-01		
244	Q9Y4L1HYUJ1	5	-1.50	-0.41	1.09	-0.07	2.24	-0.88	2.16	0.061253	1.72E-01	-4.76	1.184E+07	8.90E+06	1.80E+07	NA	1.27E+08	1.14E+08	1.00E+08	1.19E+07	-1.427	-1.475	-1.595	NA	0.094	0.177	-0.155	-1.761	-0.411	1.069	6.16E-02	1.72E-01		
245	zz1Y_FGC2Ccm00487	15	0.94	0.87	0.92	-0.06	1.91	1.40	2.20	0.062053	1.73E-01	-4.67	2.10E+08	3.46E+07	1.54E+08	NA	4.51E+08	2.67E+08	4.45E+08	NA	0.924	0.390	1.513	NA	1.982	1.509	2.108	NA	0.942	1.866	0.324	6.21E-02	1.73E-01	
246	Q14847LASF1	6	0.74	1.62	0.89	-0.07	1.85	1.98	2.09	0.063025	1.80E-01	-4.92	8.70E+07	5.25E+07	1.43E+08	3.44E+07	2.65E+08	2.56E+08	2.10E+08	0.724	0.962	1.403	-0.146	1.182	1.444	1.394	2.470	0.736	1.625	0.869	6.50E-02	1.80E-01		
247	Q606371SFPY2	2	0.53	-1.51	-2.04	-4.27	0.20	-0.29	-2.21	0.067531	1.86E-01	-4.59	3.947E+07	2.30E+07	1.02E+08	NA	1.15E+07	NA	1.1E+08	NA	0.641	-0.168	0.906	NA	-3.029	NA	0.008	NA	0.526	-1.511	-2.037	6.75E-02	1.86E-01	
248	Q165591DPY2	21	5.61	6.73	1.12	-0.10	2.33	6.17	2.07	0.067781	1.86E-01	-4.96	3.79E+09	1.70E+09	9.15E+09	8.54E+08	1.30E+10	9.21E+09	1.22E+10	2.14E+09	5.948	5.739	6.888	4.183	6.990	7.009	7.129	5.790	5.614	6.729	1.116	6.78E-02	1.86E-01	
249	zz1Y_FGC2Ccm000991	9	4.88	5.49	0.61	-0.05	1.27	5.18	2.07	0.067718	1.86E-01	-4.96	0.168E+09	8.17E+08	2.33E+09	1.10E+09	4.44E+09	3.90E+09	4.12E+09	1.64E+09	4.822	4.733	5.440	4.525	5.393	5.676	5.483	5.403	4.880	5.489	6.009	6.77E-02	1.86E-01	
250	zz1Y_FGC2Ccm00192	6	5.91	5.26	-0.65	-1.36	0.06	5.59	-2.06	0.068354	1.87E-01	-4.97	0.359E+09	2.40E+09	3.96E+09	2.04E+09	3.27E+09	2.86E+09	3.52E+09	1.98E+09	5.874	6.214	6.207	5.359	4.937	5.193	5.244	5.672	5.914	5.262	-0.652	6.84E-02	1.87E-01	
251	Q608691EDF1	2	-1.16	-2.00	-0.84	-1.77	0.08	-1.58	-2.05	0.069632	1.89E-01	-4.99	0.153E+07	7.00E+06	2.62E+07	3.62E+07	2.65E+07	3.41E+07	2.95E+07	1.98E+07	-1.685	-1.806	-1.054	-0.076	-2.245	-1.691	-2.008	-2.042	-1.155	-1.997	-0.842	6.96E-02	1.89E-01	
252	P32004L1CAM	3	-1.04	-1.69	-0.65	-1.36	0.06	-1.36	-2.05	0.069961	1.89E-01	-4.99	0.203E+07	9.61E+06	3.30E+07	2.16E+07	4.12E+07	2.43E+07	3.88E+07	1.58E+07	-1.289	-1.370	-0.720	-0.774	-1.585	-2.221	-1.598	-1.345	-1.039	-1.685	-0.649	7.00E-02	1.89E-01	
253	Q5LXK91NENF1	2	-1.65	-2.32	-0.67	-1.42	0.07	-1.98	-2.05	0.070023	1.89E-01	-4.99	0.112E+07	9.35E+06	1.41E+07	1.67E+07	2.20E+07	2.05E+07	2.77E+07	9.86E+06	-2.110	-1.407	-1.947	-1.118	-2.664	-2.483	-2.099	-2.030	-1.645	-2.319	-0.674	7.00E-02	1.89E-01	
254	Q081741PCDH1	7	0.25	-0.40	-0.66	-1.38	0.07	-0.07	-2.03	0.072096	1.94E-01	-5.02	5.28E+07	2.94E+07	7.24E+07	5.16E+07	9.92E+07	5.38E+07	7.68E+07	4.59E+07	0.034	0.166	0.416	0.402	-0.275	-0.983	-0.554	0.205	0.254	-0.402	-0.656	7.21E-02	1.94E-01	
255	Q180161SRA1	3	-1.78	-2.56	-0.78	-1.66	0.10	-2.30	-2.09	0.073207	1.95E-01	-4.77	2.134E+07	NA	1.69E+07	NA	1.64E+07	2.34E+07	2.01E+07	7.54E+06	-1.864	NA	-1.888	NA	-2.955	-2.277	-2.590	-2.419	-1.776	-2.561	-0.784	7.32E-02	1.95E-01	
256	Q150191DNUB4	2	0.52	-0.35	-0.87	-1.84	0.10	0.02	-2.05	0.073181	1.95E-01	-4.93	NA	6.47E+07	7.75E+07	3.31E+07	9.31E+07	9.23E+07	1.1E+08	2.23E+07	NA	1.251	0.515	-0.199	-0.144	-0.001	-0.845	-0.348	-0.870	7.32E-02	1.95E-01			
257	Q18V501LYSM2	2	-2.56	-3.88	-1.32	-2.83	0.19	-3.22	-2.21	0.074858	1.97E-01	-4.54	5.16E+06	NA	9.15E+06	NA	NA	8.74E+06	2.71E+06	-3.186	NA	NA	-1.932	NA	NA	-3.852	-3.907	-2.559	-3.880	-1.321	7.49E-02	1.97E-01		
258	Q18V501TFPP3	8	2.48	1.83	-0.65	-1.38	0.08	2.16	-2.01	0.074631	1.97E-01	-5.05	0.256E+08	1.20E+08	3.29E+08	3.04E+08	3.36E+08	3.86E+08	1.79E+08	2.216	2.094	2.605	3.016	1.993	1.866	1.895	2.184	2.483	1.834	-0.648	7.49E-02	1.97E-01		
259	Q18V501GF1R1	2	-2.68	-3.70	-1.02	-2.17	0.13	-3.29	-2.14	0.074033	1.97E-01	-4.68	3.87E+06	3.10E+06	NA	NA	6.96E+06	1.26E+07	1.02E+07	NA	-2.437	-2.922	NA	NA	-4.232	-3.244	-3.618	NA	-2.279	-3.698	-1.019	7.40E-02	1.97E-01	
260	zz1Y_FGC2Ccm001161	3	2.20	1.56	-0.64	-1.37	0.08	1.88	-2.01	0.074384	1.97E-01	-5.05	0.273E+08	8.84E+07	2.34E+08	2.85E+08	3.00E+08	3.01E+08	2.53E+08	1.48E+08	2.307	1.679	2.114	2.706	1.376	1.691	1.251	1.911	-2.201	-1.557	-0.644	7.40E-02	1.97E-01	
261	Q195391AGM1	2	-1.68	-3.01	-1.32	-2.84	0.19	-2.67	-2.21	0.0752	1.97E-01	-4.42	4.14E+06	NA	1.70E+07	NA	1.16E+07	1.69E+07	1.79E+07	NA	NA	NA	-1.683	NA	-3.472	-2.784	-2.760	NA	-1.683	-3.005	-1.323	7.52E-02	1.97E-01	
262	Q151811PYR	8	2.72	3.53	0.82	-0.11	1.74	3.18	2.03	0.075978	1.98E-01	-4.96	1.425E+08	1.43E+08	4.01E+08	NA	1.70E+09	1.21E+09	1.21E+09	2.55E+08	2.920	2.339	2.895	NA	3.960	3.857	3.627	2.695	2.718	3.535	0.817	7.60E-02	1.98E-01	
273	P477361PFP6P1	3	-3.23	-4.36	-1.13	-2.46	0.20	-3.79	-1.92	0.086466	2.16E-01	-5.19	0.777E+06	2.12E+06	8.80E+06	1.68E+06	9.58E+06	2.72E+06	4.44E+06	4.48E+06	-2.619	-3.448	-2.634	-4.213	-3.759	-5.623	-4.877	-3.177	-3.228	-4.359	-1.131	8.65E-02	2.16E-01	
274	Q07021C10BP	4	-0.72	0.38	1.10	-0.19	2.38	-0.17	1.92	0.08656	2.16E-01	-5.19	0.579E+07	2.30E+07	4.09E+07	6.27E+06	1.57E+08	1.84E+08	1.56E+08	3.14E+07	0.160	-0.173	-0.410	-2.441	0.411	0.931	0.523	-0.345	-0.716	0.380	1.096	8.66E-02	2.16E-01	
275	Q128601CNTN1	10	0.94	1.87	0.93	-0.17	2.03	1.41	1.90	0.086674	2.20E-01	-5.21	0.533E+07	5.68E+07	2.08E+08	6.49E+07	4.63E+08	5.33E+08	4.09E+08	7.43E+07	0.046	1.071	1.943	0.709	2.023	2.580	1.960	0.904	0.942	1.872	0.930	8.87E-02	2.20E-01	
276	REV_Q8W2421TITN	3	0.55	-0.97	-1.53	-3.36	0.30	-0.32	-1.91	0.090869	2.21E-01	-5.13	1.799E+07	3.39E+07	8.77E+07	NA	2.01E+08	7.52E+07	5.39E+07	4.69E+06	0.605	0.364	0.694	NA	0.776	-0.464	-1.094	-3.111	0.554	-0.973	-1.527	9.09E-02	2.21E-01	
277	P6199114336	10	2.63	3.27	0.64	-0.12	1.41	2.95	1.80																									

1	ProteinName	# Pepti des	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M_3_Co ntrol.ra w	M_2_Co ntrol.ra w	M_4_Co ntrol.ra w	M_1_Co ntrol.ra w	E4.2_Gr oup_Lr aw	E4.4_Gr oup_Lr aw	E4.3_Gr oup_Lr aw	E4.1_Gr oup_Lr aw	M_3_C ontr. transf	M_2_C ontr. transf	M_4_C ontr. transf	M_1_C ontr. transf	E4.2_Gr oup_Lr aw	E4.4_Gr oup_Lr aw	E4.3_Gr oup_Lr aw	E4.1_Gr oup_Lr aw	pseud o.Ctrl	pseud o.Grou pI	pseud o.log2 FC	pseudo.P. Value	pseudo.ad j.P.Val	
331	Q9Y5P6MIFPFB	2	-3.04	-1.83	1.22	-0.50	2.93	-2.07	1.72	0.134594	2.79E-01	-5.07	3	NA	2.84E+06	1.80E+09	1.80E+09	1.06E+09	4.07E+09	3.27E+09	3.44E+09	9.88E+08	4.324	2.727	5.069	4.473	5.262	5.400	5.210	4.855	4.296	5.134	0.836	1.37E-01	2.79E-01
332	P30041FPDX6	16	4.30	5.13	0.84	-0.32	1.99	4.72	1.63	0.136617	2.80E-01	-5.61	0	1.81E+09	1.90E+08	1.80E+09	1.06E+09	4.07E+09	3.27E+09	3.44E+09	9.88E+08	4.324	2.727	5.069	4.473	5.262	5.400	5.210	4.855	4.296	5.134	0.836	1.37E-01	2.80E-01	
333	Q9H299SH3L3	3	2.42	2.93	0.51	-0.19	1.21	2.67	1.63	0.13691	2.80E-01	-5.61	0	3.87E+08	1.02E+08	2.96E+08	2.54E+08	8.37E+08	8.66E+08	6.26E+08	2.80E+08	2.790	1.874	2.453	2.550	2.906	3.335	2.627	2.834	2.417	2.925	0.508	1.37E-01	2.80E-01	
334	Q9NVG81TBC13	3	-3.05	-2.56	0.49	-0.19	1.17	-2.81	1.62	0.137846	2.81E-01	-5.61	0	8.33E+06	2.39E+06	7.33E+06	2.84E+06	2.30E+07	2.06E+07	1.83E+07	6.72E+06	-2.523	-3.282	-2.899	-3.507	-2.453	-2.477	-2.728	-2.587	-3.053	-2.861	0.492	1.38E-01	2.81E-01	
335	P273481H33T	9	3.35	3.87	0.51	-0.20	1.23	3.61	1.61	0.140264	2.86E-01	-5.63	0	4.37E+08	3.37E+08	1.51E+07	9.73E+07	7.81E+07	1.13E+08	8.76E+07	5.95E+07	5.41E+07	1.356	-0.745	0.843	0.958	-0.077	-0.226	-0.943	0.443	0.603	-0.201	-0.804	1.41E-01	2.87E-01
336	P647291FD23A	8	0.60	-0.20	-0.80	-1.93	0.32	0.20	-1.61	0.141202	2.87E-01	-5.63	0	1.37E+08	4.23E+06	6.91E+06	NA	1.61E+07	1.01E+07	1.05E+07	NA	1.356	-0.745	0.843	0.958	-0.077	-0.226	-0.943	0.443	0.603	-0.201	-0.804	1.41E-01	2.87E-01	
337	Q9NVT91AFM1C1	2	-2.78	-3.38	-0.59	-1.45	0.26	-3.08	-1.63	0.145864	2.95E-01	-5.48	2	6.47E+06	NA	NA	NA	NA	NA	NA	NA	-2.872	-2.496	-2.984	NA	-2.964	-3.580	NA	-2.784	-3.378	-0.555	1.46E-01	2.95E-01		
338	P233811SYWC	2	-3.32	-2.12	1.20	-0.59	2.98	-2.42	1.70	0.146876	2.95E-01	-5.08	4	NA	NA	5.48E+06	NA	1.73E+07	NA	3.79E+07	NA	NA	-3.320	NA	NA	-2.875	NA	-1.631	-1.859	-3.320	-2.122	1.158	1.47E-01	2.95E-01	
339	P633041H33Z	11	6.17	6.65	0.48	-0.21	1.17	6.41	1.58	0.147446	2.95E-01	-5.67	0	4.00E+09	1.83E+09	5.43E+09	3.04E+09	1.03E+10	9.19E+09	8.40E+09	3.24E+09	6.274	5.839	6.664	5.896	6.646	7.006	6.565	6.390	6.163	6.632	4.483	1.47E-01	2.95E-01	
340	Q9ULL1SIFPFI2	2	2.09	1.21	-0.88	-2.14	0.38	1.59	-1.50	0.147446	2.95E-01	-5.68	2	2.84E+08	1.57E+08	1.47E+08	NA	1.13E+08	2.53E+08	2.73E+08	1.72E+08	2.362	2.468	1.438	NA	-0.062	1.422	1.368	2.128	2.090	1.209	-0.891	1.49E-01	2.95E-01	
341	P307532AAA	12	-0.66	0.92	1.98	-0.71	3.88	0.13	1.96	0.152957	3.06E-01	-5.70	0	1.35E+07	7.90E+07	9.88E+07	3.78E+06	4.46E+08	6.79E+07	1.88E+08	1.14E+08	-1.855	1.452	0.865	-3.123	1.966	-0.622	0.802	1.531	-0.665	0.320	1.585	1.52E-01	3.06E-01	
342	P225761CALB2	10	5.68	5.23	-0.45	-1.10	0.20	5.46	-1.95	0.154065	3.07E-01	-5.71	0	3.19E+09	1.69E+09	2.72E+09	2.50E+09	2.72E+09	3.20E+09	3.80E+09	1.80E+09	5.708	5.730	5.661	5.630	4.660	5.368	5.364	5.540	5.682	5.233	-0.449	1.54E-01	3.07E-01	
343	P094971CLCB	5	-2.48	-1.20	1.29	-0.62	3.19	-1.62	1.99	0.154985	3.08E-01	-5.48	2	3.12E+06	NA	2.57E+07	NA	3.50E+07	3.52E+07	5.84E+07	3.15E+07	-3.881	NA	-1.083	NA	-1.829	-1.642	-0.969	-0.344	-2.482	-1.196	1.286	1.55E-01	3.08E-01	
344	P421671LAP2B	2	-2.35	-3.46	-1.10	-2.74	0.53	-3.09	-1.99	0.155953	3.08E-01	-5.48	2	NA	1.90E+06	NA	1.68E+07	8.29E+06	1.56E+07	1.10E+07	3.71E+06	NA	-3.592	NA	-1.114	-3.975	-2.908	-3.499	-3.449	-3.458	-1.105	1.56E-01	3.08E-01		
345	P259331GBR2	3	-2.88	-1.97	0.90	-0.43	2.24	-2.42	1.98	0.155756	3.08E-01	-5.54	2	8.38E+06	NA	NA	NA	3.03E+07	2.90E+07	NA	NA	-2.514	NA	-1.998	NA	-4.114	-2.012	-1.875	-2.034	NA	-2.875	-1.974	0.902	1.56E-01	3.08E-01
346	P119891TOP2A	2	1.13	-1.06	-2.20	-5.51	1.11	-1.18	-1.61	0.156738	3.09E-01	-5.40	3	NA	6.52E+07	NA	8.10E+07	8.15E+06	1.21E+08	NA	NA	1.260	NA	1.009	-4.000	0.278	NA	0.532	1.135	-1.033	-2.198	-1.57E-01	3.09E-01		
347	REV_Q519591CCD10	2	-0.21	-0.95	-0.64	-1.98	0.30	-0.57	-1.55	0.159379	3.10E-01	-5.64	NA	1.52E+07	6.23E+07	3.60E+07	6.61E+07	9.52E+07	4.76E+07	1.83E+07	NA	-0.743	0.198	-0.086	-0.890	-0.096	-1.280	-1.130	-0.210	-0.847	-0.636	1.58E-01	3.10E-01		
348	P004921HRT	6	1.65	2.59	0.94	-0.44	2.31	2.12	1.54	0.167591	3.10E-01	-5.73	0	1.16E+08	1.11E+08	4.39E+08	3.81E+07	1.00E+09	4.85E+08	7.69E+08	1.40E+08	1.614	1.992	3.021	-0.008	3.176	2.433	2.938	1.823	1.655	2.593	0.938	1.58E-01	3.10E-01	
349	Q9B6WD11THIC	2	-2.53	-1.65	0.88	-0.48	2.24	-1.87	1.64	0.16842	3.10E-01	-5.16	4	NA	NA	9.47E+06	NA	3.97E+07	3.87E+07	3.37E+07	NA	NA	NA	-2.527	-1.640	-1.495	-1.805	NA	-2.527	-1.647	0.881	1.58E-01	3.10E-01		
350	Q9NRRF61UBQL4	4	-2.86	-3.44	-0.58	-1.44	0.29	-3.15	-1.57	0.169379	3.10E-01	-5.56	2	6.21E+06	4.47E+06	6.98E+06	NA	1.09E+07	1.27E+07	NA	3.59E+06	-2.929	-2.422	-3.222	NA	-3.561	-3.232	NA	-3.514	-2.959	-3.436	-0.578	1.59E-01	3.11E-01	
351	P409261MDH1	6	2.84	3.88	1.04	-0.50	2.58	3.36	1.52	0.162006	3.12E-01	-5.75	0	6.15E+08	2.96E+08	7.31E+08	7.05E+07	1.71E+09	1.93E+09	1.85E+09	2.59E+08	3.431	3.339	3.762	0.821	3.968	4.578	4.268	2.898	2.838	3.878	1.040	1.61E-01	3.12E-01	
352	Q157411T2D1	4	0.56	-0.40	-0.96	-2.39	0.47	0.01	-1.54	0.167077	3.12E-01	-5.65	1	1.64E+08	NA	9.47E+07	2.25E+07	1.35E+08	3.89E+07	9.90E+07	3.66E+07	1.598	NA	0.804	-0.718	0.185	-1.487	-0.170	-0.124	0.561	-0.399	-0.960	1.61E-01	3.12E-01	
353	z1Y-FGC2Ccont003741	7	0.00	-0.56	-0.56	-1.39	0.27	-2.28	-1.52	0.16347	3.12E-01	-5.75	0	4.58E+07	3.05E+07	4.07E+07	4.97E+07	6.35E+07	7.27E+07	5.51E+07	4.78E+07	-0.164	0.217	-0.417	0.350	-0.940	-0.516	-1.060	0.257	-0.004	-0.565	-0.561	1.61E-01	3.12E-01	
354	P525651GDIF1	4	0.45	-1.36	-0.92	-2.31	0.48	-1.06	-1.54	0.164961	3.18E-01	-5.53	2	NA	NA	5.29E+07	2.04E+07	2.87E+07	3.00E+07	5.28E+07	3.20E+07	NA	NA	-0.039	-0.853	-2.123	-1.891	-1.124	-0.317	-0.446	-1.164	-0.917	1.65E-01	3.18E-01	
355	P059371CALB1	13	6.36	5.80	-0.56	-1.40	0.28	6.08	1.90	0.166434	3.19E-01	-5.78	0	5.85E+09	2.81E+09	4.00E+09	2.34E+09	6.12E+09	4.73E+09	6.30E+09	1.44E+09	6.548	6.431	6.901	5.543	5.872	5.972	6.128	5.209	6.359	5.795	-0.560	1.66E-01	3.19E-01	
356	P193241DESP	3	-3.83	-2.71	1.12	-0.65	2.90	-3.55	1.60	0.167791	3.19E-01	-5.20	4	NA	2.68E+06	2.56E+06	2.07E+06	NA	NA	NA	6.19E+06	NA	-3.123	-4.422	-3.937	NA	NA	NA	-2.705	-3.827	-2.795	1.122	1.68E-01	3.19E-01	
357	P194041NDUV2	2	-0.49	-0.05	0.44	-0.22	1.10	-0.27	1.49	0.168204	3.19E-01	-5.79	0	4.79E+07	1.77E+07	2.72E+07	2.99E+07	1.20E+08	1.09E+08	9.46E+07	3.74E+07	-0.103	-0.531	-1.001	-0.334	0.006	0.110	-0.239	-0.093	-0.492	-0.504	0.438	1.68E-01	3.19E-01	
358	P209161MAG	4	2.25	1.78	-0.47	-1.18	0.24	2.01	-1.50	0.166933	3.19E-01	-5.78	0	2.56E+08	1.31E+08	2.58E+08	2.12E+08	3.95E+08	3.06E+08	2.45E+08	2.08E+08	2.217	2.218	2.253	2.303	1.785	1.718	1.202	2.404	-2.248	-1.777	-0.470	1.67E-01	3.19E-01	
359	Q101951TAGL	11	3.01	3.48	0.47	-0.24	1.81	3.24	1.90	0.168004	3.19E-01	-5.79	0	3.84E+08	2.31E+08	6.65E+08	2.68E+08	1.24E+09	1.13E+09	8.93E+08	4.43E+08	2.780	2.999	3.625	2.621	3.496	3.753	3.166	3.498	3.006	3.478	0.472	1.68E-01	3.19E-01	
360	Q9N3V71SYNPD	3	-2.69	-3.52	-0.83	-2.10	0.44	-3.24	-1.53	0.169105	3.20E-01	-5.55	2	8.33E+06	3.26E+06	NA	NA	6.14E+06	8.52E+06	1.69E+07	5.39E+06	-2.523	-2.856	NA	NA	-4.422	-3.847	-2.883	-2.910	-2.689					

1	ProteinName	# Peptides	Ctrl	Group	log2FC	C.L.L	C.L.R	Ave Expr	t	P. Value	adj.P.Val	B	nrN As	M.3.Co ntrol.ra w	M.2.Co ntrol.ra w	M.4.Co ntrol.ra w	M.1.Co ntrol.ra w	E4.2.Gr oup_Tr aw	E4.4.Gr oup_Tr aw	E4.3.Gr oup_Tr aw	E4.1.Gr oup_Tr aw	M.3_C ontrol.transf	M.2_C ontrol.transf	M.4_C ontrol.transf	M.1_C ontrol.transf	E4.2_ Group _1tran sf	E4.4_ Group _1tran sf	E4.3_ Group _1tran sf	E4.1_G roup_1 tran sf	pseud o.Ctrl	pseud o.Grou p1	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val
378	Q43795SEEA	4	0.78	1.23	0.51	-0.34	1.35	1.03	1.35	0.207699	3.75E-01	-5.97	0	9.65E+07	2.51E+07	1.70E+08	5.96E+07	2.56E+08	2.25E+08	3.08E+08	4.18E+07	0.867	0.004	1.651	0.594	1.140	1.238	1.551	1.212	0.779	1.285	0.506	2.08E-01	3.75E-01
379	P23278HMFPA1	2	-1.42	-3.20	-1.78	-4.94	1.37	-2.75	-1.43	0.208779	3.75E-01	-5.40	4	NA	NA	2.04E+07	NA	NA	4.77E+06	3.93E+07	4.18E+06	NA	NA	-1.415	NA	NA	-4.750	-1.570	-3.277	-1.415	-3.199	-1.784	2.08E-01	3.75E-01
380	Q9NVZ3INECP2	2	-5.17	-3.53	1.64	-1.19	4.47	-3.86	1.40	0.209301	3.75E-01	-5.47	3	NA	NA	1.52E+06	NA	3.02E+07	3.81E+06	1.82E+07	2.16E+06	NA	NA	-5.171	NA	-2.047	-5.099	-2.739	-4.238	-5.171	-3.531	1.640	2.08E-01	3.75E-01
381	Q95777ILSM6	3	-3.03	-2.26	0.77	-0.53	2.07	-2.59	1.35	0.21352	3.78E-01	-5.89	1	1.41E+07	1.40E+08	5.57E+06	NA	3.14E+07	3.22E+07	2.54E+07	4.88E+06	-1.793	-4.012	-3.296	NA	-1.990	-1.779	-2.235	-3.050	-3.034	-2.264	0.770	2.1E-01	3.78E-01
382	P16949ISTMNT	7	3.43	3.09	-0.34	-0.93	0.24	3.26	-1.33	0.215134	3.84E-01	-6.00	0	6.63E+08	2.85E+08	6.32E+08	4.56E+08	8.23E+08	8.21E+08	3.25E+08	3.12E+08	3.547	3.285	3.951	3.337	2.861	3.251	3.219	2.990	3.430	3.085	-0.345	2.15E-01	3.84E-01
383	Q98238HS90B	12	-1.83	-0.51	1.32	-0.94	3.58	-1.08	1.34	0.216278	3.84E-01	-5.91	1	6.66E+07	1.85E+06	1.17E+07	NA	1.65E+08	1.33E+08	2.71E+07	2.28E+07	0.354	-3.623	-2.217	NA	0.485	0.422	-2.194	-0.810	-1.831	-0.510	1.321	2.16E-01	3.84E-01
384	Q98559IDOPA	2	-2.71	-1.50	1.21	-0.92	3.34	-1.74	1.37	0.216455	3.84E-01	-5.50	3	NA	NA	8.36E+06	NA	2.03E+07	5.73E+07	3.08E+07	2.77E+07	NA	NA	-2.708	NA	-2.637	-0.886	-1.940	-0.523	-2.708	-1.438	1.210	2.16E-01	3.84E-01
385	P29762IIRAEP1	2	-3.05	-2.42	0.63	-0.48	1.74	-2.68	1.37	0.217472	3.85E-01	-5.70	3	6.35E+06	NA	5.92E+06	NA	1.64E+07	2.35E+07	2.68E+07	NA	-2.898	NA	-3.203	NA	-2.958	-2.268	-2.044	NA	-3.054	-2.423	0.630	2.17E-01	3.85E-01
386	Q4V31CCDS8	3	0.65	-0.11	-0.75	-2.03	0.53	0.27	-1.32	0.219288	3.86E-01	-6.01	0	1.25E+08	4.17E+07	1.30E+08	2.54E+07	1.45E+08	1.52E+08	1.24E+08	1.41E+07	1.226	0.646	1.268	-0.658	0.288	0.631	0.167	-1.507	0.645	-0.103	-0.751	2.18E-01	3.86E-01
387	P27816IMAP4	17	1.80	2.17	0.37	-0.26	1.00	1.99	1.31	0.220295	3.87E-01	-6.02	0	1.97E+08	9.79E+07	6.06E+08	1.64E+08	6.30E+08	4.07E+08	3.56E+08	2.26E+08	1.856	1.919	1.969	1.961	2.224	2.162	1.772	2.519	1.801	2.169	0.368	2.21E-01	3.87E-01
388	Q9H569FCDH9	2	-3.63	-4.45	-0.62	-1.71	0.47	-4.14	-1.34	0.220962	3.87E-01	-5.84	2	2.93E+06	1.00E+06	6.20E+06	NA	6.95E+06	6.42E+06	4.56E+06	NA	-3.967	-4.473	-3.049	NA	-4.230	-4.288	-4.836	NA	-3.830	-4.451	-0.622	2.21E-01	3.87E-01
389	Q9Y2HDIOLG1P4	3	-1.37	-1.89	-0.52	-1.40	0.37	-1.63	-1.31	0.220448	3.87E-01	-6.02	0	2.93E+07	1.14E+07	2.28E+07	7.22E+06	6.95E+07	2.60E+07	4.49E+07	7.80E+06	-0.629	-1.133	-1.255	-2.251	-1.649	-2.185	-1.369	-2.408	-1.367	-1.895	-0.517	2.20E-01	3.87E-01
390	Q76388ISL1	5	2.38	3.17	0.79	-0.59	2.17	2.77	1.29	0.220701	3.95E-01	-6.05	0	5.54E+08	1.37E+08	5.49E+08	5.93E+08	9.69E+08	1.20E+09	9.16E+08	2.22E+08	3.285	2.285	3.347	0.588	3.123	3.844	3.204	2.495	2.376	3.186	0.790	2.29E-01	3.95E-01
391	P0C221CC175	2	1.97	1.18	-0.80	-2.20	0.60	1.83	-1.30	0.227652	3.95E-01	-5.95	1	2.82E+08	4.68E+07	1.63E+08	3.96E+08	1.54E+08	3.96E+08	NA	9.65E+07	2.252	0.799	1.994	3.155	3.377	1.876	NA	1.285	1.976	1.179	-0.796	2.29E-01	3.95E-01
392	P60933GN16F	4	-2.18	-1.30	0.88	-0.69	2.45	-1.74	1.32	0.227298	3.95E-01	-5.87	2	1.15E+07	NA	2.97E+07	2.70E+08	NA	3.73E+07	5.52E+07	1.65E+07	-2.080	NA	-0.875	-3.575	NA	-1.552	-1.066	-1.280	-2.177	-1.296	0.881	2.27E-01	3.95E-01
393	Q05832CALD1	6	0.32	0.77	0.45	-0.33	1.23	0.54	1.29	0.227256	3.95E-01	-6.04	0	6.53E+07	2.98E+07	5.05E+07	7.59E+07	1.71E+08	1.46E+08	1.59E+08	1.06E+08	0.326	0.129	-0.105	0.920	0.539	0.965	0.544	1.416	0.318	0.766	0.448	2.27E-01	3.95E-01
394	Q14982IDPCN4	4	0.08	-0.37	-0.45	-1.22	0.33	-0.15	-1.29	0.227163	3.95E-01	-6.04	0	6.10E+07	2.32E+07	6.55E+07	2.97E+07	1.22E+08	8.15E+07	1.09E+08	1.82E+07	0.232	0.165	0.270	-0.345	0.032	-0.338	-0.027	-1.141	0.078	-0.369	-0.447	2.27E-01	3.95E-01
395	Q8NCV9SINRPE	4	-0.21	0.51	0.71	-0.56	1.98	0.27	1.31	0.228761	3.95E-01	-5.82	2	3.05E+07	NA	6.76E+07	NA	2.19E+08	1.47E+08	2.08E+08	3.00E+07	-0.727	NA	0.916	NA	0.903	0.581	0.954	-0.415	-0.206	0.806	0.711	2.29E-01	3.95E-01
396	Q9F0LQVAPA	4	1.11	-0.74	-1.95	-5.11	1.40	-0.18	-1.28	0.230696	3.97E-01	-6.05	0	1.24E+08	5.02E+07	3.29E+08	3.11E+07	2.05E+08	1.30E+08	3.05E+08	7.89E+05	1.217	0.901	2.609	-0.284	0.805	0.389	1.536	-5.700	1.111	-0.743	-1.954	2.31E-01	3.97E-01
397	P09493ITFM1	5	-5.03	-3.66	1.96	-1.19	3.92	-4.01	1.35	0.232226	3.97E-01	-5.50	4	NA	NA	9.19E+05	NA	1.63E+07	3.94E+06	1.03E+07	4.74E+06	NA	NA	NA	-5.028	NA	-2.839	-5.028	-3.095	-5.028	-3.665	1.963	2.32E-01	3.97E-01
398	P24666PPAC	2	2.78	2.46	-0.32	-0.88	0.24	2.62	-1.28	0.231813	3.97E-01	-6.06	0	3.60E+08	2.15E+08	3.78E+08	2.95E+08	5.78E+08	4.88E+08	5.52E+08	2.44E+08	2.688	2.898	2.801	2.750	2.349	2.444	2.436	2.630	2.784	2.465	-0.320	2.32E-01	3.97E-01
399	Q15491PLEC	15	0.96	0.28	-0.68	-1.91	0.55	0.51	-1.30	0.232468	3.97E-01	-5.83	2	NA	2.94E+07	NA	1.41E+08	1.48E+08	1.04E+08	1.14E+07	NA	0.165	NA	1.758	0.325	0.043	0.702	0.056	0.962	0.281	-0.680	2.32E-01	3.97E-01	
400	Q14651IPLS1	3	-4.32	-3.21	1.11	-0.98	3.19	-3.49	1.34	0.234079	3.99E-01	-5.51	4	NA	1.12E+06	NA	NA	7.96E+06	NA	1.22E+07	8.41E+06	NA	-4.321	NA	-4.036	NA	-3.345	-2.261	-4.321	-3.214	1.107	2.34E-01	3.99E-01	
401	Q9BVI9ICA050	2	-3.01	-3.53	-0.52	-1.47	0.44	-3.32	-1.31	0.235463	4.00E-01	-5.77	3	5.76E+06	NA	6.89E+06	NA	1.29E+07	9.61E+06	1.03E+07	NA	-3.034	NA	-2.989	NA	-3.315	-3.661	-3.604	NA	-3.011	-3.526	-0.515	2.35E-01	4.00E-01
402	Q15691MARE1	2	-2.86	-2.04	0.82	-0.73	2.37	-2.24	1.34	0.236081	4.00E-01	-5.52	4	NA	NA	7.54E+06	NA	3.36E+07	3.41E+07	2.08E+07	NA	NA	NA	-2.857	NA	-1.889	-1.691	-2.537	NA	-2.857	-2.039	0.818	2.36E-01	4.00E-01
403	Q15433RGN	2	-3.34	-2.00	0.34	-0.27	0.95	-2.17	1.24	0.243995	4.13E-01	-6.10	0	1.08E+07	3.80E+06	1.25E+07	6.37E+06	2.95E+07	2.59E+07	2.86E+07	1.20E+07	-2.163	-2.643	-2.125	-2.419	-2.082	-2.125	-2.054	-1.745	-2.338	-2.001	0.336	2.44E-01	4.13E-01
404	Q99502NFTXR	5	0.54	0.11	-0.44	-1.23	0.36	0.32	-1.23	0.248141	4.21E-01	-6.11	0	9.10E+07	5.16E+07	9.56E+07	2.90E+07	1.18E+08	1.16E+08	1.07E+08	4.87E+07	0.787	0.938	0.818	-0.377	-0.020	0.209	-0.056	0.292	0.542	0.106	-0.435	2.48E-01	4.19E-01
405	P07900HS90A	12	5.62	3.27	0.65	-0.54	1.83	2.94	1.23	0.250013	4.21E-01	-6.12	0	1.33E+08	2.52E+08	7.33E+08	2.08E+08	7.85E+08	6.57E+08	1.19E+09	5.22E+08	1.310	3.117	3.765	2.280	2.808	2.966	3.607	3.738	2.618	3.265	0.647	2.50E-01	4.21E-01
406	Q15819UBV2	6	0.92	1.47	0.95	-0.46	1.56	1.19	1.22	0.252441	4.24E-01	-6.13	0	1.02E+08	6.39E+07	1.83E+08	2.93E+07	3.05E+08	3.12E+08	2.78E+08	9.92E+07	0.940	1.344	1.758	-0.362	1.400	1.746	1.395	1.325	0.920	1.466	0.547	2.52E-01	4.24E-01
407	Q194856NFASC	11	1.50	1.87	0.37	-0.32	1.06	1.68	1.22	0.253695	4.24E-01	-6.13	0	1.41E+08	5.43E+07	2.29E+08	1.17E+08	3.75E+08	3.98E+08	1.56E+08	1.96E+08	1.942	1.009	2.084	1.502	1.709	1.878	1.901	1.982	1.497	1.668	0.371	2	

1	ProteinName	# Peptides	Ctrl	Group	log2FC	C.L.	C.I.R.	Ave Expr	t	P. Value	adj.P.Val	B	nrN As	M3_Co ntrol.ra w	M2_Co ntrol.ra w	M4_Co ntrol.ra w	M1_Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M3_C ontrol.tran sf	M2_C ontrol.tran sf	M4_C ontrol.tran sf	M1_C ontrol.tran sf	E4.2_Gr oup_Tr an sf	E4.4_Gr oup_Tr an sf	E4.3_Gr oup_Tr an sf	E4.1_Gr oup_Tr an sf	pseud o.Ctrl	pseud o.Grou p.t	pseud o.log2 FC	pseud o.P. Value	pseud o.adj P.Val
425	P216991NVD	2	-2.73	-3.94	-1.21	-3.84	1.41	-3.64	-1.17	0.293582	4.65E-01	-5.70	4	NA	NA	8.24E+06	NA	1.68E+07	3.24E+06	1.06E+07	NA	NA	NA	NA	NA	NA	NA	NA	-2.919	-5.347	-3.562	1.214	2.92E-01	4.65E-01
426	P280661PSA5	5	-0.17	0.93	1.10	-1.11	3.31	0.38	1.12	0.294449	4.65E-01	-6.24	0	9.06E+07	4.4E+07	1.24E+08	3.35E+06	3.73E+08	2.76E+08	2.41E+08	2.45E+07	0.781	0.635	1.199	-3.286	1.700	1.598	1.180	-0.708	-0.168	0.932	1.100	2.91E-01	4.65E-01
427	Q327521TENR	2	-1.03	-1.40	-0.36	-1.10	0.37	-1.19	-1.13	0.290654	4.65E-01	-6.16	1	2.00E+07	1.46E+07	3.32E+07	1.44E+07	5.00E+07	3.62E+07	4.70E+07	NA	-1.309	-0.796	-0.711	-1.322	-1.296	-1.600	-1.299	NA	-1.035	-1.398	-0.364	2.91E-01	4.65E-01
428	Q329051CSN5	2	-4.47	-5.03	-0.56	-1.74	0.62	-4.81	-1.15	0.292678	4.68E-01	-5.95	3	NA	1.06E+06	2.36E+06	NA	2.75E+06	5.80E+06	4.04E+06	NA	NA	-4.401	-4.538	NA	-5.622	-4.444	-5.023	NA	-4.469	-5.030	-0.560	2.93E-01	4.68E-01
429	P235981IF4B	2	-2.59	-3.49	-0.90	-2.86	1.06	-3.26	-1.16	0.294386	4.68E-01	-5.71	4	7.95E+06	NA	NA	NA	2.1E+07	7.27E+06	9.06E+06	NA	-2.588	NA	NA	NA	-2.579	-4.094	-3.796	NA	-2.588	-3.490	-0.902	2.94E-01	4.68E-01
430	P072371FOA1	5	-3.95	-2.57	0.99	-1.16	3.13	-2.81	1.16	0.295203	4.68E-01	-5.71	4	NA	NA	4.67E+06	NA	NA	1.27E+07	4.12E+07	5.13E+06	NA	NA	NA	-3.551	NA	-3.228	-1.498	-2.978	-3.551	-2.568	0.963	2.95E-01	4.68E-01
431	P321191PFDX2	5	1.74	2.41	0.67	-0.70	2.04	2.07	1.11	0.296908	4.69E-01	-6.25	0	2.72E+08	1.25E+08	3.52E+08	3.29E+07	5.06E+08	4.69E+08	4.34E+08	3.22E+08	2.301	2.152	2.706	-0.207	2.154	2.362	2.070	3.036	1.738	2.410	0.673	2.97E-01	4.69E-01
432	Q166301CPSF6	2	4.08	5.96	3.12	-3.84	3.60	1.58	-1.12	0.301933	4.74E-01	-5.78	3	9.85E+08	NA	NA	NA	6.61E+08	5.46E+08	8.08E+08	2.01E+06	4.082	NA	NA	NA	2.552	2.619	3.015	-4.344	4.082	0.361	-3.121	3.02E-01	4.74E-01
433	Q3693191WBF2	2	-0.53	0.06	0.59	-0.63	1.81	-0.24	1.09	0.302502	4.74E-01	-6.27	0	2.40E+07	1.69E+07	4.03E+07	3.71E+07	1.98E+08	1.36E+08	3.80E+07	6.16E+07	-1.060	-0.596	-0.430	-0.045	0.758	0.459	-1.621	0.632	-0.533	0.057	0.590	3.03E-01	4.74E-01
434	Q96191EPCD2	7	3.05	2.63	-0.43	-1.31	0.46	2.84	-1.09	0.302228	4.74E-01	-6.27	0	4.16E+08	1.45E+08	5.50E+08	5.61E+08	5.80E+08	4.23E+08	5.83E+08	4.15E+08	2.890	2.358	3.351	3.617	2.357	2.222	2.519	3.403	3.054	2.625	-0.429	3.02E-01	4.74E-01
435	zz1Y-FGC2Ccont004611	8	2.53	1.97	-0.56	-1.72	0.59	2.25	-1.09	0.301283	4.74E-01	-6.27	0	4.05E+08	2.59E+08	4.95E+08	7.85E+07	4.28E+08	4.90E+08	3.56E+08	1.33E+08	2.554	3.160	3.199	0.931	1.905	2.449	1.771	1.756	2.533	1.971	-0.563	3.01E-01	4.74E-01
436	Q160271PUB4	3	-2.89	-2.32	0.57	-0.65	1.78	-2.51	1.09	0.310178	4.85E-01	-6.07	2	3.99E+06	NA	1.28E+07	NA	3.29E+07	1.99E+07	2.39E+07	7.81E+06	-3.693	NA	-2.089	NA	-1.325	-2.535	-2.468	-2.369	-2.890	-2.324	0.565	3.10E-01	4.85E-01
437	Q795941PAK3	13	-1.37	-0.90	0.47	-0.53	1.47	-1.14	1.07	0.310105	4.85E-01	-6.30	0	8.99E+06	8.21E+06	2.89E+07	2.52E+07	8.03E+07	1.99E+07	2.39E+07	7.81E+06	-2.430	-1.896	-0.914	-0.564	-0.590	-0.535	-1.410	-1.095	-1.373	-0.900	0.473	3.10E-01	4.85E-01
438	Q096681AHNK	110	3.47	4.89	1.42	-1.88	4.43	4.18	1.07	0.312404	4.85E-01	-6.29	0	8.14E+07	2.05E+09	5.06E+09	6.34E+07	8.23E+09	2.44E+09	2.59E+09	1.38E+09	0.532	5.998	6.562	0.677	4.716	4.944	4.795	5.151	3.467	4.892	1.425	3.12E-01	4.85E-01
439	Q139071D11	3	-3.19	-2.63	0.56	-0.68	1.80	-2.86	1.10	0.313171	4.85E-01	-6.01	3	4.55E+06	NA	5.71E+06	NA	2.59E+07	1.16E+07	NA	8.50E+06	-3.359	NA	-3.025	NA	-2.278	-3.371	NA	-2.246	-3.182	-2.632	0.560	3.13E-01	4.85E-01
440	Q861281ISCA2	3	-0.60	-1.28	-0.48	-1.51	0.54	-1.04	-1.06	0.31373	4.85E-01	-6.30	0	3.51E+07	1.98E+07	4.06E+07	9.71E+06	5.99E+07	4.80E+07	6.87E+07	8.69E+06	-0.531	-0.379	-0.423	-1.850	-1.027	-1.159	-0.723	-2.213	-0.728	-1.261	-0.485	3.14E-01	4.85E-01
441	Q46071IREPS1	2	-3.09	-2.59	0.49	-0.95	1.53	-2.84	1.07	0.312981	4.85E-01	-6.30	0	4.86E+06	4.61E+06	5.84E+06	2.92E+06	2.74E+07	1.89E+07	3.26E+06	1.15E+07	-3.268	-2.378	-3.227	-3.471	-2.196	-2.613	-3.763	-1.805	-3.086	-2.894	0.492	3.13E-01	4.85E-01
442	Q022461CNTN2	3	-3.89	-4.47	-0.58	-1.84	0.68	-4.22	-1.05	0.322617	4.97E-01	-6.23	1	2.95E+06	1.02E+06	5.67E+06	NA	7.70E+06	2.45E+06	3.73E+06	2.02E+06	-3.960	-4.452	-3.269	NA	-4.085	-5.785	-3.689	-4.332	-3.894	-4.473	-0.579	2.23E-01	4.97E-01
443	Q8NF231CADM4	5	-1.15	-1.77	-0.62	-1.99	0.74	-1.41	-1.05	0.326038	5.00E-01	-6.24	1	1.42E+07	1.08E+07	4.29E+07	1.51E+07	7.53E+07	3.70E+07	1.49E+07	NA	-1.785	-1.203	-0.341	-1.256	-0.687	-1.565	-3.056	NA	-1.146	-1.769	-0.623	3.25E-01	5.00E-01
444	Q3LUN21NSFC	3	-1.27	-0.91	0.36	-0.42	1.13	-1.09	1.04	0.325984	5.00E-01	-6.33	0	2.46E+07	1.07E+07	1.41E+07	2.01E+07	5.64E+07	5.40E+07	5.07E+07	3.09E+07	-1.025	-1.223	-1.953	-0.871	-1.117	-0.976	-1.184	-0.369	-1.258	-0.911	0.357	3.25E-01	5.00E-01
445	Q608941DNJA2	2	-1.33	-1.29	-0.06	-2.77	1.06	-1.62	-1.05	0.328929	5.03E-01	-6.11	2	1.92E+07	2.32E+07	3.02E+07	4.29E+06	NA	NA	4.32E+07	5.24E+06	-1.369	-0.159	-0.849	-2.953	NA	NA	-1.426	-2.949	-1.333	-2.188	-0.955	3.29E-01	5.03E-01
446	Q795981PFMTB	3	-3.93	-4.74	-0.81	-2.59	0.98	-4.39	-1.04	0.328397	5.03E-01	-6.25	1	2.72E+06	1.83E+06	3.27E+06	NA	1.32E+07	6.37E+06	6.11E+06	3.30E+05	-4.071	-3.647	-4.066	NA	-3.286	-4.298	-4.394	-6.965	-3.928	-4.736	-0.808	3.29E-01	5.03E-01
447	P139291ENDB	6	2.85	3.91	1.06	-1.27	3.40	3.38	1.02	0.331964	5.06E-01	-6.34	0	7.43E+08	3.71E+08	1.36E+09	2.43E+07	1.99E+09	1.81E+09	1.95E+09	2.21E+08	3.632	3.647	4.658	-0.613	4.191	4.480	4.346	2.616	2.846	3.908	1.062	3.32E-01	5.06E-01
448	zz1Y-FGC2Ccont001821	9	6.07	5.74	-0.33	-1.04	0.39	5.91	-1.02	0.333681	5.08E-01	-6.34	0	4.34E+09	1.95E+09	5.36E+09	3.47E+09	4.47E+09	4.02E+09	5.64E+09	2.29E+09	6.135	5.930	6.645	5.564	5.403	5.722	5.960	5.888	6.069	5.743	-0.325	3.34E-01	5.08E-01
449	P503951GDI6	7	-0.48	-1.45	-0.97	-3.16	1.22	-1.03	-1.02	0.339174	5.12E-01	-6.27	1	1.46E+07	1.06E+07	1.58E+08	NA	2.54E+07	2.82E+07	1.63E+08	9.43E+06	-1.749	-1.234	1.542	NA	-2.305	-1.985	0.584	-2.095	-0.480	-1.450	-0.970	3.38E-01	5.12E-01
450	P632411IF5A1	4	1.73	2.04	0.30	-0.37	0.98	1.89	1.01	0.338232	5.12E-01	-6.35	0	2.33E+08	6.45E+07	1.98E+08	1.39E+08	5.01E+08	4.44E+08	4.28E+08	1.26E+08	2.087	1.246	1.869	1.736	2.138	2.297	2.049	1.671	1.753	2.039	0.304	3.38E-01	5.12E-01
451	O003301ODPX	2	-2.45	-2.10	0.35	-0.46	1.15	-2.22	1.01	0.343776	5.18E-01	-6.15	2	NA	4.80E+06	9.19E+06	NA	2.75E+07	2.65E+07	2.74E+07	9.98E+06	NA	-2.322	-2.572	NA	-2.188	-2.081	-2.120	-2.012	-2.447	-2.100	0.347	3.44E-01	5.18E-01
452	O005841RNT2	2	-0.28	-1.66	-1.38	-4.65	1.89	-1.11	-1.02	0.345726	5.18E-01	-6.08	3	NA	2.60E+07	NA	2.53E+07	8.60E+07	5.48E+06	1.14E+08	NA	0.000	NA	-0.559	-0.487	-4.532	0.047	NA	-0.279	-1.657	-1.378	3.46E-01	5.18E-01	
453	O43491E4L2	2	-2.99	-4.19	-1.20	-4.14	1.73	-3.89	-1.04	0.344163	5.18E-01	-5.84	4	NA	2.96E+06	NA	NA	4.05E+06	4.01E+06	2.12E+07	NA	NA	-2.987	NA	NA	-5.043	-5.019	-2.511	NA	-2.987	-4.191	-1.204	3.44E-01	5.18E-01
454	P610861UBE2K	5	-0.84	0.05	0.89	-1.15	2.93	-0.33	1.00	0.346844	5.18E-01	-6.29	1	1.34E+07	NA	1.32E+08	9.15E+06	1.71E+08	2.06E+08	1.07E+08	1.55E+07	-1.868	NA	1.290	-1.932	0.536	1.105	-0.058	-1.375	-0.837	0.052	0.888	3.47E-01	5.18E-01
455	P614571PHS	5	2.26	2.57	0.31	-0.39	1.00	2.41	0.99	0.346354	5.18E-01	-6.37	0	3.22E+08	9.05E+07	3.41E+08	1.88E+08	7.15E+08	6.10E+08	5.49E+08	2.05E+08	2.533	1.711	2.660	2.144	2.670	2.791	2.426	2.262	2.262	2.567	0.305	3.46E-01	5.18E-01
456	Q997291RDOAA	2	-2.45	-1.91	0.54	-0.68	1.75	-2.18	0.99	0.345556	5.18E-01	-6.37	0	6.11E+06	7.25E+06	1.99E+07	2.60E+06	4.74E+07	2.96E+07	3.95E+07	5.82E+06	-2.952	-1.756	-1.455	-3.628	-1.377	-1.909	-1.564	-2.797	-2.448	-1.172	0.536	3.46E-01	5.18E-01
457	zz1Y-FGC2Ccont003691	6	0.68	1.10	0.42	-0.53	1.36	0.89	0.99	0.348579	5.20E-01	-6.38	0	3.96E+07	2.84E+07	1.29E+08	6.89E+07	2.95E+08	3.11E+08	2.46E+08	4.24E+07	0.829	0.119	1.256	0.531	1.351	1.746	1.211						

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M.3_Co ntrol.ra w	M.2_Co ntrol.ra w	M.4_Co ntrol.ra w	M.1_Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3_C ontr.transf	M.2_C ontr.transf	M.4_C ontr.transf	M.1_C ontr.transf	E4.2_Gr oup_Tr ansf	E4.4_Gr oup_Tr ansf	E4.3_Gr oup_Tr ansf	E4.1_Gr oup_Tr ansf	pseud o.Ctrl	pseud o.Group Pl	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val
472	O4562UBFD1	3	-0.31	-1.07	-0.76	-2.67	1.15	-0.75	-0.92	0.385778	5.57E-01	-6.37	1	7.79E+07	NA	3.19E+07	2.23E+07	1.22E+07	1.16E+08	7.94E+07	2.65E+07	0.571	NA	-0.771	-0.723	-3.403	0.206	-0.591	-0.310	-1.073	-0.763	3.68E-01	5.57E-01	
473	P09336LCHL1	11	6.36	5.32	-0.44	-1.54	0.66	6.14	-0.91	0.3683	5.59E-01	-6.45	0	4.60E+09	3.27E+09	2.99E+09	5.67E+09	3.41E+09	5.38E+09	3.80E+09	5.43E+09	6.217	6.637	5.802	6.780	4.999	6.173	5.360	7.142	6.359	5.978	-0.440	3.68E-01	5.59E-01
474	Q95881TXD12	2	-0.64	-1.06	-0.42	-1.46	0.63	-0.85	-0.90	0.390713	5.62E-01	-6.46	0	3.34E+07	2.10E+07	4.67E+07	1.32E+07	6.80E+07	8.05E+07	8.55E+06	-0.600	-0.299	-0.220	-1.441	-0.896	-0.618	-0.476	-2.236	-0.640	-1.057	-0.417	3.91E-01	5.62E-01	
475	D753471BCA	3	2.13	1.83	-0.30	-1.07	0.46	1.99	-0.89	0.393513	5.65E-01	-6.46	0	2.22E+08	8.82E+07	2.80E+08	2.37E+08	3.07E+08	4.65E+08	2.90E+08	1.66E+08	2.018	1.675	2.373	2.454	1.408	2.367	1.460	2.070	2.130	1.826	-0.304	3.94E-01	5.65E-01
476	P304191NMT1	2	-0.66	-1.00	-0.34	-1.21	0.53	-0.83	-0.89	0.397593	5.68E-01	-6.47	0	2.09E+07	1.06E+07	4.37E+07	7.73E+07	4.75E+07	5.37E+07	1.68E+07	-1.247	-1.233	-0.314	0.141	-0.647	-1.176	-0.938	-1.256	-0.663	-1.005	-0.341	3.98E-01	5.68E-01	
477	Q9HK1ISCU	3	-1.72	-1.25	0.47	-0.72	1.66	-1.49	0.89	0.397205	5.68E-01	-6.47	0	8.76E+06	1.34E+07	1.43E+07	1.19E+07	6.52E+07	7.27E+07	1.89E+07	2.38E+07	-2.454	-0.917	-1.929	-1.572	-1.044	-0.515	-2.680	-0.752	-1.776	-1.248	0.470	3.97E-01	5.68E-01
478	zz1Y-FGC2Ccont003611	7	1.18	0.76	-0.43	-1.52	0.66	0.97	-0.88	0.399993	5.69E-01	-6.47	0	1.32E+08	5.14E+07	1.63E+08	7.55E+07	2.91E+08	1.84E+08	2.89E+08	2.49E+07	1.302	0.933	1.956	0.914	1.328	1.457	0.929	1.457	1.884	0.765	-0.427	3.99E-01	5.69E-01
479	P135291P1M1	3	-1.64	-2.08	-0.44	-1.80	0.72	-1.86	-0.89	0.401734	5.72E-01	-6.32	2	2.57E+07	4.39E+06	1.91E+07	NA	4.17E+07	2.53E+07	2.10E+07	NA	-0.964	-2.446	-1.619	NA	-1.568	-2.158	NA	-1.641	-2.082	-0.441	4.02E-01	5.72E-01	
480	P207741M1M1	2	-3.20	-3.87	0.32	-0.51	1.15	-3.03	0.88	0.402394	5.72E-01	-6.48	0	5.21E+06	2.68E+06	6.36E+06	3.15E+06	1.55E+07	1.20E+07	1.29E+07	1.10E+07	-3.173	-3.136	-3.105	-3.368	-3.040	-3.318	-3.260	-1.876	-2.874	0.322	4.02E-01	5.72E-01	
481	O43601L1TQF5	2	-2.46	-1.95	0.91	-1.51	3.34	-1.85	0.88	0.408254	5.74E-01	-6.27	2	3.04E+06	NA	2.71E+07	NA	4.09E+07	6.95E+07	7.33E+07	1.93E+06	-3.917	NA	-1.007	NA	-1.599	-0.608	-0.627	-3.365	-2.462	-1.650	0.912	4.05E-01	5.74E-01
482	zz1Y-FGC2Ccont003261	7	1.58	1.86	0.28	-0.44	1.00	1.72	0.87	0.407523	5.75E-01	-6.49	0	1.77E+08	1.03E+08	2.05E+08	6.99E+07	4.04E+08	3.39E+08	3.79E+08	1.45E+08	1.705	1.985	1.922	0.809	1.817	1.880	1.683	1.876	1.580	1.959	0.279	4.08E-01	5.75E-01
483	Q8N1C9INFCP1	3	-1.04	-1.38	-0.34	-1.25	0.56	-1.24	-0.87	0.41136	5.80E-01	-6.41	1	1.94E+07	NA	2.24E+07	2.65E+07	4.52E+07	3.77E+07	3.30E+07	2.42E+07	-1.360	NA	-1.282	-0.499	-1.446	-1.534	-0.722	-1.044	-1.385	-0.341	4.11E-01	5.80E-01	
484	P136311CTP1	5	3.16	3.64	0.48	-0.78	1.73	3.40	0.95	0.416513	5.85E-01	-6.50	0	6.75E+08	3.26E+08	8.84E+08	1.25E+08	1.69E+09	1.35E+09	1.43E+09	2.55E+08	3.859	3.470	4.037	1.590	3.956	4.021	3.880	2.639	3.164	3.639	0.475	4.16E-01	5.85E-01
485	Q86Y82STX12	8	2.76	2.48	-0.29	-1.06	0.48	2.62	-0.84	0.418769	5.90E-01	-6.51	0	5.88E+08	1.19E+08	4.52E+08	2.52E+08	5.46E+08	5.80E+08	5.51E+08	2.21E+08	3.369	2.084	3.066	2.540	2.269	2.712	2.434	2.491	2.765	2.477	-0.268	4.20E-01	5.90E-01
486	Q85782AF2A1	2	-4.04	-3.53	0.51	-0.96	1.97	-3.66	0.87	0.420909	5.90E-01	-5.99	4	2.79E+06	NA	NA	NA	9.52E+06	9.40E+06	1.40E+07	NA	-4.038	NA	NA	NA	-3.768	-3.695	-3.134	NA	-4.038	-3.532	0.506	4.21E-01	5.90E-01
487	P369571ODD2	5	1.23	1.53	0.30	-0.52	1.12	1.40	0.84	0.426646	5.97E-01	-6.44	1	NA	6.80E+07	1.59E+08	7.12E+07	3.10E+08	2.20E+08	2.78E+08	1.68E+08	NA	1.319	1.546	0.834	1.425	1.209	1.398	2.092	1.233	1.931	0.298	4.27E-01	5.97E-01
488	Q8N791V1T41	2	-1.62	-2.16	-0.54	-2.07	0.98	-1.99	-0.84	0.428968	5.99E-01	-6.31	2	NA	6.77E+06	2.09E+07	NA	2.55E+07	1.24E+07	6.24E+07	8.61E+06	NA	-1.950	-1.930	NA	-2.299	-3.265	-0.869	-2.227	-1.620	-2.165	-0.545	4.29E-01	5.99E-01
489	P609531CDD42	4	0.03	-0.38	-0.41	-1.95	0.74	-0.20	-0.83	0.431784	6.02E-01	-6.44	1	6.76E+07	1.39E+07	8.16E+07	NA	8.02E+07	1.23E+08	1.07E+08	1.77E+07	0.375	-0.861	0.589	NA	-0.591	0.302	-0.053	-1.180	0.034	-0.380	-0.415	4.32E-01	6.02E-01
490	Q136551PCP1	6	0.18	0.52	0.34	-0.60	1.28	0.37	0.82	0.432874	6.02E-01	-6.45	1	5.68E+07	NA	5.45E+07	5.17E+07	1.92E+08	2.08E+08	1.68E+08	3.06E+07	0.135	NA	0.004	0.402	0.708	1.118	0.635	-0.384	0.180	0.519	0.339	4.33E-01	6.02E-01
491	Q19UBBS1NCDN	4	-1.33	-1.00	0.33	-0.60	1.27	-1.14	0.82	0.433397	6.02E-01	-6.45	1	1.03E+07	1.52E+07	2.67E+07	NA	6.01E+07	5.10E+07	6.18E+07	1.99E+07	-2.226	-0.737	-1.030	NA	-1.023	-1.066	-0.884	-1.011	-1.331	-0.996	0.335	4.34E-01	6.02E-01
492	Q432951SRGP3	2	-3.37	-3.93	-0.57	-2.22	1.09	-3.49	-0.83	0.438325	6.06E-01	-6.07	3	5.03E+06	4.18E+06	6.59E+06	2.28E+06	NA	NA	NA	2.66E+06	-3.221	-2.511	-3.932	-3.807	NA	NA	NA	-3.934	-3.368	-3.934	-0.567	4.38E-01	6.06E-01
493	Q5T0131HY1	4	-1.14	-0.95	0.59	-1.08	2.27	-0.80	0.81	0.4395	6.06E-01	-6.45	1	2.26E+07	1.09E+07	2.57E+07	NA	NA	1.47E+08	7.89E+07	6.52E+06	-1.139	-1.202	-1.094	NA	0.375	0.579	-0.515	-2.631	-1.142	-0.548	0.594	4.39E-01	6.06E-01
494	Q161431SYUB	6	6.65	6.33	-0.32	-1.24	0.59	6.49	-0.80	0.444223	6.13E-01	-6.55	0	8.94E+09	1.96E+09	3.59E+09	9.78E+09	6.40E+09	5.98E+09	7.99E+09	3.60E+09	7.134	5.935	6.067	7.469	5.939	6.340	6.482	6.545	6.651	6.326	-0.325	4.44E-01	6.13E-01
495	P484431CFKL	5	0.29	0.66	0.37	-0.69	1.43	0.47	0.79	0.447582	6.16E-01	-6.55	0	5.43E+07	5.28E+07	9.30E+07	2.32E+07	2.13E+08	2.49E+08	1.62E+08	3.46E+07	0.070	0.972	0.778	-0.677	0.864	1.399	0.580	-0.205	0.286	0.600	3.474	-0.48E-01	6.16E-01
496	P509911TCPD	2	-3.08	-2.67	0.41	-0.85	1.67	-2.75	0.79	0.459113	6.31E-01	-6.10	3	NA	2.77E+06	NA	2.14E+07	1.93E+07	1.43E+07	7.51E+06	NA	-3.076	NA	NA	NA	-2.562	-2.575	-3.102	-2.425	-3.076	-2.666	0.410	4.59E-01	6.31E-01
497	Q437651SGTA	4	-0.49	-0.11	0.98	-0.76	1.52	-0.27	0.77	0.465349	6.37E-01	-6.49	1	1.88E+07	2.71E+07	2.10E+07	5.21E+09	1.70E+10	2.11E+10	1.69E+10	6.39E+09	8.030	8.995	8.622	6.620	7.393	8.296	7.630	7.378	8.067	7.674	-0.392	4.65E-01	6.37E-01
498	zz1Y-FGC2Ccont002611	11	8.07	7.67	-0.39	-1.95	0.77	7.87	-0.76	0.464672	6.37E-01	-6.57	0	1.71E+10	1.82E+10	1.21E+10	5.21E+09	1.70E+10	2.11E+10	1.69E+10	6.39E+09	8.030	8.995	8.622	6.620	7.393	8.296	7.630	7.378	8.067	7.674	-0.392	4.65E-01	6.37E-01
499	P620721T1M10	2	-3.15	-3.54	-0.39	-1.59	0.81	-3.35	-0.77	0.467036	6.38E-01	-6.42	2	6.11E+06	2.33E+06	6.01E+06	NA	7.30E+06	7.87E+06	2.15E+07	NA	-2.951	-3.314	-3.185	NA	-4.165	-3.971	-2.488	NA	-3.150	-3.541	-0.391	4.67E-01	6.38E-01
500	Q948111PPP	3	-1.22	-0.93	0.28	-0.96	1.13	-1.08	0.76	0.468557	6.38E-01	-6.58	0	1.65E+07	9.04E+06	2.10E+07	2.73E+07	6.70E+07	4.44E+07	4.61E+07	3.32E+07	-1.577	-1.453	-1.378	-0.460	-0.859	-1.283	-1.328	-0.265	-1.217	-0.934	0.283	4.69E-01	6.38E-01
501	P453811ACY2	2	-2.84	-2.51	0.33	-0.65	1.30	-2.67	0.75	0.469191	6.38E-01	-6.58	0	6.69E+06	6.16E+06	7.17E+06	2.63E+06	2.06E+07	2.15E+07	1.33E+07	1.15E+07	-2.827												

ProteinName	# Pepti des	Ctrl	Group	log2FC	Cl.L	Cl.R	Ave Expr	t	P.Value	adj P.Val	B	rrN As	M.3.Co ntrol.ra w	M.2.Co ntrol.ra w	M.4.Co ntrol.ra w	M.1.Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3.C ontrl.transf	M.2.C ontrl.transf	M.4.C ontrl.transf	M.1.C ontrl.transf	E4.2_ Group_1Tran sf	E4.4_ Group_1Tran sf	E4.3_ Group_1Tran sf	E4.1_Gr oup_1Tran sf	pseud o.Chrl	pseud o.Group	pseud o.log2 FC	pseudo.P. Value	pseudo.ad j.P.Val
P62253LUBG21	2	-2.12	-2.61	-0.49	-2.21	1.23	-2.40	-0.65	0.53375	7.01E-01	-6.58	1.122E+07	5.02E+06	1.127E+07	NA	4.25E+07	2.63E+07	3.07E+07	1.42E+06	-1.993	-2.262	-2.106	NA	-1.539	-2.096	-1.945	-4.951	-2.120	-2.608	-0.487	5.34E-01	7.01E-01	
P52815IPM12	2	0.47	0.26	-0.21	-0.95	0.53	0.37	-0.64	0.536394	7.05E-01	-6.66	0.61E+07	3.28E+07	6.74E+07	7.55E+07	1.46E+08	8.49E+07	1.24E+08	7.16E+07	0.344	0.317	0.313	0.914	0.306	-0.274	0.167	0.851	0.472	0.262	-0.210	5.39E-01	7.05E-01	
z1z1.FG2C2Cant001491	3	5.00	4.79	-0.22	-0.99	0.56	4.89	-0.63	0.545817	7.12E-01	-6.67	0.169E+08	5.99E+08	2.15E+09	2.35E+09	2.36E+09	2.24E+09	2.63E+09	1.31E+09	4.833	4.304	5.325	5.545	4.448	4.809	4.806	5.080	5.002	4.785	-0.216	5.46E-01	7.12E-01	
z1z1.FG2C2Cant004551	3	2.76	2.96	0.20	-0.52	0.93	2.96	0.63	0.545745	7.12E-01	-6.67	0.477E+08	1.66E+08	5.00E+08	1.94E+08	9.41E+08	2.52E+08	8.09E+08	2.43E+08	3.080	2.546	3.212	2.187	3.080	3.115	3.013	2.625	2.756	2.958	0.202	5.46E-01	7.12E-01	
P00491IPM17	2	-1.91	-2.20	-0.28	-1.36	0.80	-2.09	-0.63	0.548467	7.14E-01	-6.41	3.123E+07	6.78E+06	NA	NA	2.25E+07	2.13E+07	3.65E+07	NA	-1.978	-1.849	NA	NA	-2.495	-2.420	-1.684	NA	-1.914	-2.186	-0.282	5.49E-01	7.14E-01	
P003381LDA	3	0.86	1.14	0.27	-0.73	1.27	1.00	0.61	0.55573	7.23E-01	-6.68	0.674E+07	3.79E+07	1.75E+06	7.35E+07	1.71E+08	1.44E+08	2.93E+08	1.56E+06	0.370	0.514	1.632	0.878	0.538	0.548	1.476	1.980	0.864	1.136	0.272	5.56E-01	7.23E-01	
P13791ACPH	3	-3.14	-2.61	0.53	-1.54	2.60	-2.72	0.62	0.557967	7.23E-01	-6.22	3 NA	NA	6.21E+06	NA	1.39E+07	1.87E+07	1.26E+07	1.64E+07	NA	NA	NA	NA	-3.219	-2.625	-3.302	-1.291	-3.139	-2.609	0.529	5.59E-01	7.23E-01	
P56290CAD10	11	1.53	1.73	0.19	-0.52	0.91	1.63	0.61	0.557774	7.23E-01	-6.68	0.100E+08	1.12E+08	1.74E+08	1.19E+08	3.84E+08	2.47E+08	3.91E+08	1.49E+08	0.923	2.007	1.681	1.517	1.742	1.383	1.874	1.904	1.532	1.726	1.194	5.59E-01	7.23E-01	
Q56292VAPB	5	3.30	3.12	-0.18	-0.86	0.50	3.21	-0.60	0.565237	7.27E-01	-6.69	0.490E+08	3.33E+08	6.44E+08	3.61E+08	8.54E+08	6.99E+08	7.49E+08	4.94E+08	3.116	3.502	3.579	3.022	2.936	3.002	2.898	3.659	3.305	3.124	-0.181	5.65E-01	7.27E-01	
P170501NAGAB	3	-1.97	-1.74	0.23	-0.65	1.11	-1.84	0.60	0.56665	7.27E-01	-6.61	1.11E+07	4.18E+06	2.20E+07	NA	3.42E+07	2.99E+07	3.20E+07	1.62E+07	-2.081	-2.512	-1.309	NA	-1.863	-1.903	-1.882	-1.907	-1.967	-1.739	0.229	5.66E-01	7.27E-01	
P22061IPM17	8	3.89	4.10	0.22	-0.61	1.04	4.00	0.60	0.568585	7.27E-01	-6.69	0.124E+09	3.27E+08	1.16E+09	4.24E+08	2.18E+09	1.68E+09	1.51E+09	5.29E+08	4.404	3.476	4.426	3.239	4.329	4.365	3.965	3.758	3.886	4.104	0.218	5.66E-01	7.27E-01	
Q695761T2D3	2	-2.32	-2.66	-0.34	-1.74	1.06	-2.41	-0.61	0.565527	7.27E-01	-6.19	4.958E+06	NA	8.24E+06	8.21E+06	NA	NA	6.38E+06	-2.330	NA	-2.564	-2.077	NA	NA	NA	-2.663	-2.324	-2.663	-0.339	5.65E-01	7.27E-01		
z1z1.FG2C2Cant002951	6	-0.53	-0.21	0.32	-0.91	1.55	-0.35	0.60	0.564141	7.27E-01	-6.61	1.150E+07	2.67E+07	6.77E+07	NA	1.46E+08	6.16E+07	1.26E+08	2.70E+07	-1.712	0.032	0.088	NA	0.305	-0.773	0.198	-0.568	-0.531	-0.209	0.321	5.64E-01	7.27E-01	
P00121GAL3A	2	-1.54	-1.28	0.26	-0.75	1.26	-1.39	0.59	0.573707	7.35E-01	-6.62	1.145E+07	NA	1.73E+07	1.56E+07	1.56E+07	7.62E+07	3.45E+07	6.88E+07	9.56E+06	-1.755	NA	-1.653	-1.215	-0.668	-1.673	-0.723	-2.074	-1.541	-1.204	0.257	5.74E-01	7.35E-01
P527881SPSY	3	-1.86	-1.18	0.69	-2.09	3.46	-1.40	0.58	0.578362	7.38E-01	-6.50	2.717E+06	NA	2.73E+07	NA	4.89E+07	3.00E+07	1.54E+07	1.12E+08	NA	NA	-0.937	NA	-1.328	-1.889	-2.988	1.503	-1.864	-1.176	0.688	5.78E-01	7.38E-01	
Q153691ELOD	4	0.64	0.80	0.16	-0.47	0.79	0.72	0.58	0.577728	7.38E-01	-6.70	0.794E+07	4.72E+07	9.21E-07	5.10E+07	1.99E+08	1.33E+08	2.19E+08	7.81E+07	0.597	0.816	0.765	0.395	0.763	0.428	1.036	0.977	0.641	0.801	0.160	5.78E-01	7.38E-01	
P61891APP3	4	-2.06	-2.54	-0.47	-2.48	1.53	-2.44	-0.57	0.587294	7.47E-01	-6.25	3 NA	NA	1.31E+07	NA	4.36E+07	1.20E+07	2.78E+07	4.30E+06	NA	NA	NA	NA	-1.500	-3.319	-2.095	-3.226	-2.063	-2.537	-0.474	5.87E-01	7.47E-01	
z1z1.FG2C2Cant003561	9	1.16	0.79	-0.36	-1.84	1.12	0.95	-0.56	0.587628	7.47E-01	-6.63	1.409E+07	6.95E+07	3.11E+08	NA	1.82E+08	1.34E+08	3.05E+08	5.87E+07	-0.323	1.267	2.525	NA	0.631	0.439	1.536	0.562	1.156	0.792	0.364	5.88E-01	7.47E-01	
Q155401FAPB7	3	3.14	2.95	-0.19	-0.98	0.59	3.04	-0.56	0.591118	7.50E-01	-6.72	0.426E+08	1.78E+08	4.17E+08	7.72E+08	8.76E+08	7.39E+08	7.35E+08	2.83E+08	2.921	2.644	2.949	4.046	2.973	3.089	2.869	2.849	3.140	2.945	-0.195	5.91E-01	7.50E-01	
P16851LUGPA	5	-1.18	-0.81	0.38	-1.24	1.99	-0.96	0.56	0.592703	7.50E-01	-6.45	3 NA	NA	1.35E+07	1.97E+07	NA	9.72E+07	2.82E+07	1.02E+08	NA	NA	-0.899	-1.470	NA	-0.304	-1.989	-0.129	NA	-1.184	-0.807	0.377	5.92E-01	7.50E-01
Q507841COM1	4	-0.19	-0.38	-0.19	-0.82	0.56	-0.28	-0.95	0.595749	7.53E-01	-6.72	0.16E+07	1.68E+07	4.23E+07	4.39E+07	6.13E+07	9.80E+07	1.01E+08	3.20E+07	0.000	-0.599	-0.361	0.182	-0.992	-0.052	-0.138	-0.320	-0.195	-0.375	-0.181	5.96E-01	7.53E-01	
P07191NFM	5	-0.97	-0.70	0.27	-0.84	1.38	-0.84	0.95	0.596966	7.53E-01	-6.72	0.141E+07	7.94E+06	2.97E+07	5.23E+07	6.07E+07	7.76E+07	5.68E+07	3.08E+07	-1.798	-1.631	-0.875	0.419	-1.006	-0.413	-1.012	-0.373	-0.971	0.271	5.97E-01	7.53E-01		
z1z1.FG2C2Cant003821	5	0.62	0.34	-0.28	-1.45	0.89	0.48	-0.54	0.598604	7.54E-01	-6.72	0.840E+07	5.11E+07	1.35E+08	2.77E+07	1.42E+08	2.42E+08	1.52E+08	2.39E+07	0.675	0.926	1.313	-0.437	0.256	1.351	0.481	-0.745	0.619	0.836	-0.283	5.99E-01	7.54E-01	
P26401VID	2	-2.46	-2.82	-0.36	-1.92	1.19	-2.70	-0.95	0.601109	7.56E-01	-6.52	2 NA	NA	1.01E+07	4.44E+06	NA	1.46E+07	1.29E+07	2.29E+07	6.76E+06	NA	-1.900	-3.622	NA	-3.127	-3.199	-2.391	-2.578	-2.461	-2.824	-0.362	6.01E-01	7.56E-01
Q165951FRDA	2	-1.18	-0.89	0.29	-0.94	1.52	-1.01	0.54	0.604847	7.59E-01	-6.65	1 NA	9.14E+06	4.79E+07	9.24E+06	9.02E+07	6.11E+07	8.05E+07	1.09E+07	NA	NA	-1.438	-0.182	-1.919	-0.416	-0.784	-0.485	-1.880	-1.180	-0.891	0.288	6.05E-01	7.59E-01
P134731LAMP2	2	-1.77	-1.49	0.28	-0.91	1.46	-1.63	0.53	0.603931	7.60E-01	-6.73	0.132E+07	9.22E+06	2.24E+07	4.12E+06	7.11E+07	5.43E+07	3.73E+07	6.76E+06	-1.968	-1.426	-1.279	-3.006	-0.770	-0.969	-1.649	-2.578	-1.770	-1.492	0.278	6.09E-01	7.60E-01	
P300481PFDX3	6	-1.16	-0.97	0.19	-0.61	0.99	-1.06	0.53	0.609466	7.60E-01	-6.73	0.238E+07	6.53E+06	3.00E+07	2.13E+07	7.66E+07	5.39E+07	7.00E+07	1.39E+07	-1.070	-1.901	-0.858	-0.795	-0.659	-0.981	-0.696	-1.533	-1.156	-0.967	0.189	6.09E-01	7.60E-01	
Q9NP171PALM3	4	-2.85	-2.52	0.33	-1.11	1.76	-2.68	0.54	0.608129	7.60E-01	-6.58	2.127E+07	1.96E+06	8.17E+06	NA	2.99E+07	3.21E+07	1.19E+07	NA	-1.935	-3.863	-2.741	NA	-2.070	-2.106	-3.382	NA	-2.847	-2.519	0.328	6.08E-01	7.60E-01	
Q9K9171BT3L4	3	-2.07	-2.49	-0.42	-2.26	1.43	-2.35	-0.53	0.611484	7.62E-01	-6.53	2.968E+06	NA	1.53E+07	NA	3.18E+07	3.21E+07	3.33E+07	1.96E+06	-2.315	NA	-1.831	NA	-1.971	-1.784	-1.824	-4.379	-2.073	-2.489	-0.417	6.11E-01	7.62E-01	
P539991TCP4	3	-2.65	-1.91	0.74	-2.57	4.06	-2.16	0.53	0.619058	7.63E-01	-6.53	2.894E+05	NA	NA	4.81E+07	3.42E+07	2.84E+07	2.08E+07	1.67E+07	-5.613	NA	NA	0.306	-1.865	-1.974	-2.538	-1.267	-2.653	-1.911	0.743	6.15E-01	7.63E-01	
Q9P121NTFI	4	0.76	0.46	-0.30	-1.60	1.00	0.61	-0.52	0.619959	7.63E-01	-6.74	0.721E+07	6.60E+07	6.69E+07	7.96E+07	2.39E+08	2.36E+08	1.91E+08	1.57E+07	0.464	1.277	0.301	0.985	1.0									

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CLL	CL.R	Ave Expr	t	P.Value	adj.P.Val	B	nrAs	M_3_Co ntrol.ra	M_2_Co ntrol.ra	M_4_Co ntrol.ra	M_1_Co ntrol.ra	E4.2_Gr oup_Lr	E4.4_Gr oup_Lr	E4.3_Gr oup_Lr	E4.1_Gr oup_Lr	M_3_C ontrol	M_2_C ontrol	M_4_C ontrol	M_1_C ontrol	E4.2_Gr oup_Lr	E4.4_Gr oup_Lr	E4.3_Gr oup_Lr	E4.1_Gr oup_Lr	pseud o.Ct	pseud o.Grou p	pseud o.log2 FC	pseud o.P. Value	pseudo .adj .P.Val	
566	P54727HIC23B	11	4.33	4.21	-0.12	-0.74	0.50	4.27	-0.45	0.664671	7.97E-01	-6.77	0	1.13E+09	5.00E+08	1.19E+09	1.12E+09	1.86E+09	1.99E+09	1.46E+09	1.93E+09	3.25E+08	4.270	4.060	4.463	4.545	4.146	4.037	4.569	4.335	4.212	-0.123	6.65E-01	7.97E-01	
567	Q96A3T9YFPE	2	-0.68	-1.07	-0.39	-2.42	1.64	-0.34	-0.45	0.664071	7.97E-01	-6.57	2	NA	6.65E+06	7.60E+07	NA	7.20E+07	2.85E+07	1.33E+08	1.14E+07	NA	-1.875	0.523	NA	-0.753	-1.972	0.278	-1.821	-0.676	-1.067	-0.391	6.64E-01	7.97E-01	
568	Q3C005DIPY30	2	-0.28	0.11	0.38	-1.56	2.32	-0.06	0.45	0.663145	7.97E-01	-6.70	1	NA	4.1E+06	7.22E+07	1.00E+08	8.81E+07	8.05E+07	1.65E+08	6.13E+07	NA	-2.538	0.411	1.296	-0.451	-0.357	0.605	0.626	-0.277	0.106	0.383	6.63E-01	7.97E-01	
569	Q39757TTCDCM	3	1.50	0.97	-0.53	-3.19	2.14	1.24	-0.45	0.666049	7.97E-01	-6.78	0	3.02E+08	8.79E+07	4.50E+08	1.61E+07	3.78E+08	5.28E+08	3.81E+08	8.36E+06	2.444	1672	3.061	-1.172	1.721	2.568	1.875	-2.269	1.501	0.974	-0.528	6.66E-01	7.97E-01	
570	O1516L1SM1	2	-2.06	-1.89	0.18	-0.78	1.14	-1.96	0.45	0.670449	8.00E-01	-6.52	3	1.06E+07	NA	1.42E+07	NA	3.27E+07	3.40E+07	2.88E+07	NA	-2.186	NA	-1.944	NA	-1.944	NA	-1.944	NA	-1.928	-2.065	-1.888	0.177	6.70E-01	8.00E-01
571	Q3H446FRWDD1	2	-5.03	-4.78	0.25	-1.16	1.66	-4.91	0.45	0.669524	8.00E-01	-6.43	4	NA	NA	1.31E+06	1.19E+06	3.74E+06	NA	NA	1.92E+06	NA	NA	-5.389	-4.678	-5.161	NA	NA	-4.404	-5.033	-4.783	0.251	6.70E-01	8.00E-01	
572	Q004939IBN1	10	4.19	4.39	0.20	-0.85	1.26	4.29	0.43	0.673688	8.02E-01	-6.78	0	1.70E+09	4.38E+08	1.75E+09	3.59E+08	2.07E+09	1.82E+09	2.66E+09	6.27E+08	4.840	3.978	5.028	4.257	4.492	4.820	4.005	4.190	4.333	0.203	6.74E-01	8.02E-01		
573	P51859HDCG	2	-2.49	-2.65	-0.16	-1.05	0.72	-2.60	-0.43	0.673906	8.06E-01	-6.58	2	NA	3.73E+06	1.10E+07	NA	2.00E+07	1.90E+07	1.53E+07	7.91E+06	NA	-2.670	-2.315	NA	-2.665	-2.598	-3.006	-2.390	-2.432	-2.655	-0.163	6.78E-01	8.06E-01	
574	zz1-FG2CCent001001	17	3.02	2.96	-0.46	-2.89	1.97	2.79	-0.43	0.678949	8.06E-01	-6.78	0	1.65E+08	2.79E+08	6.30E+08	6.03E+08	1.05E+08	3.05E+08	8.68E+08	1.87E+09	1.606	3.260	3.500	3.715	-0.171	1.712	3.105	5.594	3.020	2.560	-0.460	6.79E-01	8.06E-01	
575	Q15700DLG2	6	-0.83	-1.43	-0.59	-3.79	2.61	-1.09	-0.42	0.682162	8.08E-01	-6.71	1	7.97E+06	4.43E+06	6.30E+08	9.58E+06	4.04E+07	6.14E+07	3.19E+07	NA	-2.584	-2.432	3.547	-1.870	-1.612	-0.778	-1.890	NA	-0.835	-3.427	-0.592	6.82E-01	8.08E-01	
576	Q60688CUTA	3	3.82	3.95	-0.27	-1.75	1.21	3.69	-0.41	0.68739	8.14E-01	-6.79	0	1.18E+09	7.17E+08	1.27E+09	1.51E+08	1.56E+09	1.28E+09	1.94E+09	2.12E+08	4.330	4.554	4.567	1.846	3.837	3.946	3.992	2.431	3.123	1.527	1.652	0.432	6.86E-01	8.14E-01
577	zz1-FG2CCent003591	10	1.58	1.42	-0.17	-1.10	0.77	1.49	-0.41	0.691342	8.16E-01	-6.71	1	1.03E+08	7.02E+07	2.77E+08	NA	3.76E+08	2.01E+08	3.35E+08	9.15E+07	1.032	1.363	2.359	NA	1.710	1.067	1.691	1.208	1.594	1.416	-0.168	6.91E-01	8.16E-01	
578	Q60929PPFD1	4	0.40	0.25	-0.15	-0.97	0.67	0.32	-0.41	0.692649	8.16E-01	-6.79	0	9.93E+07	3.69E+07	3.87E+07	6.42E+07	1.63E+08	1.18E+08	1.01E+08	5.40E+07	0.906	0.479	-2.359	0.694	0.462	0.232	-0.143	0.441	0.397	0.249	-0.149	6.93E-01	8.16E-01	
579	P43487FRAMG	2	-1.34	-1.47	-0.13	-0.86	0.62	-1.41	-0.39	0.702825	8.27E-01	-6.72	1	2.04E+07	NA	1.66E+07	1.81E+07	3.75E+07	4.03E+07	3.90E+07	1.81E+07	NA	-1.715	-1.010	-1.724	-1.411	-1.585	-1.144	-1.336	-1.466	-0.130	7.03E-01	8.27E-01		
580	P28070P5B4	2	-3.76	-3.52	0.24	-1.17	1.64	-3.64	0.39	0.705151	8.28E-01	-6.66	2	1.97E+06	1.67E+06	NA	4.22E+06	6.29E+06	2.12E+06	2.15E+07	NA	-4.516	-3.777	NA	-2.976	-3.975	-4.104	-2.483	NA	-3.756	-3.521	0.235	7.05E-01	8.28E-01	
581	Q172653INECA2	2	-2.43	-2.72	-0.29	-2.01	1.43	-2.62	-0.39	0.706282	8.28E-01	-6.60	2	NA	NA	4.48E+06	1.52E+07	1.52E+07	1.25E+07	1.77E+07	1.19E+07	NA	-3.612	-1.245	-3.069	-3.254	-2.780	-1.762	-2.429	-2.716	-0.287	7.06E-01	8.28E-01		
582	P247521THL	3	-3.66	-3.95	-0.29	-2.01	1.44	-3.86	-0.39	0.707533	8.28E-01	-6.60	2	3.98E+06	1.66E+06	NA	6.86E+06	3.23E+06	2.12E+07	3.30E+06	-3.546	-3.783	NA	NA	-4.257	-5.327	-2.507	-3.713	-3.664	-3.951	-0.287	7.08E-01	8.28E-01		
583	Q9HCJ6VATIL	4	-1.38	-1.76	-0.38	-2.66	1.91	-1.63	-0.39	0.709125	8.29E-01	-6.60	2	NA	NA	3.23E+07	6.63E+06	7.84E+06	4.37E+07	7.60E+07	1.87E+07	NA	NA	-0.752	-2.010	-4.059	-1.307	-0.571	-1.100	-1.381	-1.759	-0.378	7.09E-01	8.29E-01	
584	P52306IGDS1	4	-3.30	-2.91	0.39	-2.04	2.82	-3.07	0.39	0.712665	8.30E-01	-6.55	3	1.40E+06	NA	1.79E+07	NA	1.35E+07	NA	2.63E+07	4.08E+06	-4.991	NA	-1.610	NA	-3.247	NA	-2.161	-3.312	-3.301	-2.913	0.387	7.13E-01	8.30E-01	
585	Q15819MFPX1	3	-0.11	-0.39	-0.29	-1.96	1.41	-0.25	-0.38	0.715819	8.30E-01	-6.71	0	5.71E+07	3.00E+07	6.26E+07	1.86E+07	1.96E+08	1.19E+08	1.98E+08	5.45E+06	0.140	0.193	0.205	-0.972	0.741	0.250	0.328	-2.992	-0.109	-0.393	-0.285	7.14E-01	8.30E-01	
586	Q9H0E2ICLIP	2	-0.20	-0.44	-0.23	-1.63	1.16	-0.34	-0.38	0.716359	8.30E-01	-6.73	1	4.43E+07	2.22E+07	4.79E+07	NA	1.21E+08	1.26E+08	1.88E+08	8.81E+06	-0.213	-0.218	-0.183	NA	1.006	0.335	0.099	-2.194	-0.204	-0.366	-0.231	7.14E-01	8.30E-01	
587	Q8N573ICXR1	5	0.69	0.84	0.15	-0.73	1.02	0.76	0.38	0.716826	8.31E-01	-6.81	0	9.79E+07	2.02E+07	1.25E+08	8.17E+07	2.10E+08	2.18E+08	1.93E+08	6.77E+07	0.887	-0.348	1.206	1.020	0.841	1.190	0.846	0.770	0.691	0.837	0.146	7.16E-01	8.31E-01	
588	P09471GNAND	2	-3.07	-3.24	-0.17	-1.26	0.91	-3.17	-0.37	0.721057	8.32E-01	-6.73	1	4.83E+06	NA	7.99E+06	3.67E+06	6.79E+06	1.18E+07	1.51E+07	7.93E+06	-3.276	NA	-2.773	-3.161	-4.273	-3.336	-3.023	-2.476	-3.070	-3.244	-0.174	7.21E-01	8.32E-01	
589	P36558NMJ24	2	-4.56	-4.05	0.51	-2.64	3.67	-4.27	0.37	0.719727	8.32E-01	-6.73	1	1.42E+06	1.12E+06	NA	1.47E+06	3.07E+06	2.76E+06	2.99E+06	4.99E+07	-4.968	-4.323	NA	-4.396	-5.455	-5.599	-5.477	0.326	-4.562	-4.051	0.511	7.20E-01	8.32E-01	
590	P41236IPF2	2	-0.93	-0.42	0.51	-2.64	3.66	-0.53	0.37	0.718494	8.32E-01	-6.73	1	8.84E+07	NA	1.08E+08	1.34E+06	1.34E+06	1.47E+06	1.22E+06	6.94E+06	0.745	NA	1.000	-4.524	0.175	0.581	0.143	-2.562	-0.926	-0.416	0.511	7.19E-01	8.32E-01	
591	P60723IBASP1	8	1.93	2.01	0.14	-0.74	1.02	2.00	0.37	0.721617	8.32E-01	-6.81	0	1.68E+08	1.28E+08	1.29E+08	2.71E+08	3.48E+08	4.15E+08	2.48E+08	2.48E+08	1.635	2.187	1.556	2.191	1.845	2.654	1.927	2.071	0.144	7.22E-01	8.32E-01			
592	zz1-FG2CCent003421	6	-3.36	-3.51	-0.15	-1.16	0.85	-3.43	-0.35	0.734392	8.46E-01	-6.67	2	3.29E+06	2.87E+06	5.82E+06	NA	1.63E+07	7.47E+06	1.10E+07	NA	-3.807	-3.030	-3.233	NA	-2.968	-4.052	-3.501	NA	-3.356	-3.507	-0.151	7.35E-01	8.46E-01	
593	Q9533616PGL	5	-0.02	0.13	0.16	-0.89	1.20	0.07	0.34	0.740722	8.49E-01	-6.74	1	NA	2.1E+07	9.13E+07	2.59E+07	1.48E+08	1.28E+08	1.57E+08	2.49E+07	NA	-0.290	0.751	-0.527	0.320	0.367	0.531	-0.694	-0.022	0.133	0.166	7.41E-01	8.49E-01	
594	P300091KCY	3	-1.34	-1.45	-0.12	-0.89	0.65	-1.40	-0.34	0.740645	8.49E-01	-6.82	0	1.89E+07	8.01E+06	3.08E+07	1.24E+07	4.32E+07	5.41E+07	2.78E+07	1.71E+07	-1.387	-1.620	-0.622	-1.522	-1.516	-0.974	-2.097	-1.231	-1.338	-1.454	-0.117	7.41E-01	8.49E-01	
595	Q9BWS91MUDT9	2	-3.63	-3.37	0.27	-1.62	2.15	-3.45	0.33	0.743691	8.59E-01	-6.62	2	NA	NA	2.33E+06	6.16E+06	1.97E+07	1.82E+07	4.26E+06	4.50E+06	NA	-4.558	-4.207	-5.693	-2.669	-								

ProteinName	# Pepti des	Ctrl	Group	log2FC	Cl.L	Cl.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M.3.Co ntrol.ra w	M.2.Co ntrol.ra w	M.4.Co ntrol.ra w	M.1.Co ntrol.ra w	E4.2.Gr oup_1r aw	E4.4.Gr oup_1r aw	E4.3.Gr oup_1r aw	E4.1.Gr oup_1r aw	M.3.C ontr.transf	M.2.C ontr.transf	M.4.C ontr.transf	M.1.C ontr.transf	E4.2.Gr oup_1tran sf	E4.4.Gr oup_1tran sf	E4.3.Gr oup_1tran sf	E4.1.Gr oup_1tran sf	pseud o.Cht	pseud o.Grou p1	pseud o.Log2 FC	pseudo.P. Value	pseudo.ad j.P.Val	
Q15233NANO	4	-4.87	-5.06	-0.19	-2.13	1.75	-5.02	-0.25	0.813633	9.04E-01	-6.37	4	NA	NA	NA	1.03E+06	5.65E+06	2.79E+06	NA	1.63E+06	NA	NA	NA	-4.873	-4.573	-5.966	NA	-4.649	-4.873	-5.063	-0.190	8.14E-01	9.04E-01	
zz1Y-FGC2Cant00276l	8	-2.96	-2.46	0.50	-4.46	5.46	-2.96	0.24	0.815071	9.04E-01	-6.40	3	NA	NA	NA	4.27E+06	6.31E+06	7.76E+06	1.83E+07	9.57E+07	NA	NA	NA	-2.959	-4.382	-3.992	-2.733	1.273	-2.959	-2.459	0.500	8.15E-01	9.04E-01	
P55036P5M04	3	-1.13	-1.24	-0.11	-1.14	0.92	-1.21	-0.24	0.816639	9.04E-01	-6.65	2	1.60E+07	NA	NA	2.37E+07	4.66E+07	5.42E+07	5.85E+07	1.33E+07	-1.621	NA	NA	NA	-0.649	-1.433	-0.970	-0.968	-1.590	-1.125	-1.240	-0.106	8.17E-01	9.04E-01
Q9LUJIDENL	6	0.26	0.18	-0.08	-0.80	0.65	0.22	-0.24	0.818906	9.05E-01	-6.85	0	7.04E+07	2.41E+07	6.24E+07	5.60E+07	1.20E+08	1.75E+08	1.04E+08	3.88E+07	0.430	-0.108	0.201	0.510	0.013	0.852	-0.089	-0.047	0.258	0.12	-0.076	8.19E-01	9.05E-01	
P235BICMGP	4	-0.21	-0.15	0.07	-0.58	0.71	-0.18	0.23	0.821077	9.06E-01	-6.85	0	4.01E+07	1.92E+07	4.45E+07	4.42E+07	9.89E+07	8.71E+07	9.05E+07	4.66E+07	-0.349	-0.415	-0.288	0.133	-0.279	-0.234	-0.307	0.226	0.275	-0.148	0.066	8.21E-01	9.06E-01	
Q9N3JRICADM2	4	-1.14	-1.00	0.14	-1.24	1.51	-1.07	0.22	0.827225	9.11E-01	-6.86	0	2.57E+07	4.56E+06	2.43E+07	3.75E+07	8.99E+07	2.03E+07	7.77E+07	2.76E+07	-0.962	-2.394	-1.162	-0.029	-0.421	-2.497	-0.550	-0.532	-1.137	-1.000	0.137	8.27E-01	9.11E-01	
Q9LUM2IEPDR1	3	-0.46	-0.55	-0.09	-1.07	0.88	-0.50	-0.23	0.827782	9.11E-01	-6.71	2	3.91E+07	1.92E+07	3.67E+07	NA	5.75E+07	1.1E+08	6.98E+07	NA	-0.382	-0.420	-0.568	NA	-1.088	0.136	-0.699	NA	-0.457	-0.550	-0.094	8.28E-01	9.11E-01	
Q9N3JRICADM2	4	-1.14	-1.00	0.14	-1.24	1.51	-1.07	0.22	0.827782	9.11E-01	-6.86	0	2.57E+07	4.56E+06	2.43E+07	3.75E+07	8.99E+07	2.03E+07	7.77E+07	2.76E+07	-0.962	-2.394	-1.162	-0.029	-0.421	-2.497	-0.550	-0.532	-1.137	-1.000	0.137	8.27E-01	9.11E-01	
P619601UFM1	2	3.29	3.40	0.10	-0.97	1.18	3.34	0.22	0.831636	9.12E-01	-6.86	0	9.23E+08	3.05E+08	7.47E+08	1.70E+09	1.07E+09	1.30E+09	1.17E+09	2.69E+08	3.993	3.378	3.732	2.006	3.270	3.966	3.578	2.773	3.242	3.397	0.305	8.32E-01	9.12E-01	
Q9Y813EFN1	4	1.76	1.57	-0.19	-2.14	1.77	1.66	-0.22	0.831782	9.12E-01	-6.86	0	1.69E+08	3.35E+08	3.53E+08	2.07E+07	4.26E+08	4.01E+08	3.99E+08	4.85E+07	1.644	3.508	2.707	-0.831	1.897	2.140	1.945	0.286	1.757	1.567	-0.190	8.32E-01	9.12E-01	
zz1Y-FGC2Cant00142l	6	1.93	1.66	-0.27	-3.05	2.52	1.80	-0.22	0.834033	9.13E-01	-6.86	0	5.33E+08	8.23E+07	6.61E+08	2.26E+07	7.00E+08	7.11E+08	6.75E+08	1.19E+07	3.232	1.581	3.617	-0.710	2.639	3.028	2.741	-1.753	1.930	1.664	-0.266	8.34E-01	9.13E-01	
Q102952AKA12	7	0.71	0.60	-0.11	-1.23	1.02	0.65	-0.21	0.835869	9.13E-01	-6.86	0	6.79E+07	2.71E+07	7.13E+07	1.69E+08	1.08E+08	1.38E+08	1.70E+08	1.06E+08	0.380	0.056	0.394	1.996	-0.149	0.482	0.652	1.416	0.707	0.600	-0.106	8.36E-01	9.13E-01	
Q151021PA1B3	4	1.99	1.92	-0.07	-0.82	0.68	1.95	-0.21	0.836897	9.13E-01	-6.86	0	3.16E+08	1.05E+08	2.56E+08	9.92E+07	4.31E+08	3.53E+08	3.95E+08	1.72E+08	2.507	1.910	2.242	1.281	1.914	1.940	1.679	2.127	1.995	1.915	-0.070	8.37E-01	9.13E-01	
zz1Y-FGC2Cant00464l	2	0.75	0.66	-0.10	-1.15	0.96	0.70	-0.21	0.840915	9.16E-01	-6.78	1	7.55E+07	2.74E+07	NA	1.32E+08	1.68E+08	1.97E+08	1.20E+08	7.82E+07	0.528	0.067	NA	1.667	0.509	1.031	0.116	0.980	0.754	0.659	-0.095	8.41E-01	9.16E-01	
Q9LUMFOICAM5	4	-0.44	-0.32	0.12	-1.24	1.48	-0.38	0.20	0.845655	9.19E-01	-6.86	0	5.85E+07	1.87E+07	7.52E+07	8.98E+06	1.33E+08	1.13E+08	1.08E+08	1.34E+07	0.174	-0.457	0.470	-1.957	0.158	0.175	-0.032	-1.587	-0.442	-0.321	0.121	8.46E-01	9.19E-01	
zz1Y-FGC2Cant00347l	15	9.76	9.67	-0.09	-1.12	0.93	9.71	-0.20	0.846465	9.19E-01	-6.86	0	9.50E+10	4.39E+10	5.74E+10	1.88E+10	8.60E+10	5.45E+10	6.08E+10	2.90E+10	10.405	10.207	10.076	8.353	9.811	9.774	9.567	9.527	9.760	9.670	-0.091	8.46E-01	9.19E-01	
Q9C0401TRIM2	2	-3.79	-3.67	0.13	-1.40	1.65	-3.74	0.20	0.847953	9.20E-01	-6.61	3	3.69E+06	9.19E+05	6.24E+06	NA	NA	1.44E+07	6.49E+06	NA	-3.655	-4.594	-3.300	NA	-3.031	-4.303	NA	-3.793	-3.667	0.126	8.48E-01	9.20E-01		
P153741UHL3L	7	3.08	3.14	0.06	-0.64	0.76	3.11	0.19	0.850258	9.21E-01	-6.86	0	5.23E+08	2.82E+08	5.87E+08	2.29E+08	1.01E+09	8.51E+08	8.61E+08	3.08E+08	3.207	3.271	3.444	2.406	3.178	3.309	3.111	2.971	3.082	3.142	0.060	8.50E-01	9.21E-01	
Q9LUH69ISWP70	3	-3.01	-3.16	0.14	-1.96	1.67	-3.13	-0.19	0.853616	9.23E-01	-6.41	3	NA	NA	6.77E+06	NA	1.82E+07	1.64E+07	6.54E+06	6.20E+06	NA	NA	-3.013	NA	-2.800	-2.832	-4.292	-2.705	-3.013	-3.757	-0.144	8.54E-01	9.23E-01	
P014791SCDM	3	2.41	2.20	-0.20	-2.71	2.30	2.30	-0.18	0.858792	9.24E-01	-6.87	0	4.31E+08	2.02E+08	4.66E+08	6.74E+07	1.1E+09	1.08E+09	9.73E+08	1.43E+07	2.938	2.813	3.110	0.760	3.324	3.678	3.295	-1.491	2.405	2.202	-0.204	8.59E-01	9.24E-01	
P332401CSTF2	4	-1.38	-1.47	-0.09	-1.22	1.03	-1.43	-0.19	0.859663	9.24E-01	-6.86	0	3.47E+07	1.76E+07	1.58E+07	5.32E+06	5.61E+07	3.74E+07	3.17E+07	1.60E+07	-0.548	-0.535	-1.783	-2.652	-1.125	-1.547	-1.900	-1.328	-1.362	-0.093	8.57E-01	9.24E-01		
P3585461MTPN	5	3.85	3.79	-0.06	-0.75	0.64	3.82	-0.18	0.859804	9.24E-01	-6.86	0	9.16E+08	4.94E+08	6.01E+08	6.86E+08	1.17E+09	1.10E+09	1.27E+09	7.92E+08	3.981	4.043	3.479	3.887	3.403	3.713	3.703	3.444	3.848	3.791	-0.057	8.58E-01	9.24E-01	
Q1655991NF2	3	-3.53	-3.43	0.11	-1.33	1.54	-3.45	0.18	0.861716	9.26E-01	-6.41	3	NA	NA	1.99E+06	NA	1.39E+07	1.52E+07	7.63E+06	3.61E+06	NA	-3.533	NA	-3.205	-2.950	-4.057	-3.489	-3.533	-3.426	0.108	8.62E-01	9.26E-01		
Q6F5E3ICARL2	4	-1.82	-1.90	-0.08	-1.12	0.96	-1.86	-0.18	0.864019	9.27E-01	-6.87	0	2.40E+07	4.33E+06	1.85E+07	7.50E+06	1.97E+07	4.17E+07	2.31E+07	1.78E+07	-1.058	-2.466	-1.557	-2.399	-2.682	-1.980	-2.375	-1.169	-1.820	-1.901	-0.081	8.64E-01	9.27E-01	
Q817C9IAAMDC	2	-0.46	-0.51	-0.06	-0.81	0.69	-0.48	-0.17	0.869504	9.31E-01	-6.87	0	3.62E+07	1.83E+07	3.01E+07	3.85E+07	8.35E+07	6.99E+07	5.70E+07	4.18E+07	-0.492	-0.481	-0.854	0.005	-0.532	-0.577	-1.008	0.070	-0.456	-0.512	-0.056	8.70E-01	9.31E-01	
P0111FRASN	3	-3.44	-3.31	0.14	-2.05	2.32	-3.34	0.16	0.881706	9.38E-01	-6.39	4	NA	NA	5.04E+06	NA	1.98E+07	1.65E+07	6.01E+06	NA	NA	-3.441	NA	-2.676	-2.821	-4.421	NA	-3.441	-3.306	0.135	8.82E-01	9.38E-01		
P684021PA1B2	8	2.89	2.75	-0.13	-2.03	1.76	2.82	-0.16	0.878025	9.38E-01	-6.87	0	7.51E+08	3.59E+08	8.80E+08	4.47E+07	1.10E+09	7.72E+08	8.70E+08	1.06E+08	3.707	3.605	4.031	0.207	3.312	3.156	3.127	1.424	2.888	2.755	-0.133	8.78E-01	9.38E-01	
Q9LUM501NFU1	3	0.35	0.25	-0.11	-1.66	1.45	0.30	-0.15	0.881831	9.38E-01	-6.87	0	1.08E+08	5.12E+07	1.20E+08	1.10E+07	1.95E+08	1.65E+08	1.58E+08	1.95E+07	1.023	0.929	1.150	-1.686	0.734	0.757	0.538	-1.037	0.354	0.248	-0.106	8.82E-01	9.38E-01	
zz1Y-FGC2Cant00284l	14	0.59	0.80	0.22	-3.13	3.56	0.68	0.16	0.880991	9.38E-01	-6.62	3	NA	NA	1.98E+07	8.92E+07	1.87E+08	NA	NA	5.01E+07	2.76E+08	NA	-1.084	0.718	2.133	NA	NA	-1.205	2.813	0.589	0.804	0.215	8.81E-01	9.38E-01
Q9NHFH31NF43	2	-0.47	-0.26	0.21	-3.01	3.44	-0.35	0.15	0.884244	9.40E-01	-6.79	1	NA	NA	5.42E+07	1.42E+08	2.29E+06	2.12E+08	1.45E+08	1.62E+08	4.96E+06	NA	1.006	1.390	-3.799	0.861	0.561	0.581	-3.028	-0.468	-0.256	0.211	8.84E-01	9.40E-01
P497731HINT1	3	-0.60	-0.73	-0.13	-2.21	1.95	-0.67	-0.15	0.889971	9.40E-01	-6.73	2	1.36E+08	9.27E+06	1.64E+07	NA	1.27E+08	7.08E+07	3.51E+07	NA	1.347	-1.419	-1.733	NA	-0.099	-0.557	-1.743	NA	-0.602	-0.734	-0.132	8.86E-01	9.40E-01	
P248211TE1NA	3	-3.97	-3.89	0.07	-1.21	1.36	-3.92	0.14	0.892745	9.43E-01	-6.62	3	2.80E+06	NA	3.65E+06	NA	7.03E+06	5.99E+06	NA	4.82E+06	-4.031	NA	-3.908	NA	-4.220	-4.394	NA	-3.070	-3.969	-3.895	0.075	8.93E-01	9.43E-01	
P593271TFD52	4	2.66	2.72	0.06	-0.84	0.95	2.69	0.14	0.893396	9.43E-01	-6.87	0	6.88E+08	1.03E+08	3.27E+08	2.58E+08	5.83E+08	7.11E+08	8.08E+08	2.17E+08	3.585	1.895	2.599	2.567	2.365	3.028	3.014	2.464	2.661	2.718	0.056	8.90E-01	9.43E-01	
Q4572201PLPR4	2	-3.16	-3.07	0.09	-1.40	1.58	-3.11	0.14	0.891639	9.43E-01	-6.8																							

1	ProteinName	# Peptides	Ctrl	Group	log2FC	C.I.L	C.I.R	Ave Expr	t	P.Value	adj.P.Val	B	nrnAs	M.3_Co ntrol.ra w	M.2_Co ntrol.ra w	M.4_Co ntrol.ra w	M.1_Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3_C ontr.transf	M.2_C ontr.transf	M.4_C ontr.transf	M.1_C ontr.transf	E4.2_Group _1.tran sf	E4.4_Group _1.tran sf	E4.3_Group _1.tran sf	E4.1_G roup_1.tran sf	pseud o.Ctrl	pseud o.Grou p1	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val
660	P95637HIF6	2	-0.95	-0.90	0.05	-1.93	2.03	-0.91	0.06	0.952955	3.77E-01	-6.40	4.259E+07	NA	NA	3.01E+08	3.55E+07	8.63E+07	7.93E+07	3.28E+07	NA	-0.951	NA	NA	NA	-0.482	-0.381	-1.844	NA	-0.951	-0.902	0.049	9.52E-01	9.77E-01
661	Q96X68HINT2	3	1.48	1.52	0.04	-1.46	1.55	1.50	0.07	0.948456	3.77E-01	-6.88	0.179E+08	9.65E+07	3.01E+08	3.55E+07	4.87E+08	3.98E+08	4.08E+08	3.68E+07	1.720	1.800	2.478	-0.090	2.096	2.126	1.977	-0.115	1.477	1.521	0.044	9.48E-01	9.77E-01	
662	Q9NQR4HINT2	2	-3.55	-3.58	-0.03	-1.02	0.97	-3.57	-0.06	0.952247	3.77E-01	-6.63	3.39E+06	1.67E+06	5.46E+06	NA	3.39E+06	1.08E+07	1.13E+07	NA	NA	-3.775	-3.325	NA	-2.368	-1.978	-3.463	NA	-3.550	-3.576	-0.026	9.52E-01	9.77E-01	
663	Q9UC2EPIFR	10	0.32	0.35	0.02	-0.74	0.78	0.94	0.07	0.94769	3.77E-01	-6.88	0.110E+08	6.13E+07	1.40E+08	4.13E+07	2.05E+08	1.80E+08	2.08E+08	8.70E+07	1.053	1.175	1.367	0.101	0.805	0.894	0.954	1.135	0.924	0.947	0.023	9.48E-01	9.77E-01	
664	zz1Y-FGC2Ccm00349I	8	2.86	2.84	-0.03	-1.05	0.99	2.85	-0.06	0.949841	3.77E-01	-6.88	0.587E+08	1.60E+08	7.49E+08	1.45E+08	6.39E+08	6.39E+08	6.64E+08	3.76E+08	3.367	2.497	3.797	1.796	2.503	2.861	2.716	3.260	2.864	2.835	-0.029	9.50E-01	9.77E-01	
665	Q12769SCFNT1	3	3.60	3.63	0.03	-1.13	1.19	3.61	0.06	0.955263	3.78E-01	-6.88	0.822E+08	5.20E+08	1.02E+09	1.94E+08	1.88E+09	1.31E+09	1.42E+09	2.58E+08	3.832	4.113	4.251	2.186	3.944	3.974	3.871	2.713	3.596	3.625	0.030	9.55E-01	9.78E-01	
666	Q13522PPFRIA	2	0.10	0.18	0.08	-3.32	3.48	0.14	0.06	0.956539	3.78E-01	-6.63	3.146E+08	3.11E+06	1.88E+08	NA	NA	1.09E+08	1.31E+08	NA	1.441	-2.921	1.794	NA	NA	0.108	0.260	NA	0.104	0.184	0.080	9.57E-01	9.78E-01	
667	Q95747IDXSF1	6	0.01	0.00	-0.02	-0.78	0.74	0.01	-0.05	0.958635	3.79E-01	-6.88	0.356E+07	3.63E+07	7.37E+07	3.01E+07	1.59E+08	8.67E+07	1.11E+08	3.41E+07	-0.514	0.455	0.442	-0.325	0.416	-0.205	0.001	-0.226	0.014	-0.004	-0.018	9.59E-01	9.79E-01	
668	Q13404IJBZV1	5	-0.77	-0.74	0.03	-1.22	1.27	-0.75	0.05	0.960702	3.79E-01	-6.43	3.1NA	NA	3.20E+07	NA	6.32E+07	5.73E+07	6.51E+07	3.19E+07	NA	NA	-0.768	NA	-0.946	-0.894	-0.805	-0.330	-0.226	-0.768	-0.741	0.026	9.61E-01	9.79E-01
669	Q95191DDP2	2	0.27	0.30	0.02	-1.03	1.08	0.29	0.05	0.963576	3.80E-01	-6.88	0.364E+07	1.77E+07	1.03E+08	9.19E+07	1.25E+08	1.10E+08	1.15E+08	7.55E+07	-0.491	-0.527	0.927	1.178	0.064	0.130	0.063	0.928	0.274	0.296	0.022	9.64E-01	9.80E-01	
670	Q99426TBCB	2	-0.25	-0.28	-0.03	-1.62	1.56	-0.27	-0.05	0.964741	3.80E-01	-6.68	2.130E+07	NA	NA	4.20E+07	1.47E+08	3.10E+07	1.27E+08	4.47E+07	-0.616	NA	NA	0.123	0.312	-1.799	0.212	0.166	-0.246	-0.277	-0.031	9.65E-01	9.80E-01	
671	Q9N1J2PFA49B	8	0.65	0.62	-0.03	-1.51	1.45	0.63	-0.04	0.965796	3.80E-01	-6.88	0.102E+08	9.03E+07	1.47E+08	1.26E+07	2.41E+08	1.74E+08	1.48E+08	4.43E+07	0.946	1.707	1.437	-1.495	1.048	0.837	0.443	0.153	0.649	0.620	-0.029	9.66E-01	9.80E-01	
672	P1940PAGP1	4	2.29	2.31	0.02	-1.00	1.03	2.30	0.03	0.973799	3.83E-01	-6.88	0.532E+08	1.71E+08	2.68E+08	6.28E+07	6.77E+08	4.29E+08	4.68E+08	1.82E+08	3.231	2.587	2.308	1.038	2.588	2.241	2.188	2.210	2.291	2.320	0.015	9.74E-01	9.83E-01	
673	Q13765NACA	2	-1.28	-1.26	0.02	-1.08	1.12	-1.27	0.04	0.971524	3.83E-01	-6.81	1.340E+07	NA	1.72E+07	1.10E+07	3.29E+07	4.57E+07	4.11E+07	3.03E+07	-0.497	NA	-1.662	-1.687	-1.922	-1.235	-1.604	-0.397	-1.262	-1.264	0.018	9.72E-01	9.83E-01	
674	Q9B711PACN1	3	-0.91	-0.93	-0.02	-1.51	1.47	-0.92	-0.03	0.974682	3.83E-01	-6.54	4.248E+07	NA	3.55E+07	NA	1.01E+08	NA	4.36E+07	NA	-1.011	NA	-0.617	NA	-0.263	NA	-1.144	NA	-0.894	-0.834	-0.020	9.75E-01	9.83E-01	
675	zz1Y-FGC2Ccm00248I	5	3.46	3.45	-0.01	-0.85	0.83	3.45	-0.03	0.974906	3.83E-01	-6.88	0.638E+08	2.94E+08	3.79E+08	3.87E+08	9.35E+08	1.67E+09	1.04E+09	3.07E+09	3.533	3.328	3.854	3.117	3.072	4.352	3.392	2.968	3.498	3.446	-0.012	9.75E-01	9.83E-01	
676	P30043BLVFB	2	2.07	2.09	0.02	-2.34	2.39	2.08	0.02	0.983432	3.88E-01	-6.88	0.519E+08	2.68E+08	5.71E+08	1.24E+07	5.29E+08	5.47E+08	6.40E+08	7.23E+07	3.195	3.203	3.404	-1.526	2.219	2.621	2.660	0.865	2.069	2.091	0.022	9.83E-01	9.88E-01	
677	zz1Y-FGC2Ccm00193I	4	-1.21	-1.22	-0.01	-1.00	0.98	-1.22	-0.02	0.983043	3.88E-01	-6.81	1.154E+07	1.08E+07	3.21E+07	NA	4.00E+07	3.10E+07	7.84E+07	2.15E+07	-1.676	-1.304	-0.759	NA	-1.628	-1.840	-0.524	-0.898	-1.213	-1.222	-0.009	9.83E-01	9.88E-01	
678	zz1Y-FGC2Ccm00332I	3	-0.53	-0.54	-0.01	-0.92	0.90	-0.53	-0.02	0.980861	3.89E-01	-6.88	0.401E+07	2.09E+07	5.55E+07	1.26E+07	1.19E+08	7.90E+07	6.77E+07	1.97E+07	-0.348	-0.304	0.030	-1.497	0.000	-0.385	-0.747	-1.027	-0.530	-0.540	0.010	9.81E-01	9.88E-01	
679	zz1Y-FGC2Ccm00319I	12	4.62	4.61	-0.01	-0.66	0.65	4.62	-0.02	0.985122	3.98E-01	-6.88	0.166E+09	6.61E+08	1.62E+09	9.41E+08	2.75E+09	2.22E+09	2.55E+09	7.28E+08	4.803	4.443	4.917	4.313	4.680	4.799	4.755	4.221	4.619	4.514	-0.006	9.85E-01	9.88E-01	
680	P61604CH10	6	4.93	4.93	0.00	-0.76	0.77	4.93	0.01	0.989914	3.91E-01	-6.88	0.174E+09	6.31E+08	2.04E+09	1.84E+09	2.79E+09	3.66E+09	2.46E+09	1.05E+09	4.871	4.378	5.244	5.218	4.698	5.577	4.705	4.749	4.928	4.932	0.004	9.90E-01	9.91E-01	
681	Q14637HIFG1	3	-2.74	-2.74	-0.01	-1.36	1.35	-2.74	-0.01	0.993025	3.93E-01	-6.81	1.118E+07	2.90E+06	6.12E+06	NA	1.72E+07	4.19E+07	1.66E+07	2.89E+06	-2.040	-3.013	-3.160	NA	-2.886	-1.376	-2.877	-3.834	-2.738	-2.743	-0.005	9.93E-01	9.93E-01	
682	REV_Q81W99ITEX2	2	NA	-2.23	NA	NA	NA	-2.23	NA	NA	NA	NA	4.1NA	NA	NA	NA	2.04E+07	2.24E+07	4.17E+07	7.41E+06	NA	NA	NA	NA	-2.632	-2.347	-1.480	-2.444	-4.066	-2.226	1.840	0.00E+00	0.00E+00	
683	Q00267SPTSH	2	NA	-3.08	NA	NA	NA	-3.08	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.74E+07	5.90E+06	2.29E+07	6.62E+06	NA	NA	NA	NA	-2.670	-4.444	-2.395	-2.609	-4.066	-3.080	0.966	0.00E+00	0.00E+00	
684	Q00299CLIC1	6	NA	-0.06	NA	NA	NA	-0.06	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.40E+08	8.99E+07	1.13E+08	3.00E+07	NA	NA	NA	NA	0.240	-0.085	0.036	-0.415	-4.066	-0.056	4.010	0.00E+00	0.00E+00	
685	Q00391QSDQX1	7	NA	-0.51	NA	NA	NA	-0.51	NA	NA	NA	NA	4.1NA	NA	NA	NA	8.76E+07	2.62E+07	1.78E+08	3.47E+07	NA	NA	NA	NA	-0.460	-2.103	0.719	-0.201	-4.066	-5.512	3.555	0.00E+00	0.00E+00	
686	Q00623CCN1	3	NA	0.65	NA	NA	NA	0.65	NA	NA	NA	NA	4.1NA	NA	NA	NA	2.02E+08	1.67E+08	9.95E+07	9.02E+07	NA	NA	NA	NA	0.786	0.778	-0.161	1.187	-4.066	0.547	4.714	0.00E+00	0.00E+00	
687	Q146721ADA10	3	NA	0.60	NA	NA	NA	0.60	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.42E+08	1.51E+08	2.66E+08	4.54E+07	NA	NA	NA	NA	0.265	0.617	1.327	0.198	0.649	4.665	0.00E+00	0.00E+00		
688	Q43237DCIL2	2	NA	-1.97	NA	NA	NA	-1.97	NA	NA	NA	NA	4.1NA	NA	NA	NA	4.00E+07	1.28E+07	2.95E+07	2.01E+07	NA	NA	NA	NA	-1.629	-3.214	-2.059	-0.993	-4.066	-1.973	2.093	0.00E+00	0.00E+00	
689	Q43390HMFPR	3	NA	-2.95	NA	NA	NA	-2.95	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.93E+07	8.82E+06	1.78E+07	7.10E+06	NA	NA	NA	NA	-2.713	-3.794	-2.773	-2.506	-4.066	-2.947	1.119	0.00E+00	0.00E+00	
690	Q43707ACTN4	10	NA	0.75	NA	NA	NA	0.75	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.79E+08	1.74E+08	1.82E+08	6.96E+07	NA	NA	NA	NA	0.599	0.841	0.754	0.811	-4.066	0.751	4.617	0.00E+00	0.00E+00	
691	Q60749SNX2	2	NA	-3.82	NA	NA	NA	-3.82	NA	NA	NA	NA	4.1NA	NA	NA	NA	8.60E+06	4.56E+06	8.93E+06	4.19E+06	NA	NA	NA	NA	-3.920	-4.252	-3.819	-3.275	-4.066	-3.816	0.250	0.00E+00	0.00E+00	
692	Q75533SFB1	2	NA	-2.88	NA	NA	NA	-2.88	NA	NA	NA	NA	4.1NA	NA	NA	NA	2.80E+07	1.59E+07	1.57E+07	3.59E+06	NA	NA	NA	NA	-2.169	-2.914	-2.965	-3.498	-4.066	-2.894	1.182	0.00E+00	0.00E+00	
693	Q75787RENR	5	NA	-0.51	NA	NA	NA	-0.51	NA	NA	NA	NA	4.1NA	NA	NA	NA	7.12E+07	3.9E+07	1.99E+08	1.60E+07	NA	NA	NA	NA	-0.769	-0.498	0.551	-1.324	-4.066	-5.510	3.566	0.00E+00	0.00E+00	
694	Q96084PPRS23	7	NA	3.76	NA	NA	NA	3.76	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.51E+09	1.86E+09	1.14E+09	3.56E+08	NA	NA	NA	NA	3.780	4.526	3.636	3.161	-4.066	3.756	7.822	0.00E+00	0.00E+00	
695	Q95160TNF15	2	NA	-2.63	NA	NA	NA	-2.63	NA	NA	NA	NA	4.1NA	NA	NA	NA	1.99E+07	1.66E+0																

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M.3_Co ntrol.ra w	M.2_Co ntrol.ra w	M.4_Co ntrol.ra w	M.1_Co ntrol.ra w	E4.2_Gr oup_Tr aw	E4.4_Gr oup_Tr aw	E4.3_Gr oup_Tr aw	E4.1_Gr oup_Tr aw	M.3_C ontrol. transf	M.2_C ontrol. transf	M.4_C ontrol. transf	M.1_C ontrol. transf	E4.2_ Group _1tran sf	E4.4_ Group _1tran sf	E4.3_ Group _1tran sf	E4.1_G roup_1 tran sf	pseud o.Ctrl	pseud o.Grou p1	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val
707	P06748NFM	2	NA	-1.41	NA	NA	NA	-1.41	NA	NA	NA	NA	4	NA	NA	NA	NA	3.61E+07	4.43E+07	6.40E+07	1.21E+07	NA	NA	NA	NA	-1.782	-1.286	-0.832	-1.732	-4.066	-1.408	2.658	0.00E+00	0.00E+00
708	P06753TFM3	3	NA	-4.28	NA	NA	NA	-4.28	NA	NA	NA	NA	4	NA	NA	NA	NA	4.74E+06	8.54E+06	7.16E+06	2.06E+06	NA	NA	NA	NA	-4.807	-3.844	-4.153	-4.303	-4.066	-4.277	-0.211	0.00E+00	0.00E+00
709	P06756ITAV	5	NA	-1.41	NA	NA	NA	-1.41	NA	NA	NA	NA	4	NA	NA	NA	NA	5.80E+07	4.51E+07	4.12E+07	1.16E+07	NA	NA	NA	NA	-1.074	-1.257	-1.501	-1.752	-4.066	-1.406	2.660	0.00E+00	0.00E+00
710	P07858CATE	3	NA	0.09	NA	NA	NA	0.09	NA	NA	NA	NA	4	NA	NA	NA	NA	1.59E+08	1.36E+08	1.19E+08	2.67E+07	NA	NA	NA	NA	0.415	0.454	0.074	-0.582	-4.066	0.090	4.766	0.00E+00	0.00E+00
711	P07942LAM61	19	NA	1.94	NA	NA	NA	1.94	NA	NA	NA	NA	4	NA	NA	NA	NA	5.55E+06	2.09E+08	4.74E+08	1.73E+08	NA	NA	NA	NA	2.233	1.130	2.204	2.134	-4.066	1.940	6.006	0.00E+00	0.00E+00
712	P07996TFP1	20	NA	5.01	NA	NA	NA	5.01	NA	NA	NA	NA	4	NA	NA	NA	NA	2.24E+06	2.13E+09	4.20E+09	1.66E+09	NA	NA	NA	NA	4.370	4.736	5.514	5.416	-4.066	5.093	9.076	0.00E+00	0.00E+00
713	P08253IMF2	8	NA	2.53	NA	NA	NA	2.53	NA	NA	NA	NA	4	NA	NA	NA	NA	7.39E+08	3.76E+08	8.35E+08	1.96E+08	NA	NA	NA	NA	2.718	2.039	3.063	2.313	-4.066	2.533	6.599	0.00E+00	0.00E+00
714	P08572CDA2	12	NA	1.40	NA	NA	NA	1.40	NA	NA	NA	NA	4	NA	NA	NA	NA	2.00E+08	3.03E+08	2.70E+08	1.36E+08	NA	NA	NA	NA	0.757	1.702	1.354	1.778	-4.066	1.401	5.467	0.00E+00	0.00E+00
715	P08648IT A5	7	NA	0.47	NA	NA	NA	0.47	NA	NA	NA	NA	4	NA	NA	NA	NA	1.63E+08	1.41E+08	1.43E+08	5.37E+07	NA	NA	NA	NA	0.519	0.539	0.389	0.434	-4.066	0.465	4.531	0.00E+00	0.00E+00
716	P08856IRSSA	2	NA	-2.92	NA	NA	NA	-2.92	NA	NA	NA	NA	4	NA	NA	NA	NA	1.39E+07	8.54E+06	2.91E+07	6.65E+06	NA	NA	NA	NA	-3.202	-3.844	-2.027	-2.601	-4.066	-2.918	1.148	0.00E+00	0.00E+00
717	P10321IC07	2	NA	-0.23	NA	NA	NA	-0.23	NA	NA	NA	NA	4	NA	NA	NA	NA	1.03E+08	6.16E+07	1.15E+08	4.05E+07	NA	NA	NA	NA	-0.224	-0.773	0.056	0.022	-4.066	-0.230	3.836	0.00E+00	0.00E+00
718	P10644IKAP0	2	NA	-3.31	NA	NA	NA	-3.31	NA	NA	NA	NA	4	NA	NA	NA	NA	1.52E+07	5.19E+06	1.36E+07	7.86E+06	NA	NA	NA	NA	-3.069	-4.618	-3.184	-2.359	-4.066	-3.308	0.759	0.00E+00	0.00E+00
719	P1047LAMC1	23	NA	4.30	NA	NA	NA	4.30	NA	NA	NA	NA	4	NA	NA	NA	NA	2.29E+09	1.69E+09	2.00E+09	6.96E+08	NA	NA	NA	NA	4.401	4.272	4.391	4.156	-4.066	4.305	8.371	0.00E+00	0.00E+00
720	P12643BMP2	2	NA	-3.41	NA	NA	NA	-3.41	NA	NA	NA	NA	4	NA	NA	NA	NA	1.69E+07	1.72E+07	7.60E+06	2.74E+06	NA	NA	NA	NA	-2.923	-2.753	-4.064	-3.993	-4.066	-3.408	0.859	0.00E+00	0.00E+00
721	P13493FIN1	3	NA	0.08	NA	NA	NA	0.08	NA	NA	NA	NA	4	NA	NA	NA	NA	9.69E+07	9.29E+07	1.19E+08	6.16E+07	NA	NA	NA	NA	-0.311	-0.117	0.115	0.630	-4.066	0.079	4.146	0.00E+00	0.00E+00
722	P13639IEF2	5	NA	-0.85	NA	NA	NA	-0.85	NA	NA	NA	NA	4	NA	NA	NA	NA	5.79E+07	5.71E+07	7.53E+07	2.22E+07	NA	NA	NA	NA	-1.079	-0.890	-0.585	-0.951	-4.066	-0.951	3.216	0.00E+00	0.00E+00
723	P14209ICD99	2	NA	-1.38	NA	NA	NA	-1.38	NA	NA	NA	NA	4	NA	NA	NA	NA	3.80E+07	2.84E+07	4.83E+07	2.66E+07	NA	NA	NA	NA	-1.705	-1.976	-1.258	-0.590	-4.066	-1.382	2.684	0.00E+00	0.00E+00
724	P14543IND1	16	NA	2.16	NA	NA	NA	2.16	NA	NA	NA	NA	4	NA	NA	NA	NA	6.89E+08	3.08E+08	4.97E+08	1.61E+08	NA	NA	NA	NA	2.612	1.729	2.277	2.025	-4.066	2.161	6.227	0.00E+00	0.00E+00
725	P14625IENPL	4	NA	-3.40	NA	NA	NA	-3.40	NA	NA	NA	NA	4	NA	NA	NA	NA	1.46E+07	5.09E+06	1.25E+07	7.05E+06	NA	NA	NA	NA	-3.130	-4.647	-3.308	-2.518	-4.066	-3.401	0.655	0.00E+00	0.00E+00
726	P14966IHNPPL	5	NA	-1.27	NA	NA	NA	-1.27	NA	NA	NA	NA	4	NA	NA	NA	NA	7.80E+07	7.92E+07	3.14E+07	9.01E+06	NA	NA	NA	NA	-0.633	-0.381	-1.913	-2.162	-4.066	-1.272	2.794	0.00E+00	0.00E+00
727	P15144AMPN	3	NA	-2.40	NA	NA	NA	-2.40	NA	NA	NA	NA	4	NA	NA	NA	NA	4.84E+07	1.76E+07	2.78E+07	3.76E+06	NA	NA	NA	NA	-1.346	-2.720	-2.094	-3.432	-4.066	-2.398	1.668	0.00E+00	0.00E+00
728	P15311E2FI	6	NA	-2.41	NA	NA	NA	-2.41	NA	NA	NA	NA	4	NA	NA	NA	NA	3.20E+07	2.84E+07	1.55E+07	6.14E+06	NA	NA	NA	NA	-1.960	-1.975	-2.981	-2.719	-4.066	-2.409	1.657	0.00E+00	0.00E+00
729	P16870ICBPE	7	NA	-1.17	NA	NA	NA	-1.17	NA	NA	NA	NA	4	NA	NA	NA	NA	1.04E+08	5.44E+07	5.03E+07	8.10E+06	NA	NA	NA	NA	-0.201	-0.966	-1.198	-2.315	-4.066	-1.170	2.896	0.00E+00	0.00E+00
730	P17813IEGLN	5	NA	1.90	NA	NA	NA	1.90	NA	NA	NA	NA	4	NA	NA	NA	NA	5.11E+08	3.83E+08	5.08E+08	8.18E+07	NA	NA	NA	NA	2.169	2.069	2.310	1.045	-4.066	1.898	5.964	0.00E+00	0.00E+00
731	P19109ML12A	7	NA	-1.18	NA	NA	NA	-1.18	NA	NA	NA	NA	4	NA	NA	NA	NA	8.80E+07	4.98E+07	4.71E+07	1.11E+07	NA	NA	NA	NA	-0.453	-1.102	-1.296	-1.855	-4.066	-1.176	2.890	0.00E+00	0.00E+00
732	P20700MLN81	7	NA	-1.86	NA	NA	NA	-1.86	NA	NA	NA	NA	4	NA	NA	NA	NA	6.19E+07	2.22E+07	3.21E+07	8.54E+06	NA	NA	NA	NA	-0.980	-2.361	-1.877	-2.239	-4.066	-1.865	2.202	0.00E+00	0.00E+00
733	P21990ITGM2	5	NA	0.48	NA	NA	NA	0.48	NA	NA	NA	NA	4	NA	NA	NA	NA	1.87E+08	1.76E+08	2.26E+08	2.46E+07	NA	NA	NA	NA	0.669	0.860	1.079	-0.699	-4.066	0.477	4.543	0.00E+00	0.00E+00
734	P22392INDKB	2	NA	-1.36	NA	NA	NA	-1.36	NA	NA	NA	NA	4	NA	NA	NA	NA	5.00E+07	3.09E+07	3.39E+07	2.90E+07	NA	NA	NA	NA	-1.296	-1.845	-1.823	-0.462	-4.066	-1.356	2.710	0.00E+00	0.00E+00
735	P26038MDE S	13	NA	1.11	NA	NA	NA	1.11	NA	NA	NA	NA	4	NA	NA	NA	NA	2.34E+08	2.38E+08	1.75E+08	1.04E+08	NA	NA	NA	NA	1.007	1.327	0.693	1.397	-4.066	1.106	5.172	0.00E+00	0.00E+00
736	P27658ICD8A1	3	NA	0.89	NA	NA	NA	0.89	NA	NA	NA	NA	4	NA	NA	NA	NA	1.90E+08	1.65E+08	1.86E+08	9.94E+07	NA	NA	NA	NA	0.691	0.759	0.790	1.327	-4.066	0.892	4.968	0.00E+00	0.00E+00
737	P28799IGFN	2	NA	-0.37	NA	NA	NA	-0.37	NA	NA	NA	NA	4	NA	NA	NA	NA	8.04E+07	7.44E+07	8.21E+07	4.34E+07	NA	NA	NA	NA	-0.588	-0.542	-0.453	0.123	-4.066	-0.365	3.701	0.00E+00	0.00E+00
738	P29279CCN2	3	NA	4.51	NA	NA	NA	4.51	NA	NA	NA	NA	4	NA	NA	NA	NA	2.29E+09	2.78E+09	2.09E+09	6.38E+08	NA	NA	NA	NA	4.405	5.146	4.455	4.029	-4.066	4.509	8.575	0.00E+00	0.00E+00
739	P29629IEFD	6	NA	-1.07	NA	NA	NA	-1.07	NA	NA	NA	NA	4	NA	NA	NA	NA	5.31E+07	5.48E+07	8.92E+07	1.15E+07	NA	NA	NA	NA	-1.206	-0.954	-0.316	-1.811	-4.066	-1.072	2.995	0.00E+00	0.00E+00
740	P29966MAPCS	8	NA	0.57	NA	NA	NA	0.57	NA	NA	NA	NA	4	NA	NA	NA	NA	8.01E+07	1.54E+08	1.82E+08	1.10E+08	NA	NA	NA	NA	-0.594	0.647	0.752	1.475	-4.066	0.570	4.636	0.00E+00	0.00E+00
741	P30626SDRCN	4	NA	-0.15	NA	NA	NA	-0.15	NA	NA	NA	NA	4	NA	NA	NA	NA	1.10E+08	7.99E+07	9.07E+07	4.51E+07	NA	NA	NA	NA	-0.121	-0.368	-0.303	0.178	-4.066	-0.153	3.913	0.00E+00	0.00E+00
742	P31949IS10AB	2	NA	0.86	NA	NA	NA	0.86	NA	NA	NA	NA	4	NA	NA	NA	NA																	

1	ProteinName	# Peptides	Ctrl	Group	log2FC	Cl.L	Cl.R	Ave Expr	t	P.Value	adj.P.Val	B	nrN As	M.3_Co ntrol.ra w	M.2_Co ntrol.ra w	M.4_Co ntrol.ra w	M.1_Co ntrol.ra w	E4.2_Gr oup_1r aw	E4.4_Gr oup_1r aw	E4.3_Gr oup_1r aw	E4.1_Gr oup_1r aw	M.3_C ontr. transf	M.2_C ontr. transf	M.4_C ontr. transf	M.1_C ontr. transf	E4.2_Gr oup_1tran sf	E4.4_Gr oup_1tran sf	E4.3_Gr oup_1tran sf	E4.1_Gr oup_1tran sf	pseud o.Ctrl	pseud o.Group pl	pseud o.log2 FC	pseudo.P. Value	pseudo.adj P.Val
754	P49802RGS7	2	NA	-3.72	NA	NA	NA	-3.72	NA	NA	NA	NA	4	NA	NA	NA	NA	8.93E+06	7.30E+06	9.62E+06	4.35E+06	NA	NA	NA	NA	-3.864	-4.088	-3.708	-3.219	-4.066	-3.719	0.347	0.00E+00	0.00E+00
755	P50281MMVPH	2	NA	-0.73	NA	NA	NA	-0.73	NA	NA	NA	NA	4	NA	NA	NA	NA	6.52E+07	6.58E+07	6.56E+07	2.72E+07	NA	NA	NA	NA	-0.899	-0.669	-0.795	-0.954	-4.066	-0.730	3.337	0.00E+00	0.00E+00
756	P52272HNVFFM	2	NA	-2.99	NA	NA	NA	-2.99	NA	NA	NA	NA	4	NA	NA	NA	NA	7.23E+06	6.06E+06	2.89E+07	154E+07	NA	NA	NA	NA	-4.179	-4.375	-2.036	-1.367	-4.066	-2.994	10.72	0.00E+00	0.00E+00
757	P55011S1A2	2	1.38	NA	NA	NA	NA	-1.38	NA	NA	NA	NA	4	2.66E+07	9.41E+06	2.43E+07	8.53E+06	NA	NA	NA	NA	-0.914	-1.398	-1.164	-2.026	NA	NA	NA	-1.376	-3.762	-2.387	0.00E+00	0.00E+00	
758	P62741PP2A8	4	NA	-3.18	NA	NA	NA	-3.18	NA	NA	NA	NA	4	NA	NA	NA	NA	2.1E+07	1.19E+07	1.65E+07	2.63E+06	NA	NA	NA	NA	-2.584	-3.331	-2.847	-3.951	-4.066	-3.178	0.688	0.00E+00	0.00E+00
759	P62826FRAN	2	NA	-3.76	NA	NA	NA	-3.76	NA	NA	NA	NA	4	NA	NA	NA	NA	6.28E+06	6.88E+06	1.43E+07	3.98E+06	NA	NA	NA	NA	-4.393	-4.179	-3.109	-3.346	-4.066	-3.757	0.309	0.00E+00	0.00E+00
760	P62873IGBE1	3	NA	-2.85	NA	NA	NA	-2.85	NA	NA	NA	NA	4	NA	NA	NA	NA	2.03E+07	157E+07	2.60E+07	3.24E+06	NA	NA	NA	NA	-2.639	-2.301	-2.198	-3.645	-4.066	-2.846	1.220	0.00E+00	0.00E+00
761	P67809YBCK1	5	NA	-2.95	NA	NA	NA	-2.95	NA	NA	NA	NA	4	NA	NA	NA	NA	2.30E+07	142E+07	1.46E+07	1.42E+07	NA	NA	NA	NA	-2.593	-3.051	-3.074	-1.496	-4.066	-2.554	1.513	0.00E+00	0.00E+00
762	P78471GCTC1	2	NA	-3.52	NA	NA	NA	-3.52	NA	NA	NA	NA	4	NA	NA	NA	NA	3.04E+07	5.93E+06	7.61E+06	3.4E+06	NA	NA	NA	NA	-2.038	-4.411	-4.060	-3.573	-4.066	-3.521	0.546	0.00E+00	0.00E+00
763	P84090IERH	2	NA	-1.75	NA	NA	NA	-1.75	NA	NA	NA	NA	4	NA	NA	NA	NA	1.06E+07	3.19E+07	4.82E+07	3.19E+07	NA	NA	NA	NA	-3.616	-1.797	-1.261	-0.342	-4.066	-1.754	2.312	0.00E+00	0.00E+00
764	P89082DAE2	5	NA	-2.51	NA	NA	NA	-2.51	NA	NA	NA	NA	4	NA	NA	NA	NA	2.97E+07	1.16E+07	2.42E+07	8.39E+06	NA	NA	NA	NA	-2.125	-3.345	-2.308	-2.267	-4.066	-2.511	1.555	0.00E+00	0.00E+00
765	P89179PEM3	2	NA	-1.46	NA	NA	NA	-1.46	NA	NA	NA	NA	4	NA	NA	NA	NA	6.95E+07	4.40E+07	3.53E+07	1.49E+07	NA	NA	NA	NA	-0.684	-1.297	-2.193	-1.477	-4.066	-1.461	2.605	0.00E+00	0.00E+00
766	Q01638ILFL1	3	NA	1.65	NA	NA	NA	1.65	NA	NA	NA	NA	4	NA	NA	NA	NA	3.95E+08	2.91E+08	3.93E+08	1.05E+08	NA	NA	NA	NA	1.562	1.588	1.659	1.411	-4.066	1.555	5.621	0.00E+00	0.00E+00
767	Q01252LLNMB2	4	NA	-2.59	NA	NA	NA	-2.59	NA	NA	NA	NA	4	NA	NA	NA	NA	1.96E+07	1.94E+07	1.53E+07	9.4E+06	NA	NA	NA	NA	-2.692	-2.568	-2.997	-2.098	-4.066	-2.589	1.477	0.00E+00	0.00E+00
768	Q08629TICN1	5	NA	-0.50	NA	NA	NA	-0.50	NA	NA	NA	NA	4	NA	NA	NA	NA	7.70E+07	1.16E+08	4.64E+07	3.38E+07	NA	NA	NA	NA	-0.653	0.204	-1.321	-0.240	-4.066	-0.502	3.564	0.00E+00	0.00E+00
769	Q08753FFPD	3	NA	-2.10	NA	NA	NA	-2.10	NA	NA	NA	NA	4	NA	NA	NA	NA	4.64E+07	3.99E+07	1.93E+07	7.77E+06	NA	NA	NA	NA	-1.409	-1.447	-3.160	-2.376	-4.066	-2.098	1.968	0.00E+00	0.00E+00
770	Q12805FBLN3	3	NA	-0.98	NA	NA	NA	-0.98	NA	NA	NA	NA	4	NA	NA	NA	NA	5.17E+07	3.51E+07	7.62E+07	2.90E+07	NA	NA	NA	NA	-1.248	-1.646	-0.566	-0.465	-4.066	-0.981	3.085	0.00E+00	0.00E+00
771	Q12906ILF3	4	NA	-1.53	NA	NA	NA	-1.53	NA	NA	NA	NA	4	NA	NA	NA	NA	3.07E+07	2.85E+07	3.29E+07	3.32E+07	NA	NA	NA	NA	-2.023	-1.973	-1.842	-0.267	-4.066	-1.526	2.540	0.00E+00	0.00E+00
772	Q12631TIFB1	5	NA	-0.01	NA	NA	NA	-0.01	NA	NA	NA	NA	4	NA	NA	NA	NA	9.02E+07	9.38E+07	1.18E+08	5.19E+07	NA	NA	NA	NA	-0.416	-0.118	0.092	0.384	-4.066	-0.015	4.051	0.00E+00	0.00E+00
773	Q13435SFB2	4	NA	-2.20	NA	NA	NA	-2.20	NA	NA	NA	NA	4	NA	NA	NA	NA	3.71E+07	2.04E+07	3.25E+07	6.27E+06	NA	NA	NA	NA	-1.742	-2.492	-1.960	-2.688	-4.066	-2.196	1.871	0.00E+00	0.00E+00
774	Q13444ADA15	6	NA	1.11	NA	NA	NA	1.11	NA	NA	NA	NA	4	NA	NA	NA	NA	2.73E+08	2.04E+08	2.07E+08	8.99E+07	NA	NA	NA	NA	1.234	1.086	0.947	1.181	-4.066	1.112	5.178	0.00E+00	0.00E+00
775	Q13740CD165	6	NA	-0.21	NA	NA	NA	-0.21	NA	NA	NA	NA	4	NA	NA	NA	NA	1.57E+08	1.75E+08	9.76E+07	1.07E+07	NA	NA	NA	NA	0.412	0.647	-0.191	-1.906	-4.066	-0.210	3.856	0.00E+00	0.00E+00
776	Q13753LAMC2	18	NA	1.28	NA	NA	NA	1.28	NA	NA	NA	NA	4	NA	NA	NA	NA	3.31E+08	2.55E+08	2.90E+08	7.46E+07	NA	NA	NA	NA	1.522	1.437	1.237	0.912	-4.066	1.277	5.343	0.00E+00	0.00E+00
777	Q14767LTP2	9	NA	3.31	NA	NA	NA	3.31	NA	NA	NA	NA	4	NA	NA	NA	NA	1.08E+09	7.84E+08	8.24E+08	5.19E+08	NA	NA	NA	NA	3.291	3.181	3.044	3.712	-4.066	3.307	7.373	0.00E+00	0.00E+00
778	Q16075EEA1	2	NA	-3.12	NA	NA	NA	-3.12	NA	NA	NA	NA	4	NA	NA	NA	NA	7.54E+06	2.84E+07	2.67E+07	2.17E+06	NA	NA	NA	NA	-4.117	-1.977	-2.165	-4.229	-4.066	-3.119	0.947	0.00E+00	0.00E+00
779	Q16942ZYX	9	NA	1.49	NA	NA	NA	1.49	NA	NA	NA	NA	4	NA	NA	NA	NA	2.58E+08	2.55E+08	3.02E+08	1.40E+08	NA	NA	NA	NA	1.163	1.437	1.523	1.824	-4.066	1.484	5.551	0.00E+00	0.00E+00
780	Q16363LAM4	2	NA	-2.82	NA	NA	NA	-2.82	NA	NA	NA	NA	4	NA	NA	NA	NA	1.37E+07	3.30E+07	2.78E+07	2.16E+06	NA	NA	NA	NA	-3.222	-1.741	-2.099	-4.233	-4.066	-2.824	1.242	0.00E+00	0.00E+00
781	Q16543CDC37	2	NA	-2.48	NA	NA	NA	-2.48	NA	NA	NA	NA	4	NA	NA	NA	NA	1.91E+07	2.20E+07	1.60E+07	1.1E+07	NA	NA	NA	NA	-2.613	-2.375	-2.932	-1.813	-4.066	-2.843	1.593	0.00E+00	0.00E+00
782	Q16610ECM1	8	NA	-1.00	NA	NA	NA	-1.00	NA	NA	NA	NA	4	NA	NA	NA	NA	3.61E+07	3.56E+07	8.45E+07	3.53E+07	NA	NA	NA	NA	-1.781	-1.624	-0.411	-0.177	-4.066	-0.998	3.068	0.00E+00	0.00E+00
783	Q16658FSCN1	6	NA	-0.12	NA	NA	NA	-0.12	NA	NA	NA	NA	4	NA	NA	NA	NA	7.02E+07	8.34E+07	1.21E+08	5.60E+07	NA	NA	NA	NA	-0.790	-0.301	0.134	0.494	-4.066	-0.116	3.950	0.00E+00	0.00E+00
784	Q26213CAVN1	4	NA	1.51	NA	NA	NA	1.51	NA	NA	NA	NA	4	NA	NA	NA	NA	3.51E+08	3.91E+08	3.34E+08	6.22E+07	NA	NA	NA	NA	1.609	2.101	1.674	0.646	-4.066	1.508	5.574	0.00E+00	0.00E+00
785	Q27270TARSH	8	NA	0.62	NA	NA	NA	0.62	NA	NA	NA	NA	4	NA	NA	NA	NA	1.64E+08	1.51E+08	1.48E+08	7.67E+07	NA	NA	NA	NA	0.473	0.617	0.438	0.951	-4.066	0.620	4.686	0.00E+00	0.00E+00
786	Q86U10A8	3	NA	-1.16	NA	NA	NA	-1.16	NA	NA	NA	NA	4	NA	NA	NA	NA	6.77E+07	3.76E+07	3.44E+07	2.89E+07	NA	NA	NA	NA	-0.845	-1.540	-1.774	-0.468	-4.066	-1.167	2.909	0.00E+00	0.00E+00
787	Q8VDF2AHNK2	5	NA	-2.79	NA	NA	NA	-2.79	NA	NA	NA	NA	4	NA	NA	NA	NA	1.76E+07	1.61E+07	1.06E+07	1.08E+07	NA	NA	NA	NA	-2.849	-2.862	-3.557	-1.904	-4.066	-2.793	1.273	0.00E+00	0.00E+00
788	Q8WUB1SULF1	3	NA	-3.38	NA	NA	NA	-3.38	NA	NA	NA	NA	4	NA	NA	NA	NA	1.13E+07	2.16E+07	3.89E+06	7.02E+06	NA	NA	NA	NA	-3.516	-2.399	-5.077	-2.524	-4.066	-3.679	0.687	0.00E+00	0.00E+00
789	Q8NIX71SPART	2	NA	-1.60	NA	NA	NA	-1.60	NA	NA	NA	NA	4	NA	NA	NA	NA	2.76E+07	4.52E+07	3.04E+07	1.97E+07	NA	NA	NA	NA	-2.180	-1.253	-1.963	-1.023	-4.066	-1.605	2.461	0.00E+00	0.00E+00
790	Q8NBJAIGDLM1	5	NA	-0.41	NA	NA	NA	-0.41	NA	NA	NA	NA	4	NA	NA	NA	NA	4.06E+07	7.81E+07	1.15E+08	4.94E+07	NA	NA	NA	NA	-1.607	-0.404	0.061	0.313	-4.066	-0.409	3.657	0.00E+00	0.00E+00
791	Q8W2791ROBO4	7	NA	1.98	NA	NA	NA	1.98	NA	NA	NA	NA	4	NA	NA	NA	NA	5.09E+08	4.04E+08	2.85E+08	1.78E+08	NA	NA	NA	NA	2.163	2.151	1.435	2.174	-4.066	1.981	6.047	0.00E+00	0.00E+00
792	Q32520FAM3C	3	NA	-1.97	NA	NA	NA	-1.97	NA	NA	NA	NA	4	NA	NA	NA	NA	4.12E+07	1.85E+07	4.57E+07	8.02E+06	NA	NA	NA	NA	-1.585	-2.642	-1.341	-2.330	-4.066	-1.974	2.092	0.00E+00	0.00E+00
793	Q32626PXDN	6	NA	0.88	NA	NA	NA	0.88	NA	NA	NA	NA	4	NA	NA	NA	NA	1.54E+08	1.39E+08	1.99E+08	1.32E+08	NA	NA	NA	NA	0.38								

1	ProteinName	# Peptides	Ctrl	Group	log2FC	CI.L	CI.R	Ave Expr	t	P.Value	adj.P.Val	B	rrN As	M.3_Co ntrol.ra w	M.2_Co ntrol.ra w	M.4_Co ntrol.ra w	M.1_Co ntrol.ra w	E4.2_Gr oup_1_r aw	E4.4_Gr oup_1_r aw	E4.3_Gr oup_1_r aw	E4.1_Gr oup_1_r aw	M.3_C ontrol. transf	M.2_C ontrol. transf	M.4_C ontrol. transf	M.1_C ontrol. transf	E4.2_Gr oup_1_tran sf	E4.4_Gr oup_1_tran sf	E4.3_Gr oup_1_tran sf	E4.1_Gr oup_1_tran sf	pseud o.Ctrl	pseud o.Group pl	pseud o.log2 FC	pseudo.P. Value	pseudo.P. Val
801	Q9HMB9IEHD1	2	NA	-2.05	NA	NA	NA	-2.05	NA	NA	NA	NA	4	NA	NA	NA	NA	3.32E+07	3.84E+07	3.39E+07	5.02E+06	NA	NA	NA	NA	-1.905	-1.507	-1.794	-3.011	-4.066	-2.054	2.012	0.00E+00	0.00E+00
802	Q9HRL6IMVFN2	7	NA	-0.24	NA	NA	NA	-0.24	NA	NA	NA	NA	4	NA	NA	NA	NA	1.64E+08	6.92E+07	1.84E+08	1.31E+07	NA	NA	NA	NA	0.476	-0.592	0.774	-1.613	-4.066	-0.239	3.827	0.00E+00	0.00E+00
803	Q9H663INE T4	2	NA	-3.10	NA	NA	NA	-3.10	NA	NA	NA	NA	4	NA	NA	NA	NA	2.30E+07	1.35E+07	8.51E+06	5.40E+06	NA	NA	NA	NA	-2.456	-3.135	-3.892	-2.904	-4.066	-3.097	0.970	0.00E+00	0.00E+00
804	Q9P2E7IPCD10	4	NA	-3.17	NA	NA	NA	-3.17	NA	NA	NA	NA	4	NA	NA	NA	NA	1.1E+07	1.27E+07	1.28E+07	6.38E+06	NA	NA	NA	NA	-3.541	-3.224	-3.271	-2.662	-4.066	-3.175	0.892	0.00E+00	0.00E+00
805	Q9P2E9IRPEP1	13	NA	-0.72	NA	NA	NA	-0.72	NA	NA	NA	NA	4	NA	NA	NA	NA	5.93E+07	5.71E+07	1.36E+08	1.67E+07	NA	NA	NA	NA	-1.041	-0.889	0.313	-1.266	-4.066	-0.721	3.345	0.00E+00	0.00E+00
806	Q9LBR2ICAT2	2	NA	-2.26	NA	NA	NA	-2.26	NA	NA	NA	NA	4	NA	NA	NA	NA	3.17E+07	2.67E+07	2.36E+07	6.43E+06	NA	NA	NA	NA	-1.978	-2.072	-2.346	-2.652	-4.066	-2.262	1.804	0.00E+00	0.00E+00
807	Q9LUJ9IGNFTG	3	NA	-1.86	NA	NA	NA	-1.86	NA	NA	NA	NA	4	NA	NA	NA	NA	3.78E+07	3.57E+07	3.96E+07	6.95E+06	NA	NA	NA	NA	-1.714	-1.619	-1.558	-2.539	-4.066	-1.858	2.209	0.00E+00	0.00E+00
808	Q9LUMY4ISNK12	2	NA	-0.72	NA	NA	NA	-0.72	NA	NA	NA	NA	4	NA	NA	NA	NA	5.77E+07	5.25E+07	6.77E+07	3.86E+07	NA	NA	NA	NA	-1.082	-1.020	-0.746	-0.046	-4.066	-0.723	3.343	0.00E+00	0.00E+00
809	Q9LNN8IEPCR	4	NA	2.35	NA	NA	NA	2.35	NA	NA	NA	NA	4	NA	NA	NA	NA	7.18E+08	5.35E+08	4.38E+08	1.63E+08	NA	NA	NA	NA	2.677	2.587	2.087	2.049	-4.066	2.350	6.416	0.00E+00	0.00E+00
810	Q9LPN3IMACF1	7	NA	-2.17	NA	NA	NA	-2.17	NA	NA	NA	NA	4	NA	NA	NA	NA	3.17E+07	1.31E+07	2.59E+07	1.62E+07	NA	NA	NA	NA	-1.976	-3.177	-2.201	-1.312	-4.066	-2.166	1.900	0.00E+00	0.00E+00
811	Q9LQ80IPA2G4	5	NA	-2.44	NA	NA	NA	-2.44	NA	NA	NA	NA	4	NA	NA	NA	NA	2.65E+07	3.86E+07	1.48E+07	5.22E+06	NA	NA	NA	NA	-2.242	-1.499	-3.050	-2.954	-4.066	-2.436	1.630	0.00E+00	0.00E+00
812	Q9Y4K0ILOXL2	10	NA	3.67	NA	NA	NA	3.67	NA	NA	NA	NA	4	NA	NA	NA	NA	1.37E+09	1.13E+09	1.20E+09	4.97E+08	NA	NA	NA	NA	3.635	3.754	3.613	3.667	-4.066	3.667	7.733	0.00E+00	0.00E+00
813	Q9Y624IJAM1	2	NA	-1.55	NA	NA	NA	-1.55	NA	NA	NA	NA	4	NA	NA	NA	NA	5.46E+07	3.68E+07	3.66E+07	1.18E+07	NA	NA	NA	NA	-1.166	-1.574	-1.691	-1.768	-4.066	-1.547	2.519	0.00E+00	0.00E+00
814	zzfY-FGCZCont000271	2	NA	4.42	NA	NA	NA	4.42	NA	NA	NA	NA	4	NA	NA	NA	NA	1.80E+09	2.05E+09	1.36E+09	1.37E+09	NA	NA	NA	NA	4.044	4.673	3.804	5.139	-4.066	4.416	8.491	0.00E+00	0.00E+00
815	zzfY-FGCZCont002221	7	NA	-1.89	NA	NA	NA	-1.89	NA	NA	NA	NA	4	NA	NA	NA	NA	4.23E+07	1.68E+07	3.73E+07	1.35E+07	NA	NA	NA	NA	-1.546	-2.792	-1.650	-1.576	-4.066	-1.891	2.175	0.00E+00	0.00E+00
816	zzfY-FGCZCont002301	6	NA	-3.08	NA	NA	NA	-3.08	NA	NA	NA	NA	4	NA	NA	NA	NA	1.51E+07	1.19E+07	1.56E+07	5.33E+06	NA	NA	NA	NA	-3.079	-3.326	-2.976	-2.924	-4.066	-3.076	0.990	0.00E+00	0.00E+00
817	zzfY-FGCZCont003331	4	NA	-3.99	NA	NA	NA	-3.99	NA	NA	NA	NA	4	NA	NA	NA	NA	5.66E+06	1.27E+07	6.85E+06	2.72E+06	NA	NA	NA	NA	-4.544	-3.233	-4.222	-3.902	-4.066	-3.975	0.091	0.00E+00	0.00E+00
818	zzfY-FGCZCont004541	2	NA	0.77	NA	NA	NA	0.77	NA	NA	NA	NA	4	NA	NA	NA	NA	1.59E+08	1.27E+08	1.86E+08	1.15E+08	NA	NA	NA	NA	0.432	0.349	0.785	1.534	-4.066	0.775	4.641	0.00E+00	0.00E+00

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