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Draft genome sequences of two wide-host-range phages of *Listeria monocytogenes* from food processing environments in the United States

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ABSTRACT *Listeria monocytogenes* is notorious for persistence in food facilities. Phages can significantly impact the ecology of *Listeria*, but there is a dearth of genome sequence data for *Listeria* phages from food processing ecosystems. We report the genome sequences of two *Listeria* phages from turkey processing facilities in the USA.

KEYWORDS *Listeria*, bacteriophages, food-borne pathogens, genomes, intracellular pathogens

Listeria monocytogenes is notorious for persistence in food processing environments (FPEs) and subsequent food contamination leading to severe disease (listeriosis) (1–3). Virulent, wide-host-range (WHR) phages can significantly impact the ecology and population dynamics of *L. monocytogenes* in FPEs, but most investigated WHR phages have been from farms and sewage (4–6). We previously isolated WHR phages from two turkey processing plants in the USA and showed that *L. monocytogenes* of serotypes 1/2a and 1/2b were frequently resistant to these phages (7). Here, we report the draft genome sequences of two of these phages, 20422-1 and 805405-1 (Table 1). Both phages were members of family *Herelleviridae*, genus *Pecentumvirus*. They had identical size (134 kb) and 3,347 bp direct terminal redundancies, and almost identical sequences (100% coverage, 99.96% nt identity) highly similar to the WHR phages A511 (GCF_000871125.1) and P100 (DQ004855.1) (97% coverage, approx. 98.4% nt identity) (genus *Pecentumvirus*) isolated in 1990 and 1997 from sewage in Germany (8, 9). The high genomic similarity of 20422-1 and 805405-1 raises the possibility that they represent different isolates of a WHR listeria phage disseminated in different turkey processing plants in the USA.

Bacteriophages were isolated as described (7) and propagated using the soft-agar overlay method with 1/2 brain-heart infusion (Biolife, Milan, Italy) as bottom agar (1% agar) and LC as top agar (0.4% lysogeny broth agar with 10mM CaCl₂, 2mM MgSO₄, and 10g/L glucose). Briefly, 200 μL log-phase host bacteria (*Listeria ivanovii* WSLC 3009, OD [Optical Density]₆₀₀ = 0.5) were mixed with 10 μL phage dilution adjusted for semi-confluent lysis in 5 mL liquid top agar at 47°C and overlaid on the bottom agar. Plates were incubated at 30°C. Progeny virions were extracted from 40 semi-confluent plates using 5 mL SM buffer/plate (100mM NaCl, 8mM MgSO₄, 50mM Tris, pH 7.4) and filter-sterilized (0.2 μm) to obtain lysates. Lysates were digested with DNase I (10 μg/mL) and RNase A (1 U/10 mL, 37°C, 30 min), and phages concentrated by polyethylene glycol (PEG) precipitation (7% PEG 8000 and 1 M NaCl) at 4°C for 16 hours. Phages were resuspended in 5 mL SM buffer, purified by CsCl isopycnic centrifugation (10), and dialyzed twice against 1,000× excess SM buffer. CsCl-purified phages were digested with proteinase K (200 μg/mL, 55°C, 30 min, in SM buffer with 10 mM EDTA, pH 8.0), and phage DNA was purified with two additional purification steps using phenol:chloroform:isoamyl alcohol (25:24:1) followed by DNA precipitation in ethanol as described (11). Library preparation

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TABLE 1 Bacteriophages investigated in this study

Characteristics	20422-1	805405-1
Source	Food processing environment, conveyor belt, USA, Plant A	Food processing environment, floor drain, USA, Plant B
Isolation date	June 2004	May 2005
Family	<i>Herelleviridae</i>	<i>Herelleviridae</i>
Genus	<i>Pecentumvirus</i>	<i>Pecentumvirus</i>
Species	nA ^a	nA ^a
No. of contigs	1	1
Total length (bp)	134,755	134,754
N50 (bp)	134,755	134,754
Coverage (X)	6,742	10,832
No. of reads	6,056,756	9,731,674
GC content	35.9%	36.0%
Sequencing method	Illumina NovaSeq6000	Illumina NovaSeq6000
Read quality control tools	CLC Genomics Workbench version 2.0	CLC Genomics Workbench version 2.0
GenBank accession	PP101685.1	PP101686.1
BioSample accession	SAMN38501085	SAMN38501086
Sequence read archive accession	SRR26988306	SRR26988305

^anA; Not available.

and Illumina sequencing (2 × 150 bp, 5M reads) of purified DNA was performed by Eurofins Genomics Europe Sequencing GmbH (Constance, Germany). Sequence analysis and quality control utilized CLC Genomics version 2.0 (QIAGEN Bioinformatics). Adapter trimming was not required. Single contigs were obtained for both genomes by *de novo* assembly using the CLC Genomics Workbench. Terminal redundancies were identified by mapping Illumina reads to the unit genome, revealing a characteristic duplication of sequencing coverage over a 3,347 nt region. Coding DNA sequence identification and annotation was performed using the RAST server (12). Default parameters were used for all software unless otherwise specified.

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DATA AVAILABILITY

This Whole Genome Shotgun project has been deposited in NCBI under BioProject [PRJNA1046509](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1046509). The version described in this paper is the first version.

REFERENCES

- Painter J, Slutsker L. 2007. Listeriosis in humans, p 85–109. In ET Ryser, EH Marth (ed), *Listeria*, listeriosis and food safety, 3rd ed. CRC Press, Boca Raton, FL.
- Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson M-A, Roy SL, Jones JL, Griffin PM. 2011. Foodborne illness acquired in the United States—major pathogens. *Emerg Infect Dis* 17:7–15. <https://doi.org/10.3201/eid1701.p11101>
- de Noordhout CM, Devleeschauwer B, Angulo FJ, Verbeke G, Haagsma J, Kirk M, Havelaar A, Speybroeck N. 2014. The global burden of Listeriosis: a systematic review and meta-analysis. *Lancet Infect Dis* 14:1073–1082. [https://doi.org/10.1016/S1473-3099\(14\)70870-9](https://doi.org/10.1016/S1473-3099(14)70870-9)
- Vongkamjan K, Switt AM, den Bakker HC, Fortes ED, Wiedmann M. 2012. Silage collected on dairy farms harbors an abundance of listeriophages with considerable host range and genome size diversity. *Appl Environ Microbiol* 78:8666–8675. <https://doi.org/10.1128/AEM.01859-12>
- Klumpp J, Loessner MJ. 2013. *Listeria* phages: genomes, evolution, and application. *Bacteriophage* 3:e26861. <https://doi.org/10.4161/bact.26861>
- Denes T, Vongkamjan K, Ackermann H-W, Moreno Switt AI, Wiedmann M, den Bakker HC. 2014. Comparative genomic and morphological analyses of *Listeria* phages isolated from farm environments. *Appl Environ Microbiol* 80:4616–4625. <https://doi.org/10.1128/AEM.00720-14>
- Kim JW, Siletzky RM, Kathariou S. 2008. Host ranges of *Listeria*-specific bacteriophages from the turkey processing plant environment in the United States. *Appl Environ Microbiol* 74:6623–6630. <https://doi.org/10.1128/AEM.01282-08>
- Carlton RM, Noordman WH, Biswas B, de Meester ED, Loessner MJ. 2005. Bacteriophage P100 for control of *Listeria monocytogenes* in foods: genome sequence, bioinformatic analyses, oral toxicity study, and application. *Regul Toxicol Pharmacol* 43:301–312. <https://doi.org/10.1016/j.yrtph.2005.08.005>
- Habann M, Leiman PG, Vandersteegen K, Van den Bossche A, Lavigne R, Shneider MM, Biemann R, Eugster MR, Loessner MJ, Klumpp J. 2014. *Listeria* Phage A511, a model for the contractile tail Machineries of Spo1-related Bacteriophages. *Mol Microbiol* 92:84–99. <https://doi.org/10.1111/mmi.12539>
- Sambrook J, Russell DW. 2001. *Molecular cloning: a laboratory manual*. 3rd ed. Vol. 1. Cold Spring Harbor Laboratory Press, New York.
- Green MR, Sambrook J. 2017. Isolation of high-molecular-weight DNA using organic solvents. *Cold Spring Harb Protoc* 2017:db. <https://doi.org/10.1101/pdb.prot093450>
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, et al. 2008. The RAST Server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>